

GENETIKA ÉS POPULÁCIÓGENETIKA

Genetikai polimorfizmusok



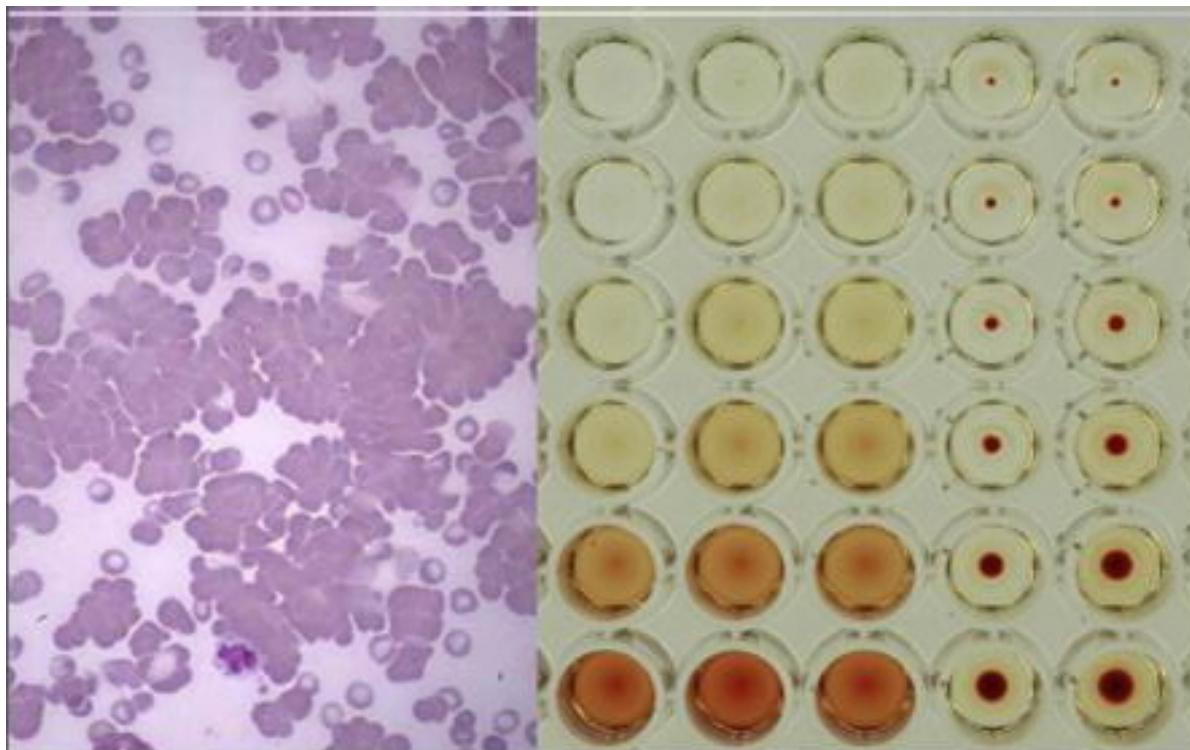
ELTE TTK Genetikai Tanszék

Recesszív gén hőmérséklet szenzitív expressziója

- TYR gén ► tirozináz enzim ► melanin szintézis (sötét szín)
 - Defektív tirozináz ► funkcióvesztés normál testhőmérsékleten
 - sötét színárnyalat csak az alacsonyabb testhőmérsékletű helyeken



Első genetikai marker: ABO vércsoport-rendszer



FELIX BERNSTEIN (1933)

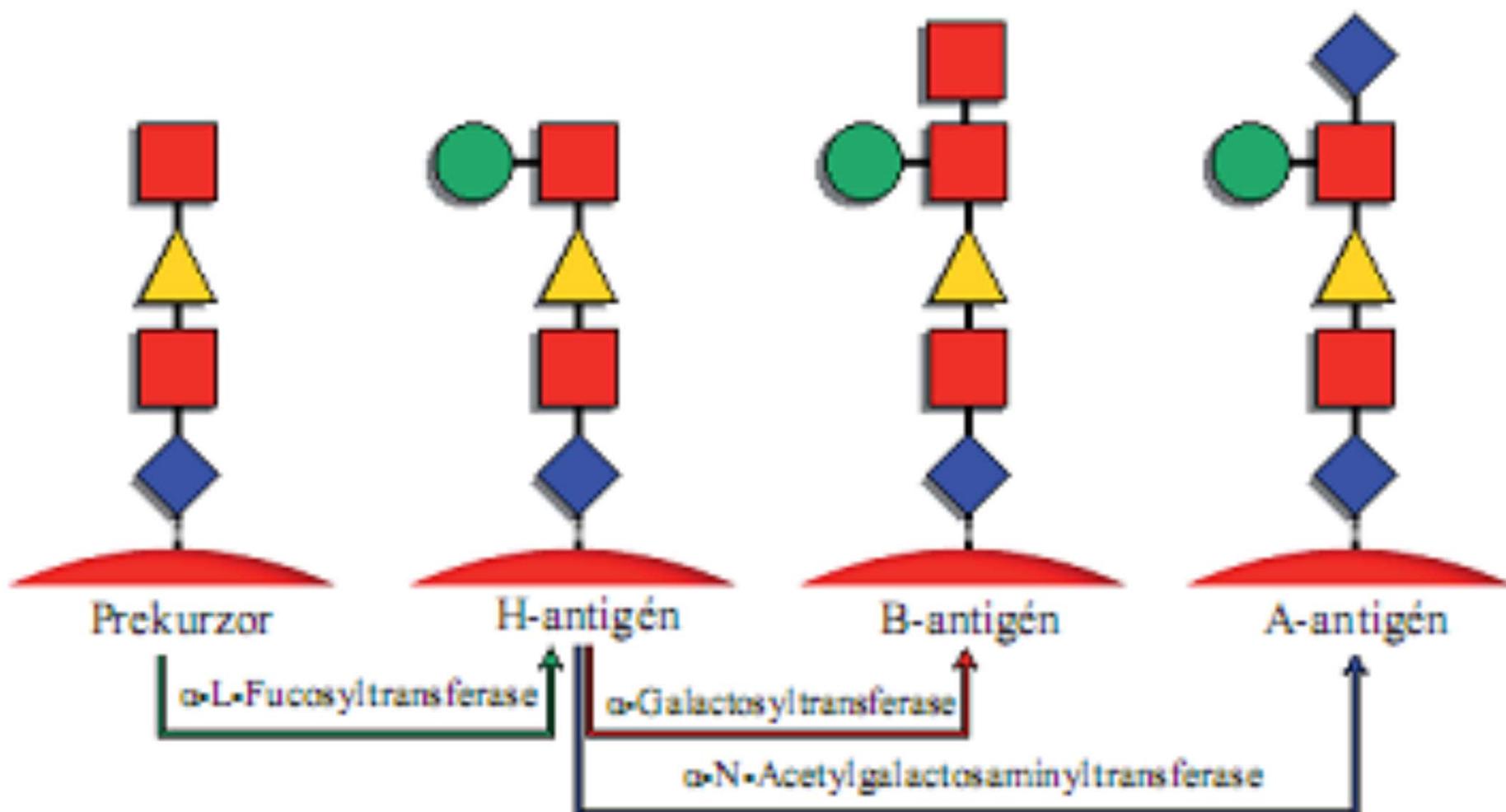
Two hypotheses of blood group inheritance

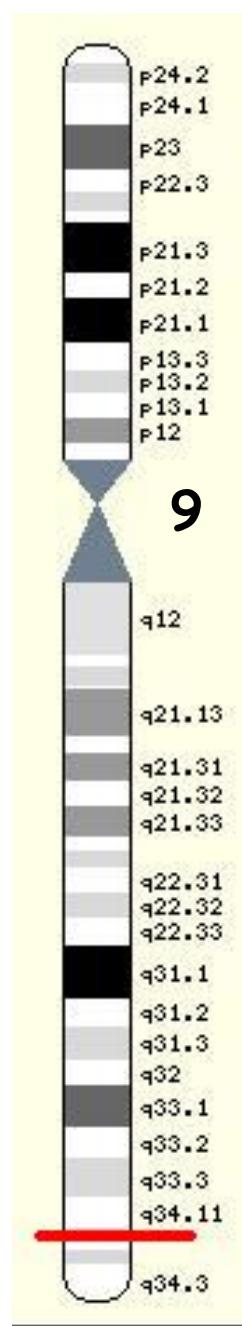
Group	VON DUNGERN and HIRZFELD		BERNSTEIN		Observed proportion
	Genotype	Expected proportion	Genotype	Expected proportion	
O	$aa\ bb$	$p_a^2\ p_b^2$	OO	p_O^2	0.294
A	$A-\ bb$	$(1 - p_a^2)p_b^2$	AA, OA	$p_A^2 + 2p_Op_A$	0.422
B	$aa\ B-$	$p_a^2(1 - p_b^2)$	BB, OB	$p_B^2 + 2p_Op_B$	0.206
AB	$A-\ B-$	$(1 - p_a^2)(1 - p_b^2)$	AB	$2p_Ap_B$	0.078
Total		1		1	1.000

The expected proportions assume Hardy-Weinberg ratios and linkage equilibrium. The observed proportions are from 502 Japanese (BERNSTEIN 1925).

- L-Fucose
- D-Galactose
- ◆ N-Acetyl galactosamine
- ▲ N-Acetyl glucosamine

AB0 antigének kialakulása





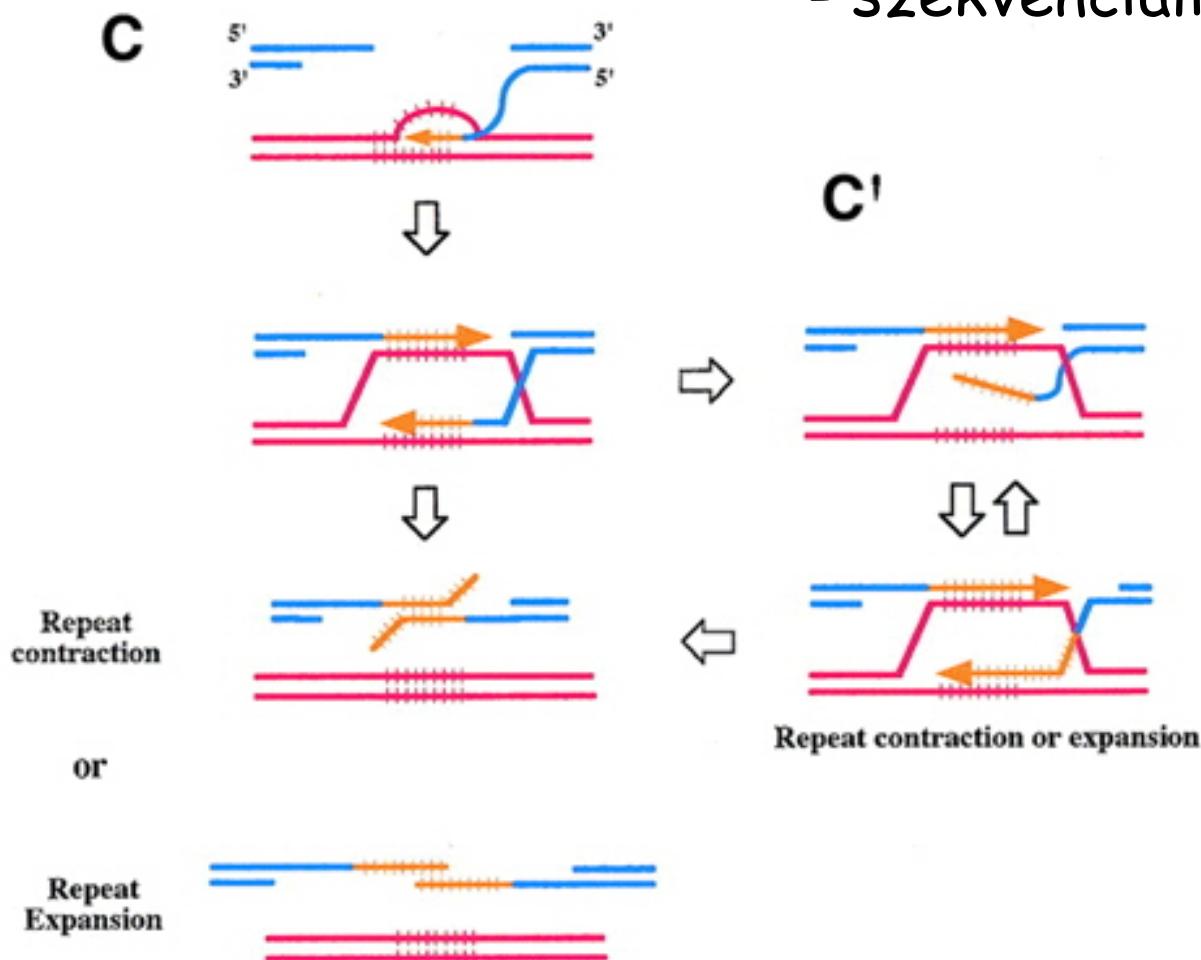
Various Alleles at the ABO Locus

Exon Number	6										7									
Nucleotide Position	2	2	4	5	6	6	6	7	7	7	7	8	8	8	8	8	9	1	1	
	6	9	6	2	4	5	8	0	7	9	0	0	2	7	1	3	0	0	0	
	1	7	7	6	6	7	1	3	1	6	2	3	9	9	1	0	5	6	4	
A alleles																				
A101	G	A	C	C	T	C	G	G	C	C	G	G	G	G	G	G	C	C	C	
A102	*	*	T	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	
A201	*	*	T	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	
A301	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	A	*	*	
Ax01	*	*	*	*	A	*	*	*	*	*	*	*	*	*	*	*	*	*	*	
cis-AB01	*	*	T	*	*	*	*	*	*	*	*	*	C	*	*	*	*	*	*	
B alleles																				
B101	*	G	*	G	*	T	*	A	*	A	*	C	*	*	A	*	*	*	*	
B301	*	G	*	G	*	T	*	A	*	A	*	C	*	*	A	T	*	*	*	
B(A)01	*	G	*	G	*	*	*	*	*	A	*	C	*	*	A	*	*	*	*	
O alleles																				
O01	△	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*	*
O02	△	G	*	*	A	*	A	*	T	*	*	A	*	*	*	*	*	*	*	
O03	*	G	*	G	*	*	*	*	*	A	*	*	*	*	*	*	*	*	*	
Possible Amino Acid Change	Frameshift	No change	P156L	R176G	F216I	No change	G235S	No change	L266M	G268R	G268A	V277M	D291N	No change	R352W	Frameshift				

A humán genom projekt eredményei

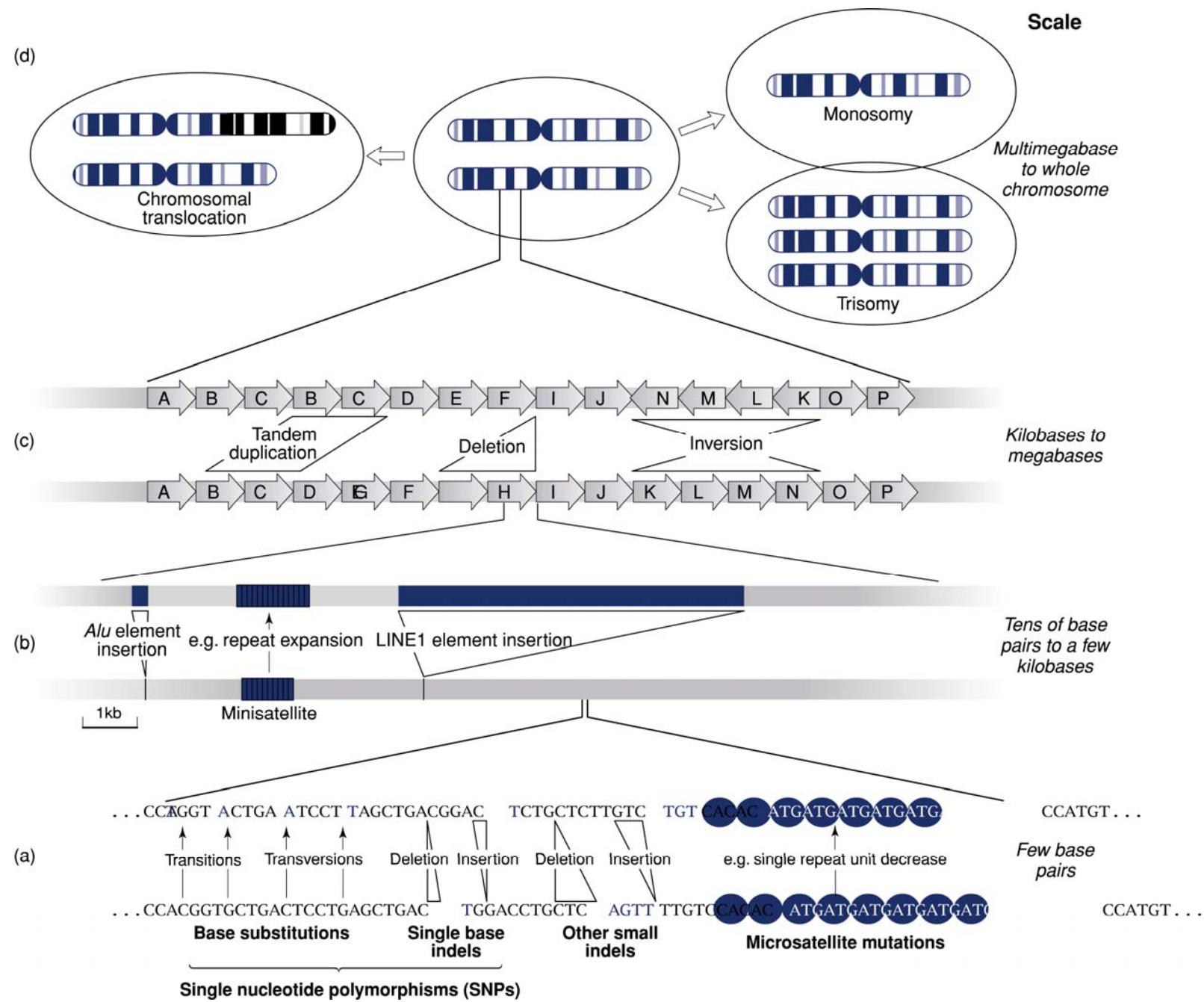
- Előzetes publikálás 2001-ben (Science, Nature)
- Az eddig leírt legnagyobb méretű teljes genom
- Az eukarióta genomra jellemző szerkezeti és szerveződési tulajdonságok (modellszervezetek)
- Meglepően alacsony a fehérje kódoló gének száma (~22000)
- RNS gének növekvő száma (tRNA, snRNA, miRNA, stb.)
- Orphan gének: nincs homológja, humán gének ~ 1/3-a
- Protein kódoló szekvenciák aránya: < 1 %
- Repetitív szekvenciák túlnyomó aránya!

REKOMBINÁCIÓ



Polimorfizmus hajtóereje:

- pontmutáció
- szekvenciális átrendeződés

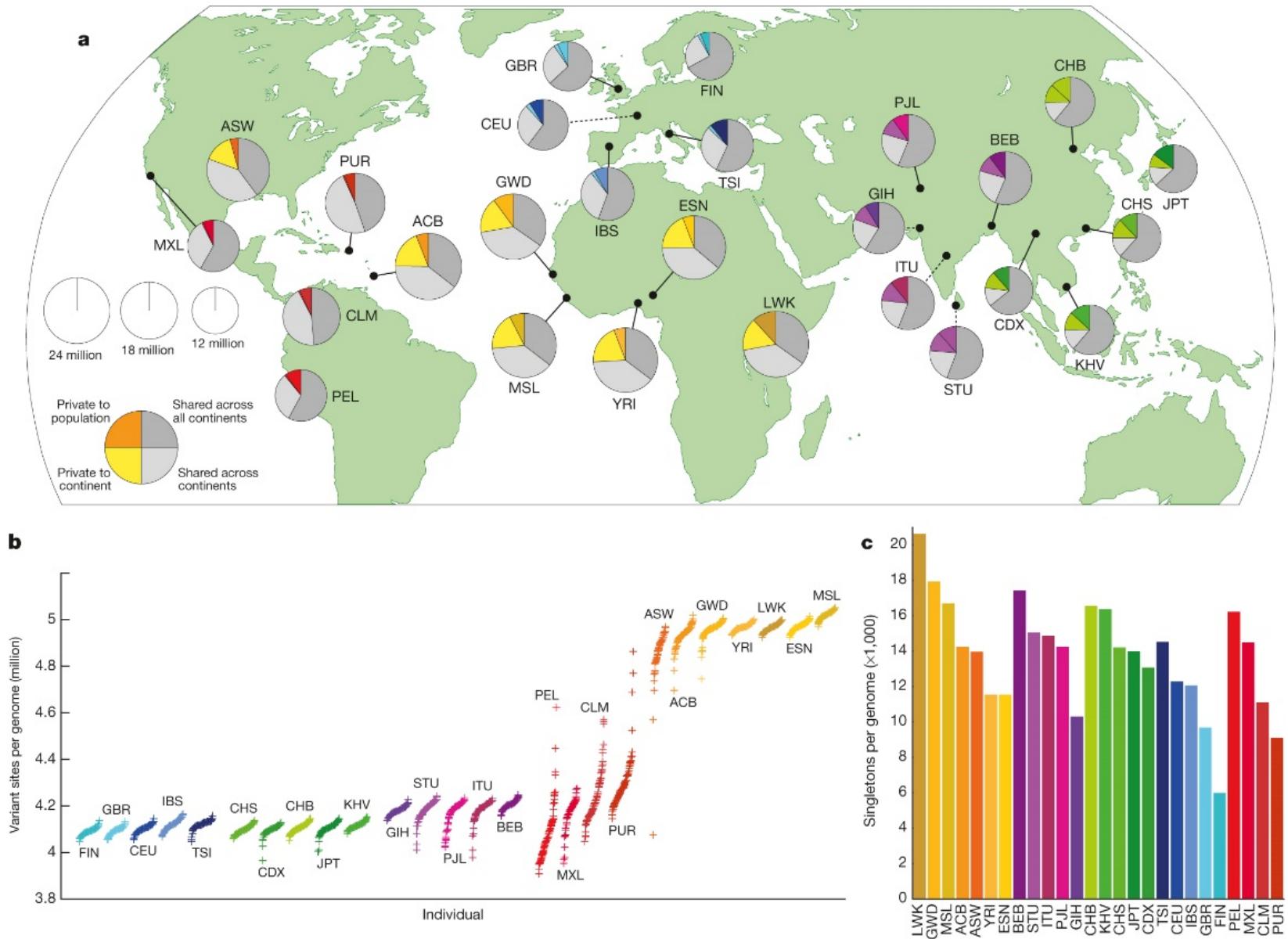


A global reference for human genetic variation

The 1000 Genomes Project Consortium*

The 1000 Genomes Project set out to provide a comprehensive description of common human genetic variation by applying whole-genome sequencing to a diverse set of individuals from multiple populations. Here we report completion of the project, having reconstructed the genomes of 2,504 individuals from 26 populations using a combination of low-coverage whole-genome sequencing, deep exome sequencing, and dense microarray genotyping. We characterized a broad spectrum of genetic variation, in total over 88 million variants (84.7 million single nucleotide polymorphisms (SNPs), 3.6 million short insertions/deletions (indels), and 60,000 structural variants), all phased onto high-quality haplotypes. This resource includes >99% of SNP variants with a frequency of >1% for a variety of ancestries. We describe the distribution of genetic variation across the global sample, and discuss the implications for common disease studies.

Population sampling



nature

A global reference for human genetic variation

The 1000 Genomes Project Consortium*

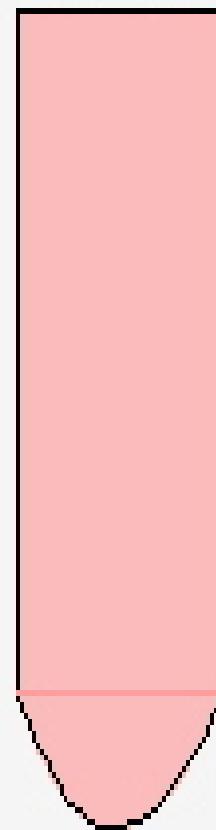
Table 1 | Median autosomal variant sites per genome

	AFR		AMR		EAS		EUR		SAS	
Samples	661		347		504		503		489	
Mean coverage	8.2		7.6		7.7		7.4		8.0	
	Var. sites	Singletons								
SNPs	4.31M	14.5k	3.64M	12.0k	3.55M	14.8k	3.53M	11.4k	3.60M	14.4k
Indels	625k	-	557k	-	546k	-	546k	-	556k	-
Large deletions	1.1k	5	949	5	940	7	939	5	947	5
CNVs	170	1	153	1	158	1	157	1	165	1
MEI (Alu)	1.03k	0	845	0	899	1	919	0	889	0
MEI (L1)	138	0	118	0	130	0	123	0	123	0
MEI (SVA)	52	0	44	0	56	0	53	0	44	0
MEI (MT)	5	0	5	0	4	0	4	0	4	0
Inversions	12	0	9	0	10	0	9	0	11	0
Nonsynon	12.2k	139	10.4k	121	10.2k	144	10.2k	116	10.3k	144
Synon	13.8k	78	11.4k	67	11.2k	79	11.2k	59	11.4k	78
Intron	2.06M	7.33k	1.72M	6.12k	1.68M	7.39k	1.68M	5.68k	1.72M	7.20k
UTR	37.2k	168	30.8k	136	30.0k	169	30.0k	129	30.7k	168
Promoter	102k	430	84.3k	332	81.6k	425	82.2k	336	84.0k	430
Insulator	70.9k	248	59.0k	199	57.7k	252	57.7k	189	59.1k	243
Enhancer	354k	1.32k	295k	1.05k	289k	1.34k	288k	1.02k	295k	1.31k
TFBSs	927	4	759	3	748	4	749	3	765	3
Filtered LoF	182	4	152	3	153	4	149	3	151	3
HGMD-DM	20	0	18	0	16	1	18	2	16	0
GWAS	2.00k	0	2.07k	0	1.99k	0	2.08k	0	2.06k	0
ClinVar	28	0	30	1	24	0	29	1	27	1

See Supplementary Table 1 for continental population groupings. CNVs, copy-number variants; HGMD-DM, Human Gene Mutation Database disease mutations; k, thousand; LoF, loss-of-function; M, million; MEI, mobile element insertions.

Satellite DNA

uniform



centrifugation



gradient

satellite bands

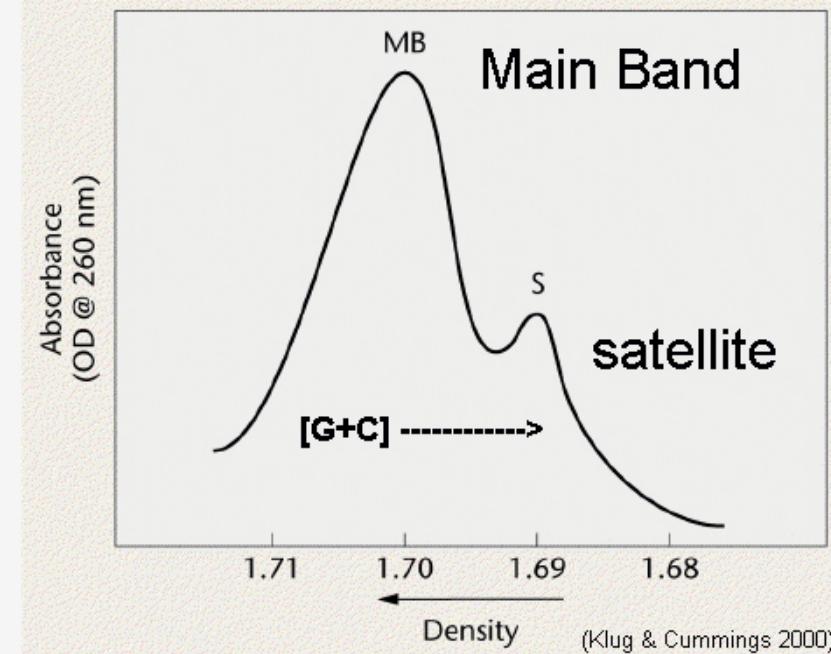
main band

Optical density

Main band DNA
1.700
1.692 $(ACAAACT)_n$

1.688 $(ATAAACT)_n$
1.671 $(ACAAATT)_n$

← Bouyant density In CsCl

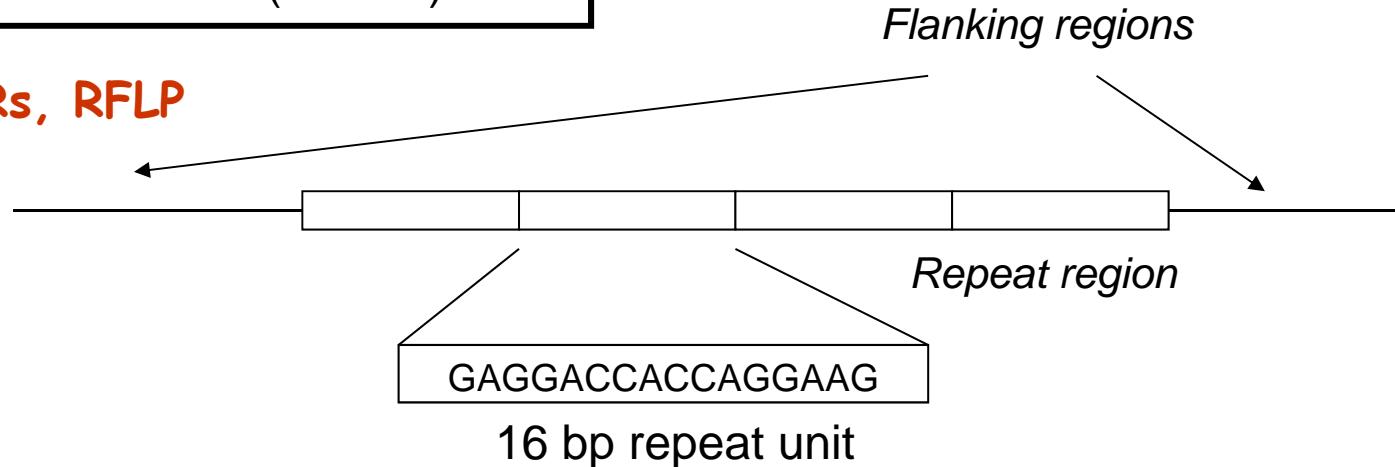


1.71 1.70 1.69 1.68

← Density (Klug & Cummings 2000)

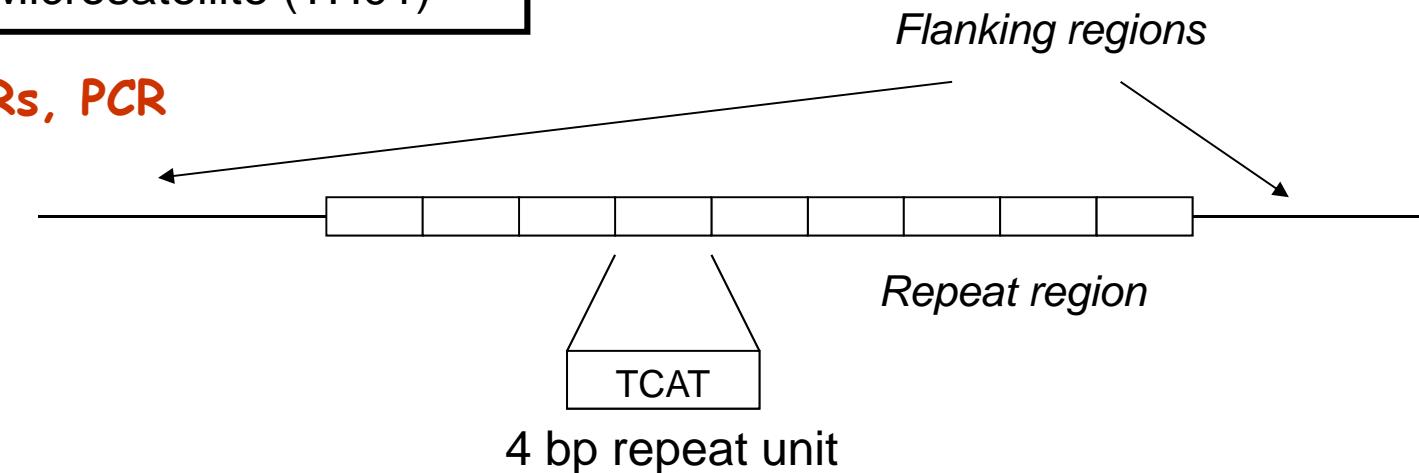
Minisatellite (D1S80)

VNTRs, RFLP



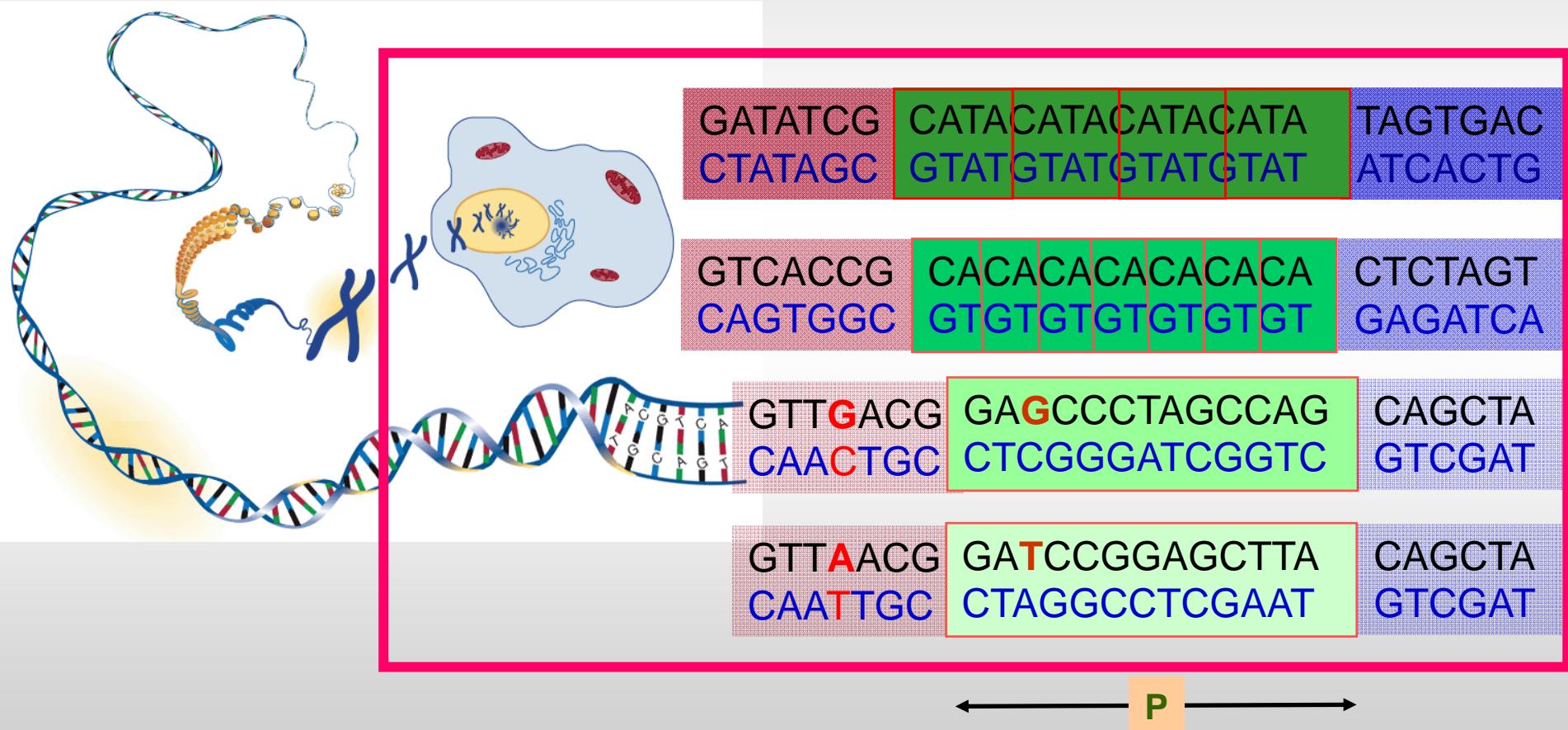
Microsatellite (TH01)

STRs, PCR



DNS polimorfizmusok

polimorfizmus - célrégió a PCR vizsgálatokban

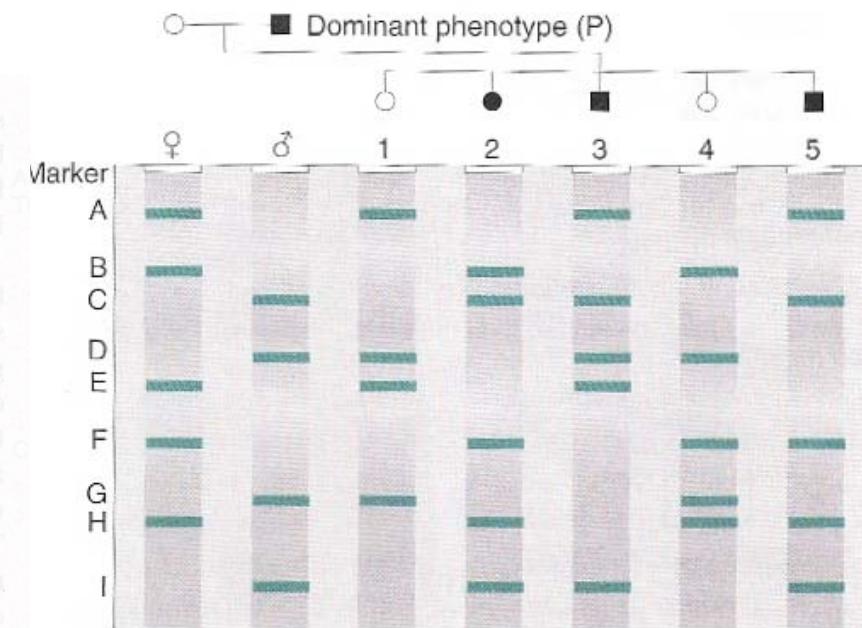


P : polimorf célszakasz (marker, lokusz, allél)



VNTR assay markerek: RFLP analitika

1985 - Sir Alec Jeffreys

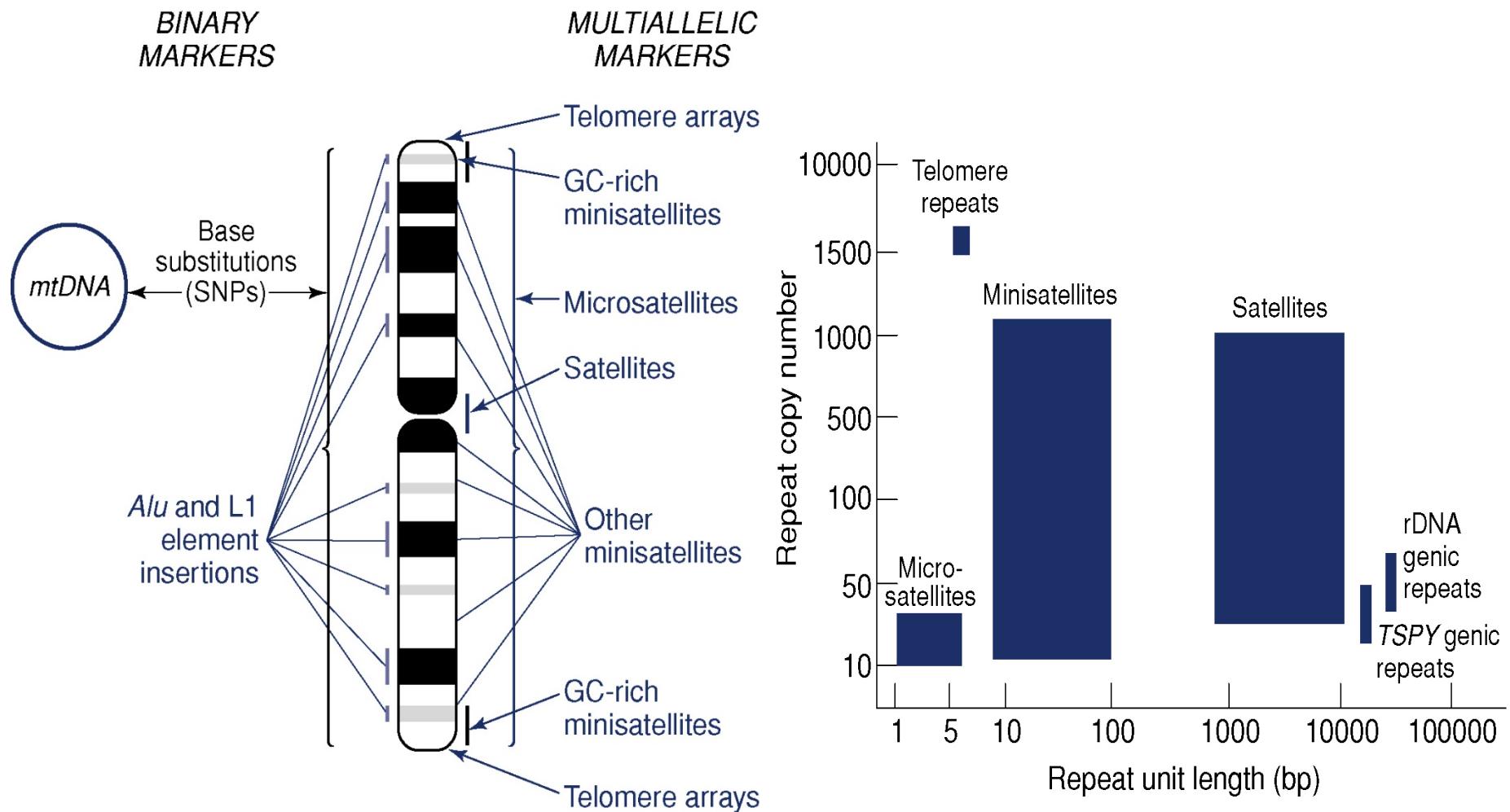


ANALYSIS EXAMPLES

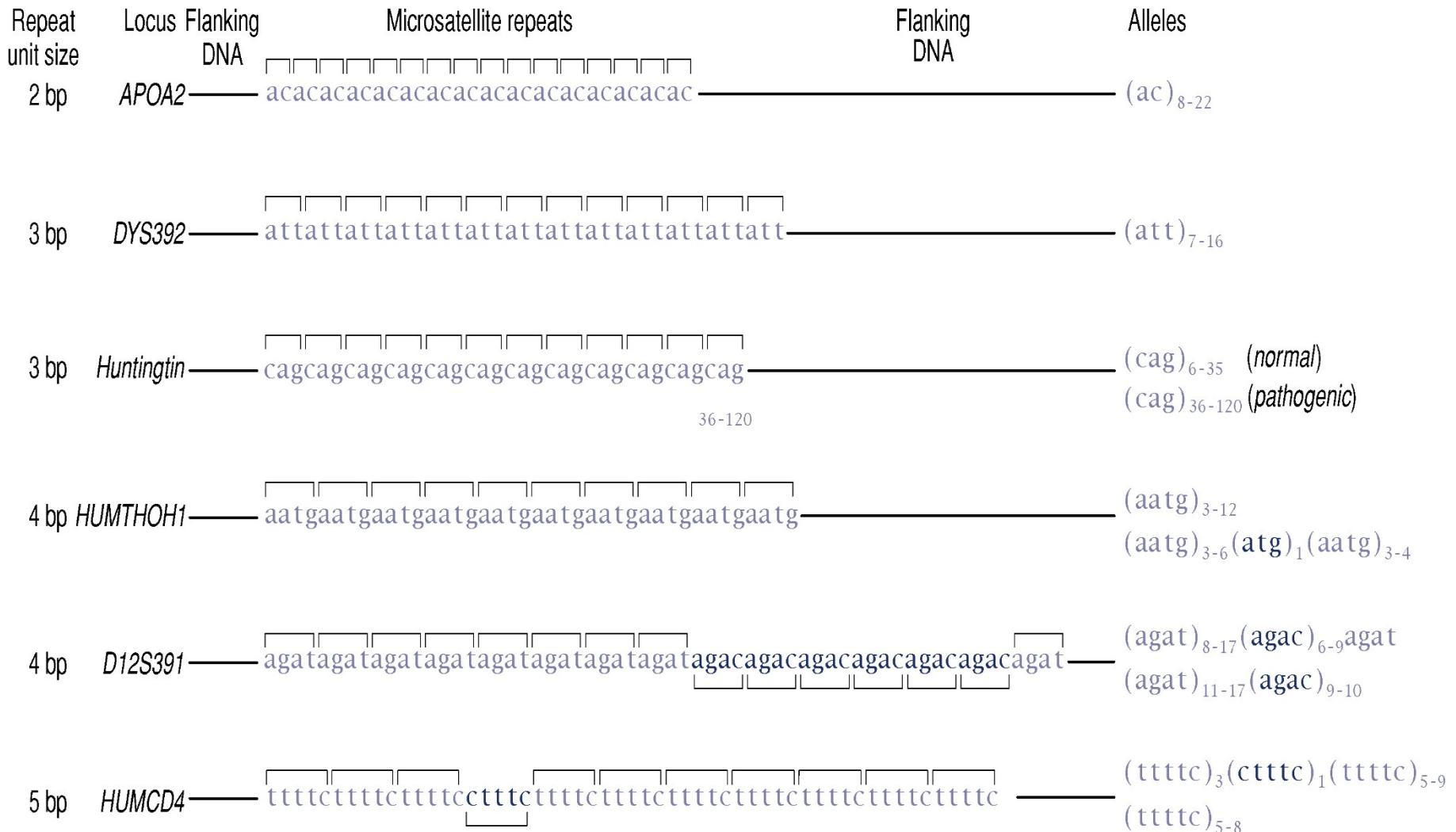
- F and H Always inherited together — linked?
- A and B In progeny, always *either A or B* — "allelic"?
- A and D Four combinations; A and D, A, D, or neither — unlinked?
- F, H, and E Always *either F and H or E* — closely linked in trans?
- Allele P Possibly linked to I and C.

Genetikai térképezés

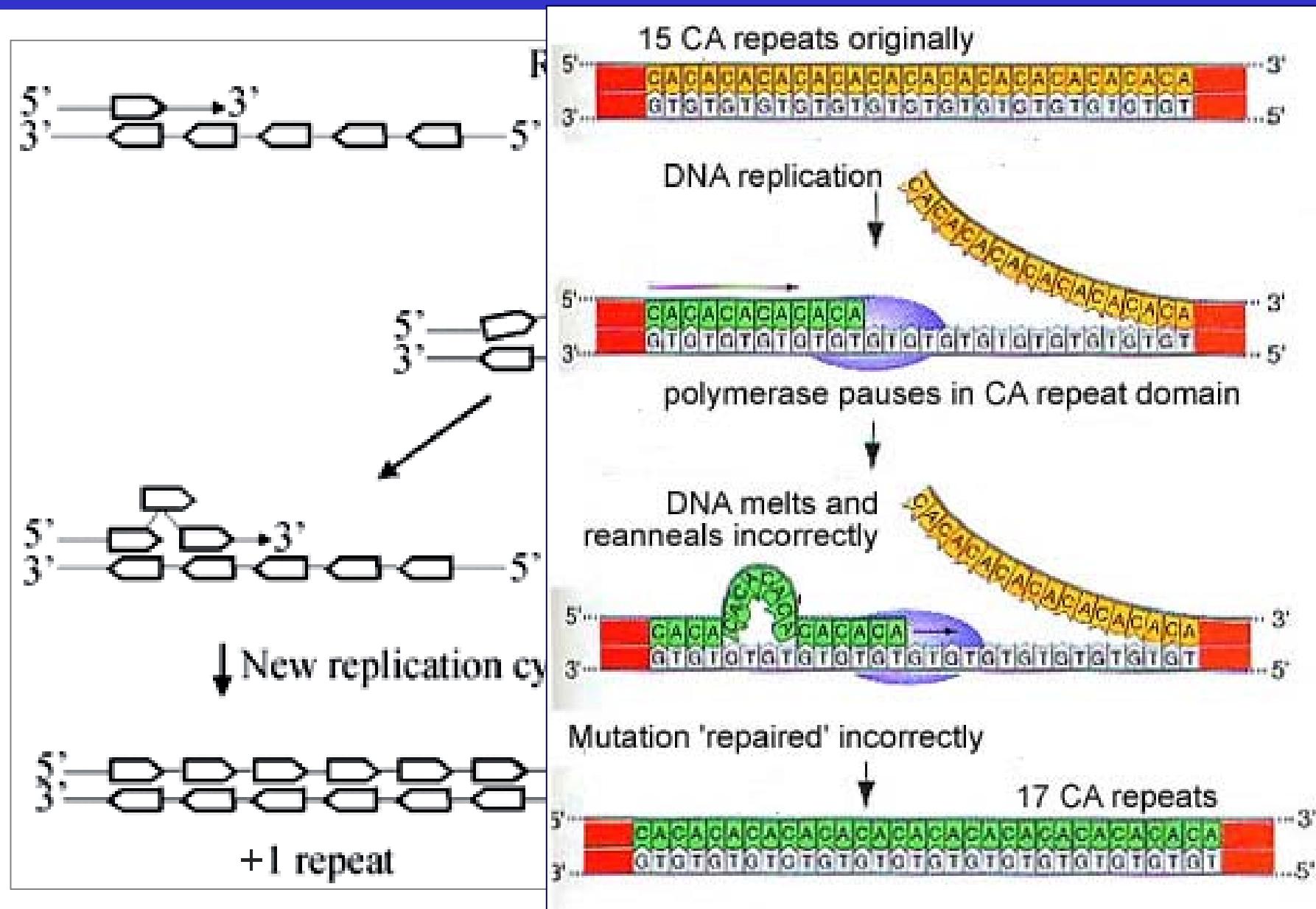
Polimorf markerek eloszlása a genomban



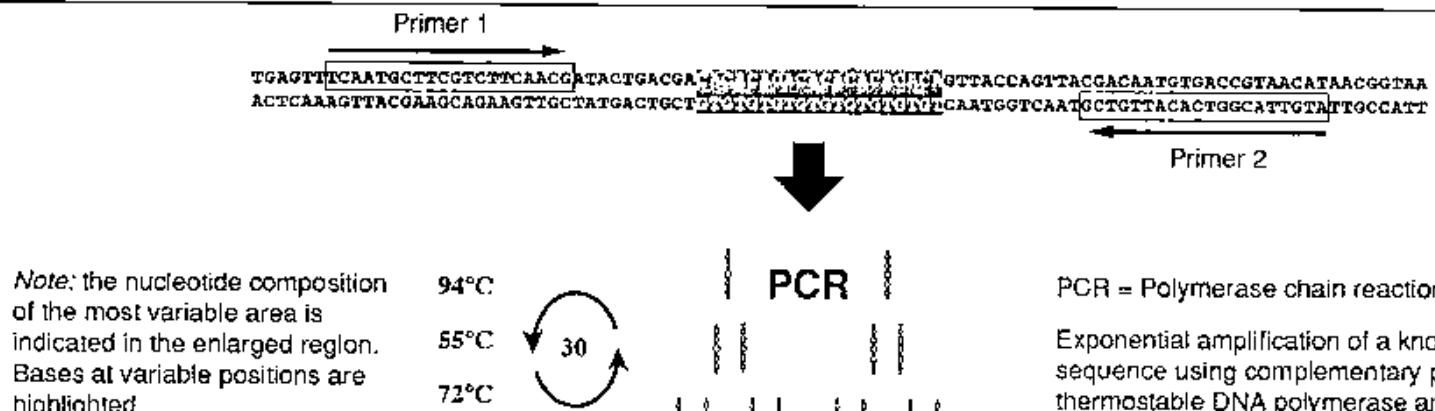
Mikroszatellit struktúra



Mikroszatellita evolúció

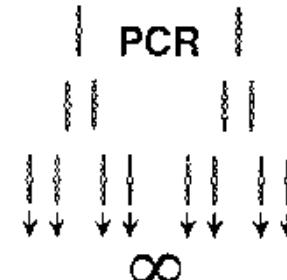


Mikroszatellita genotipizálás PCR-rel



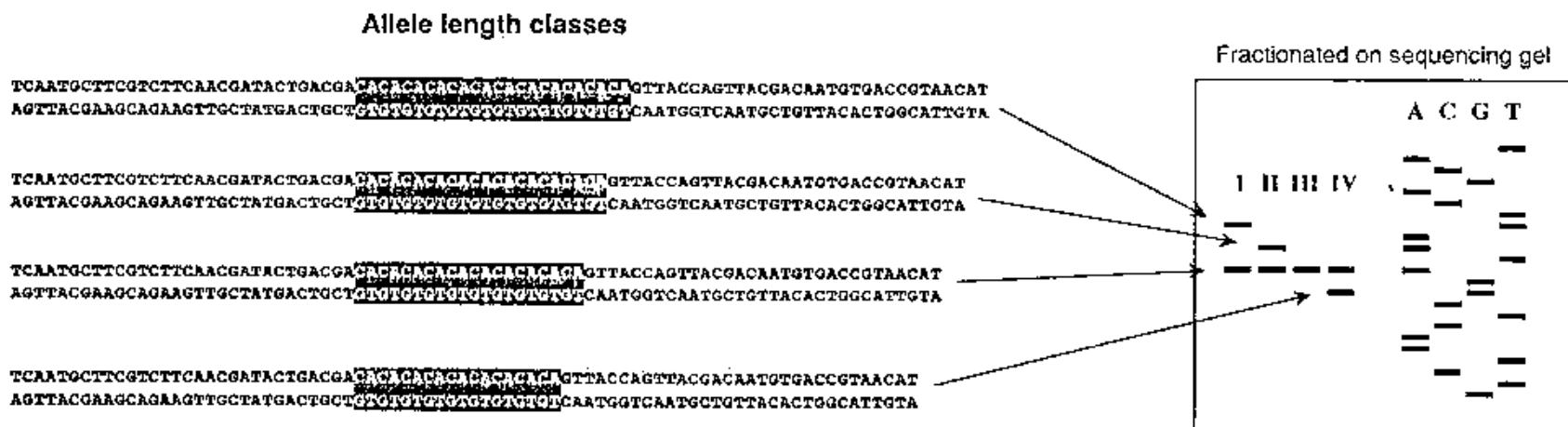
Note: the nucleotide composition of the most variable area is indicated in the enlarged region. Bases at variable positions are highlighted.

94°C
55°C
72°C



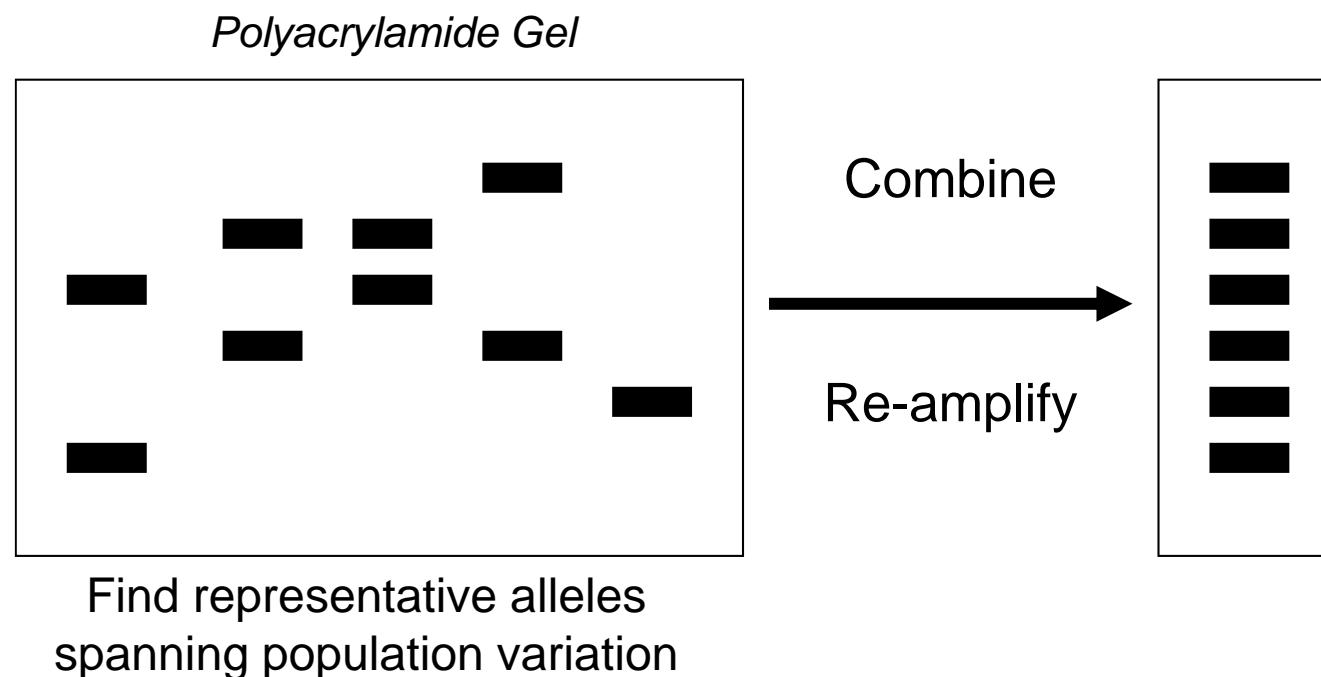
PCR = Polymerase chain reaction

Exponential amplification of a known DNA sequence using complementary primers, a thermostable DNA polymerase and a temperature cycling protocol

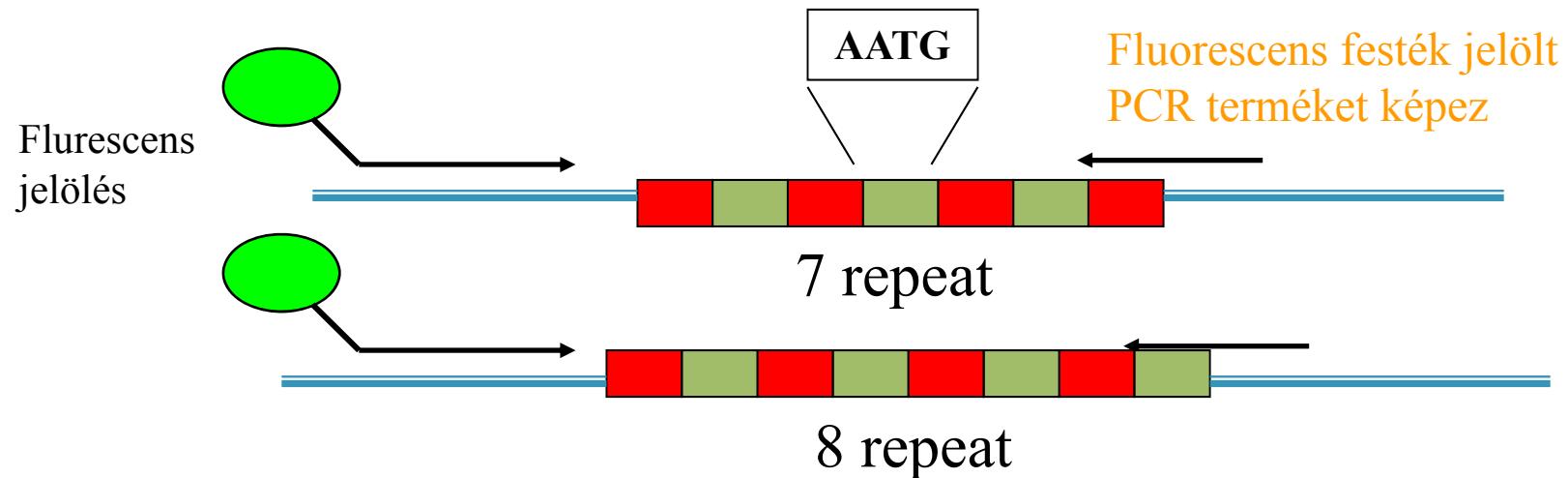


STR allélek genotipizálása: multiallélek

Separate PCR products from various samples amplified with primers targeted to a particular STR locus



Mikroszatellita - STR - markerek (Short Tandem Repeat)



a repeat régió variabilis az egyes minták között míg a flanking régió ahol a PCR primerek tapadnak állandó

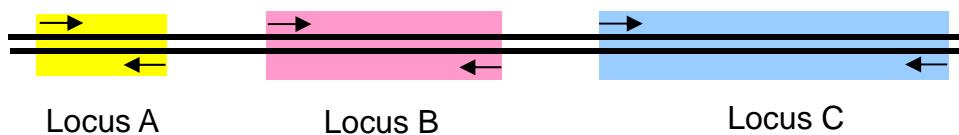
Homozigóta = a két allél megegyezik

Heterozigóta = allélek különbözők és egymástól elválaszthatók

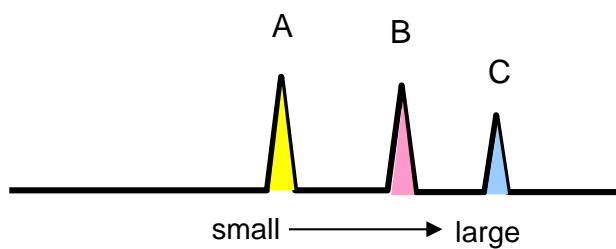
A primerek kötési helye meghatározza a PCR termék méretét!

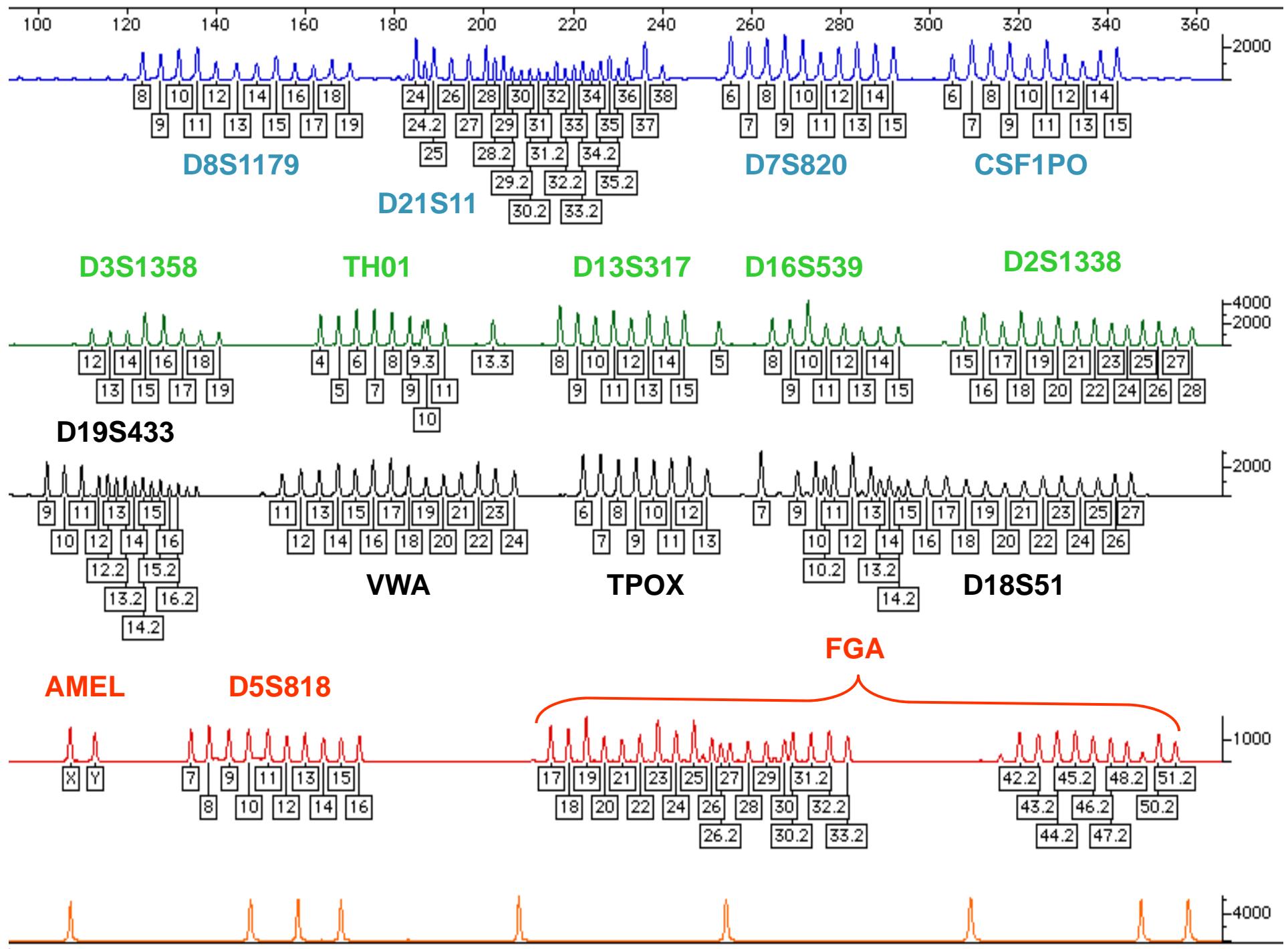
Multiplex - PCR

(A) Három lokusz szimultán amplifikálása egy reakcióban



(B) PCR termékek elválasztása frgments méret alapján

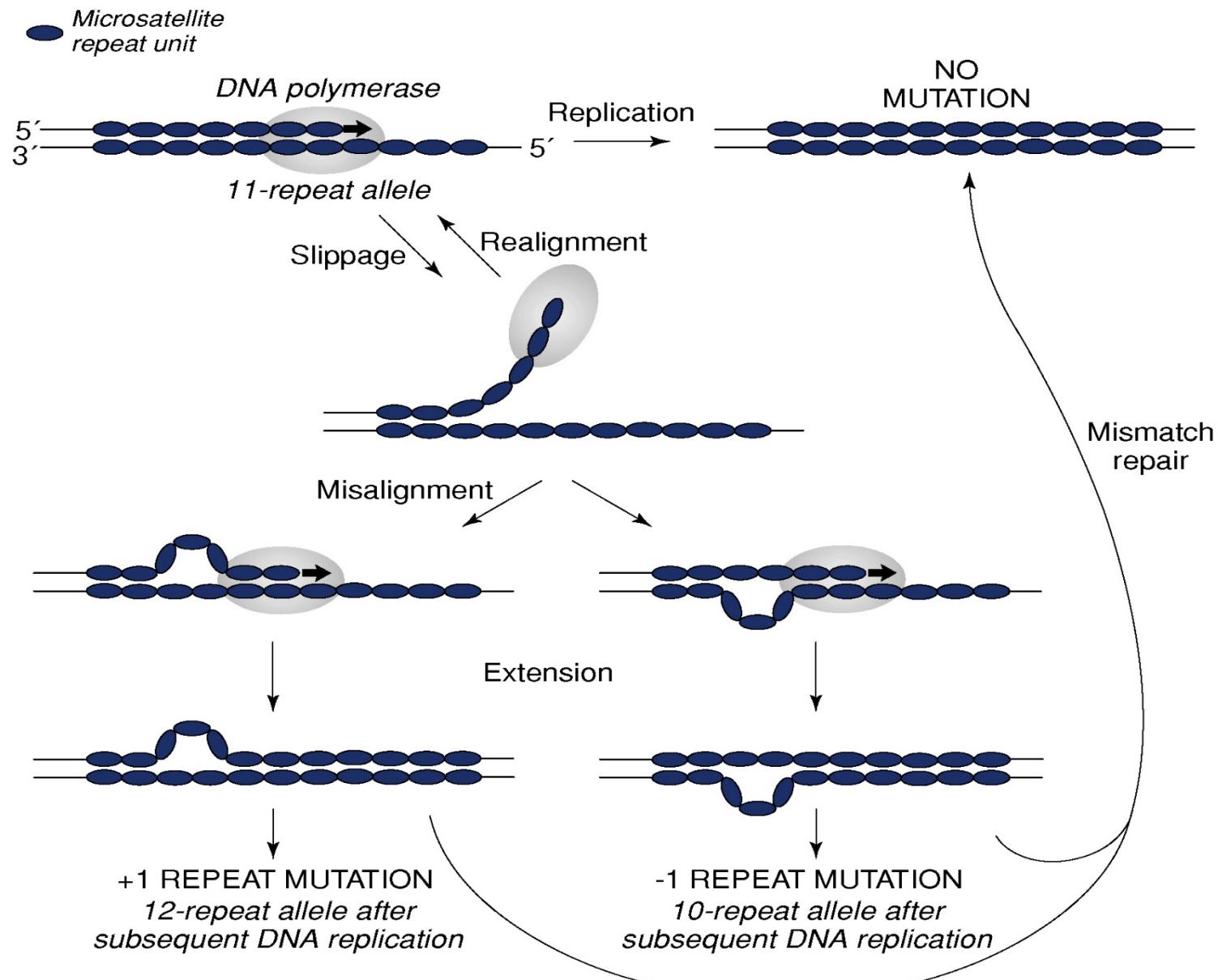




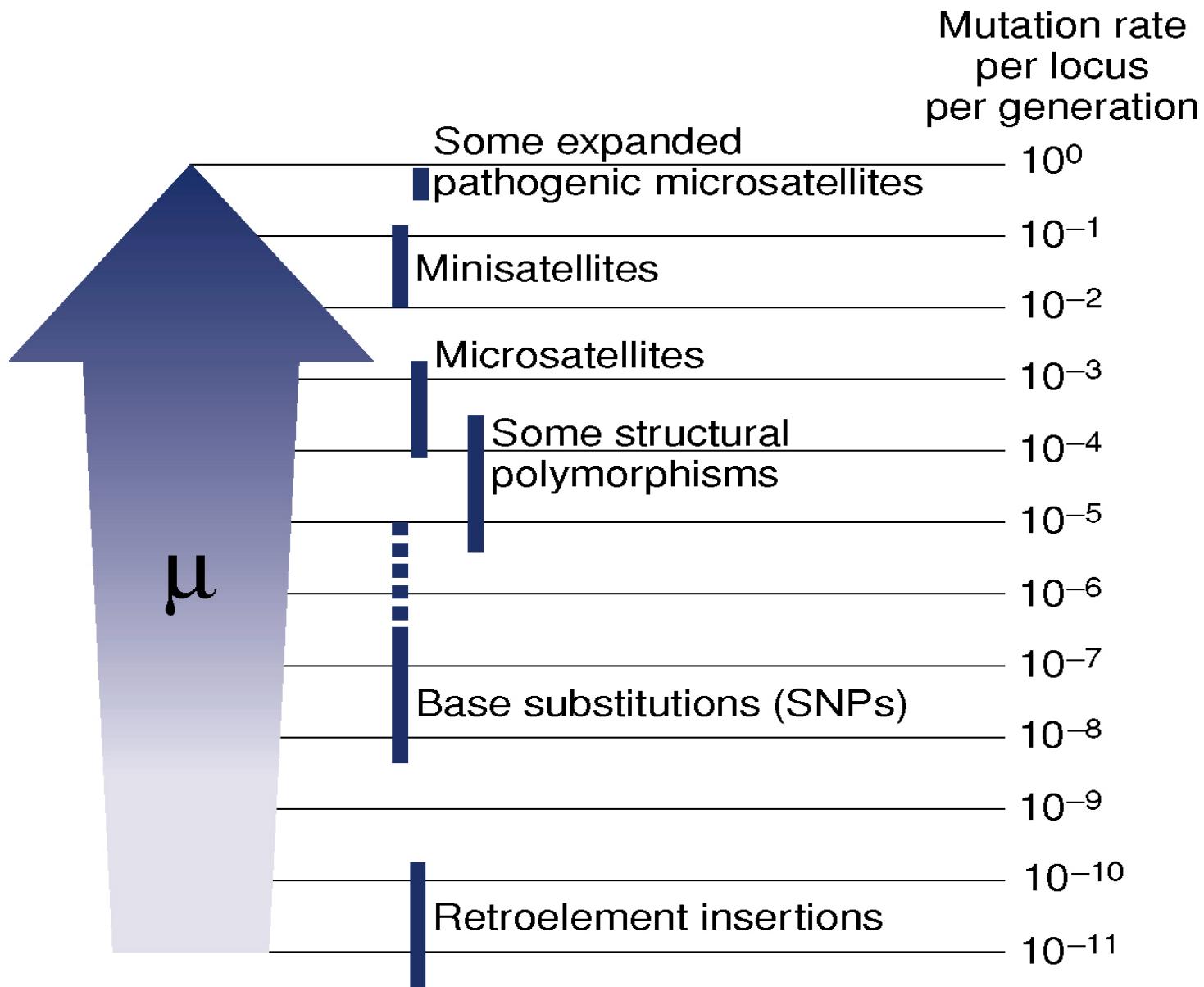
STR System	Maternal Meioses (%)	Paternal Meioses (%)	Number from either	Total Number of Mutations	Mutation Rate
CSF1PO	95/304,307 (0.03)	982/643,118 (0.15)	410	1,487/947,425	0.16%
FGA	205/408,230 (0.05)	2,210/692,776 (0.32)	710	3,125/1,101,006	0.28%
TH01	31/327,172 (0.009)	41/452,382 (0.009)	28	100/779,554	0.01%
TPOX	18/400,061 (0.004)	54/457,420 (0.012)	28	100/857,481	0.01%
VWA	184/564,398 (0.03)	1,482/873,547 (0.17)	814	2,480/1,437,945	0.17%
D3S1358	60/405,452 (0.015)	713/558,836 (0.13)	379	1,152/964,288	0.12%
D5S818	111/451,736 (0.025)	763/655,603 (0.12)	385	1,259/1,107,339	0.11%
D7S820	59/440,562 (0.013)	745/644,743 (0.12)	285	1,089/1,085,305	0.10%
D8S1179	96/409,869 (0.02)	779/489,968 (0.16)	364	1,239/899,837	0.14%
D13S317	192/482,136 (0.04)	881/621,148 (0.14)	485	1,558/1,103,282	0.14%
D16S539	129/467,774 (0.03)	540/494,465 (0.11)	372	1,041/962,239	0.11%
D18S51	186/296,244 (0.06)	1,094/494,098 (0.22)	466	1,746/790,342	0.22%
D21S11	464/435,388 (0.11)	732/526,708 (0.15)	580	1,816/962,096	0.19%
Penta D	12/18,701 (0.06)	21/22,501 (0.09)	24	57/41,202	0.14%
Penta E	29/44,311 (0.065)	75/55,719 (0.135)	59	163/100,030	0.16%
D2S1338	15/72,830 (0.021)	157/152,310 (0.10)	90	262/225,140	0.12%
D19S433	38/70,001 (0.05)	78/103,489 (0.075)	71	187/173,490	0.11%
SE33 (ACTBP2)	0/330 (<0.30)	330/51,610 (0.64)	None reported	330/51,940	0.64%

STR lókusok mutációs rátája: $10^{-3} - 10^{-4}$ / meiózis

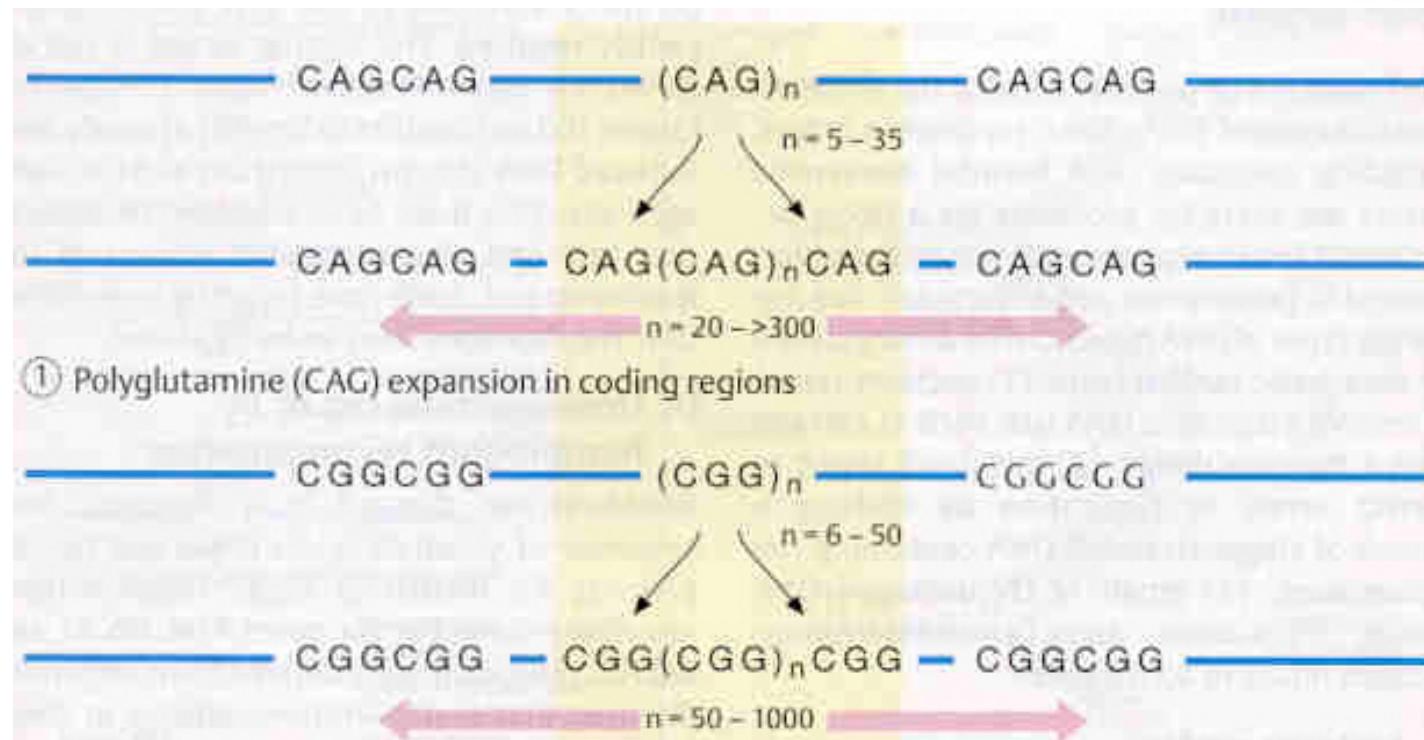
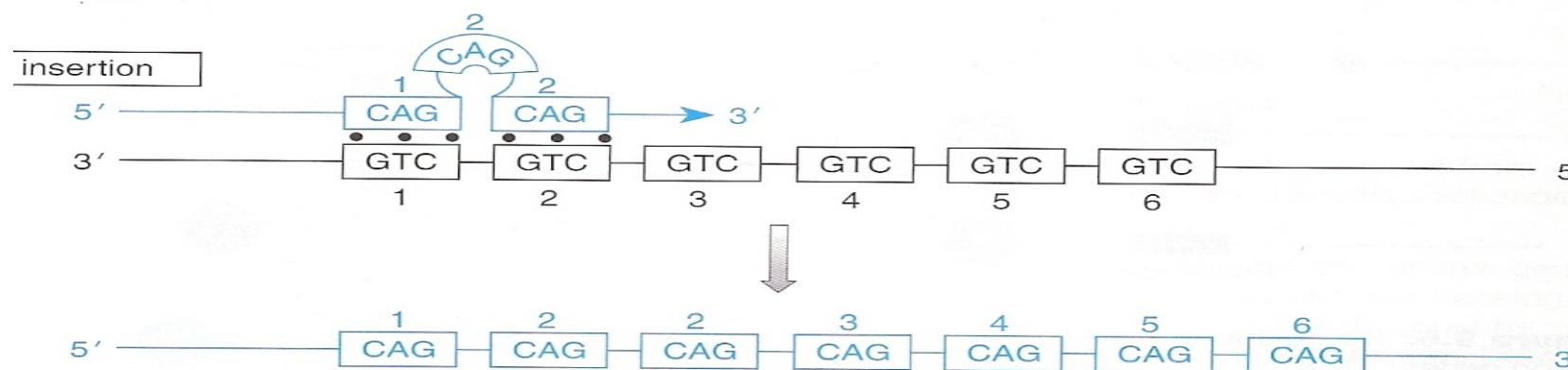
„Replikációs slippage” - Mikroszatellita mutáció



Polimorf szekvenciák mutációs rátája (μ)



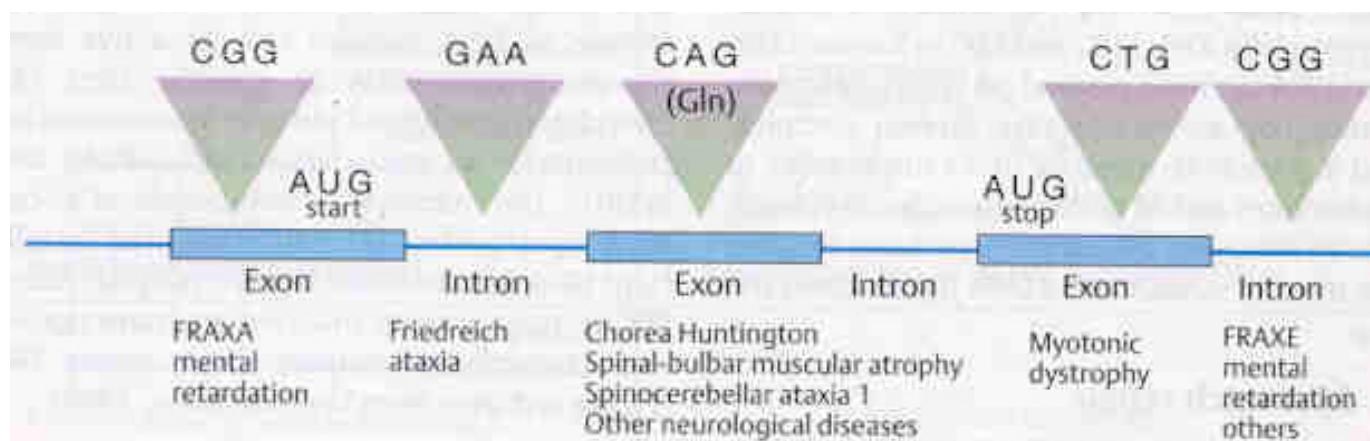
Trinukleotid repeat expansió I.



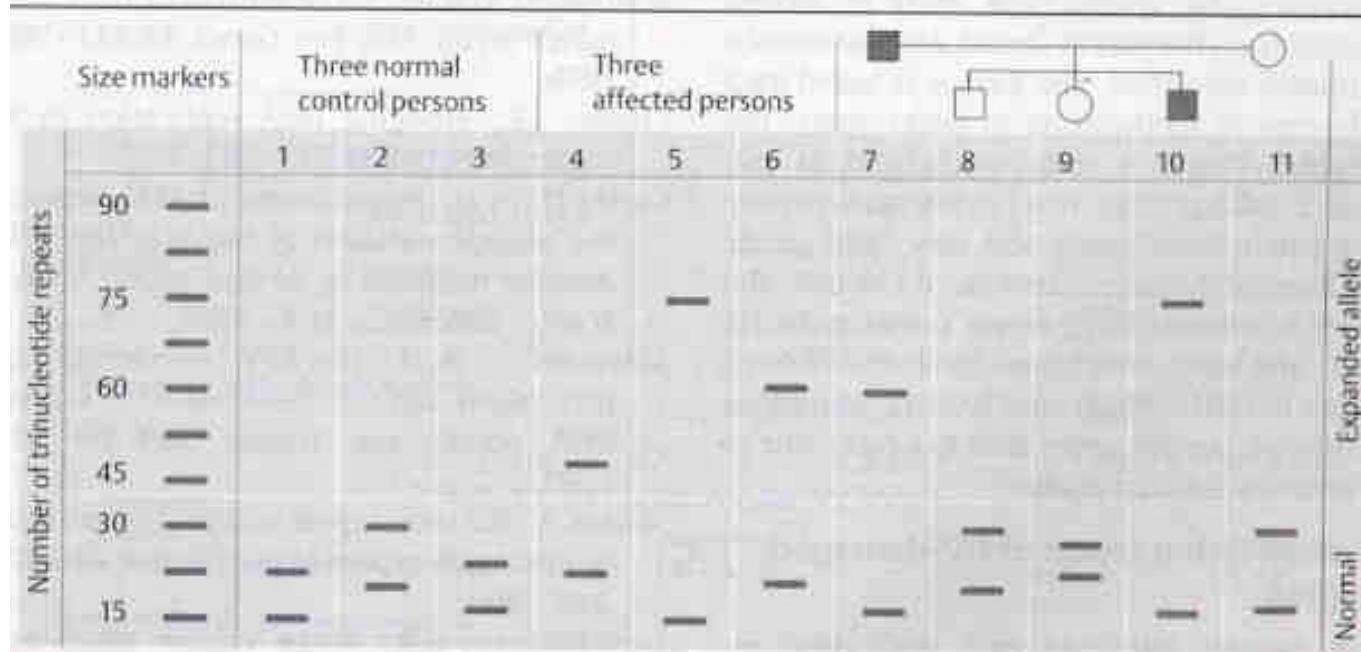
A. Different types of trinucleotide repeat expression

Instabil mutáció!

Trinukleotid repeat expansió II.



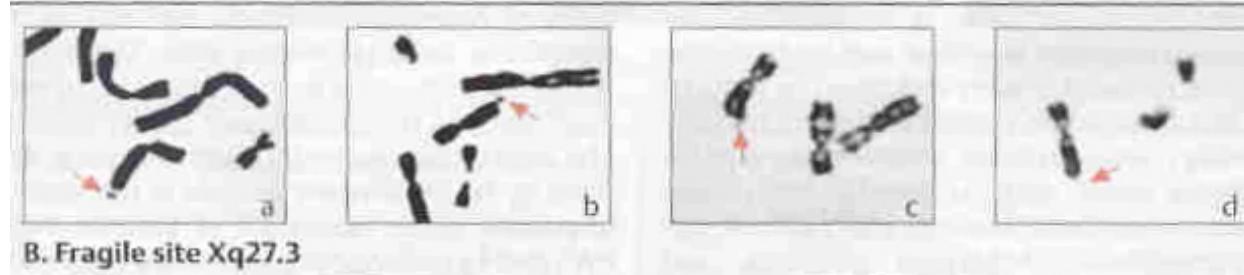
B. Unstable trinucleotide repeats in different diseases



C. Principle of laboratory diagnosis of unstable trinucleotide repeats leading to expansion

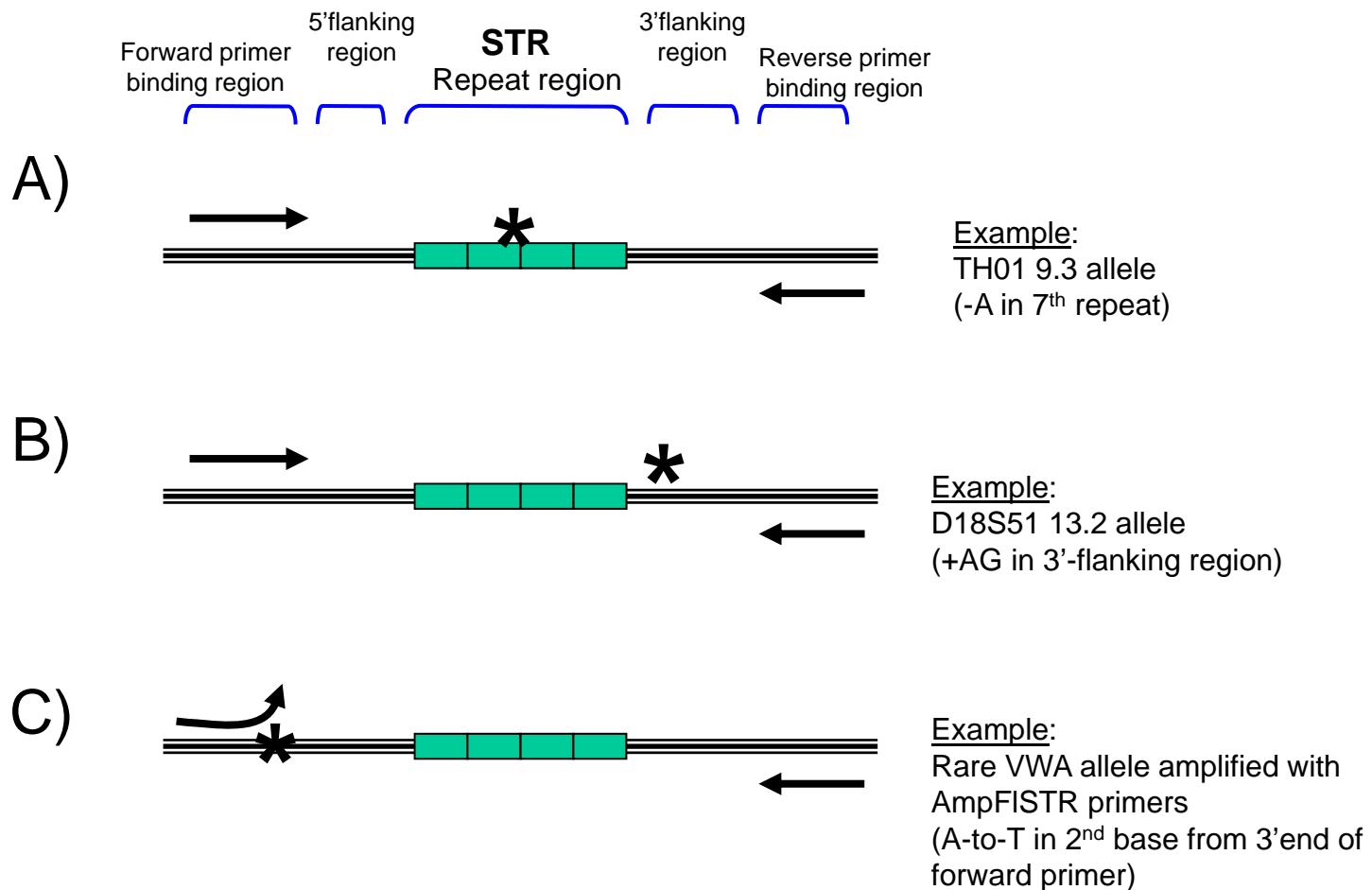
Repeat expanziók okozta defektusok

Disease (Examples)	Gene	Frequency	Tri-nucleotide	Normal Number	Mutant Allele	Chromosome
Huntington disease	HD	1:10 000	(CAG) _n	0–26	36–121	4p16.3
Fragile X syndrome	FMR1	1:5 000	(CGG) _n	6–50	52–500	Xq27.3
Myotonic dystrophy	DMPK	1:8 000	(CTG) _n	5–37	50–500	19q13.2
Spinal-bulbar muscular atrophy (Kennedy)	SBMA	<1:50 000	(CAG) _n	11–31	36–65	Xq11-12

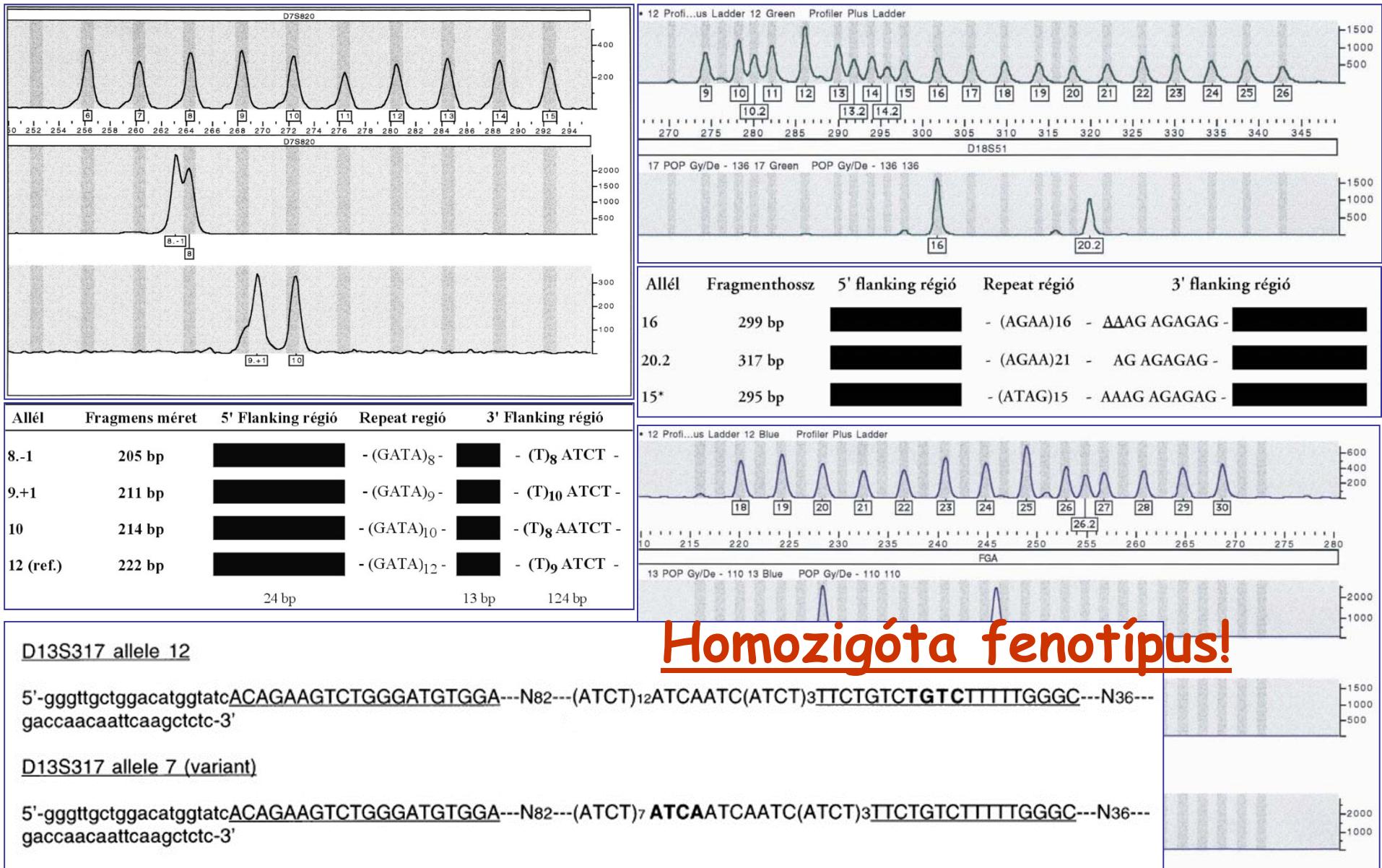


Fragilis X
Huntington disease
Myotonic dystrophy
Friedrich ataxia
stb.

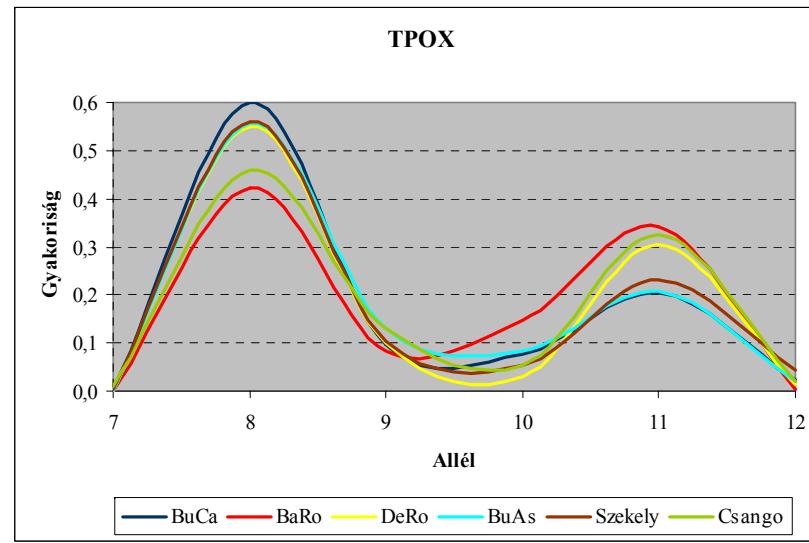
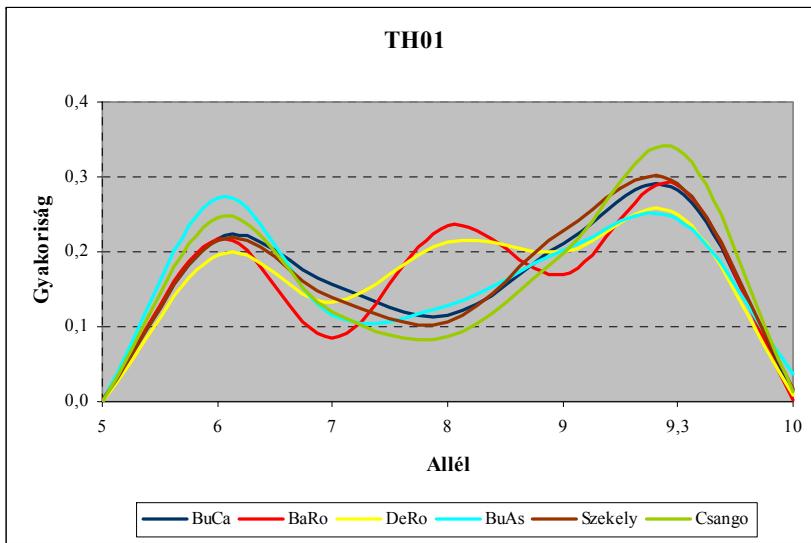
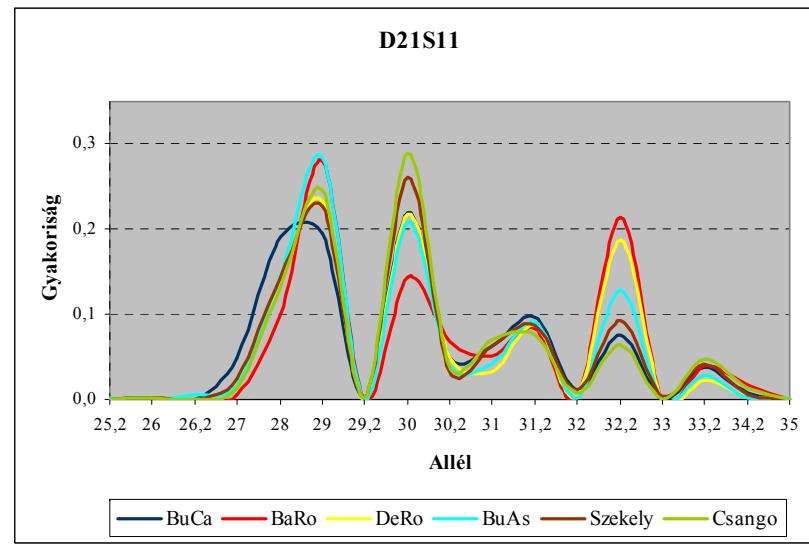
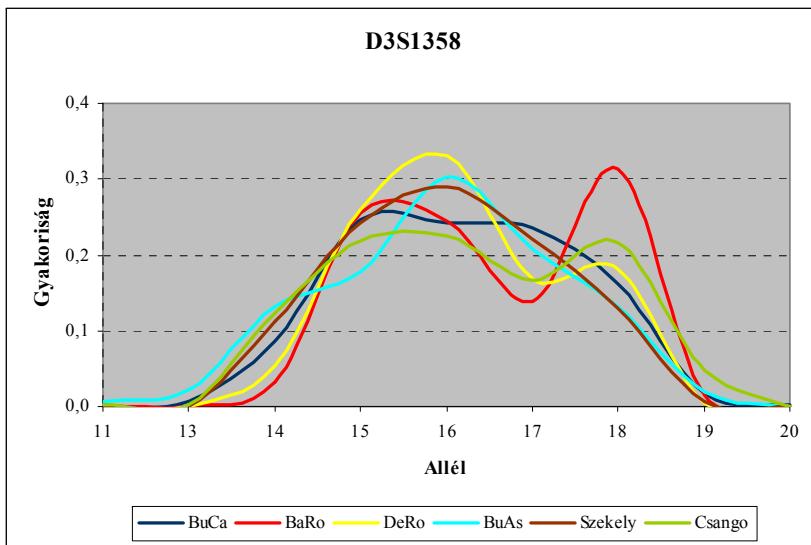
Mikroszatellita pontmutációk



Variáns mikroszatellita allélek: Null-allélek

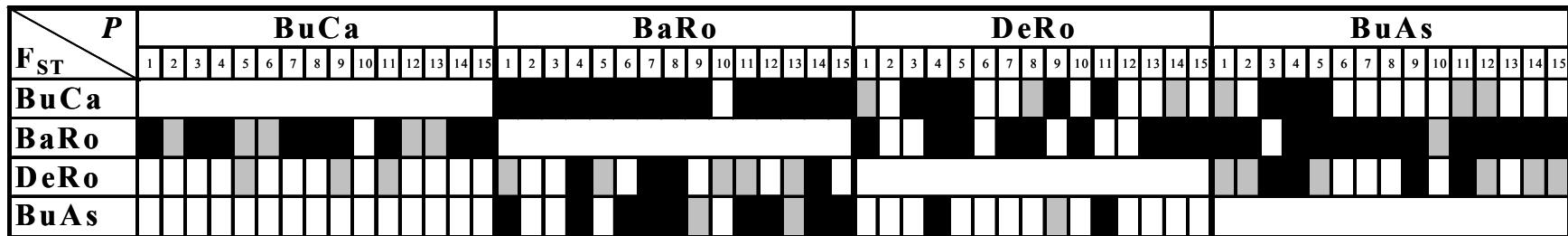


Mikroszatellita allélgyakoriság diagramok

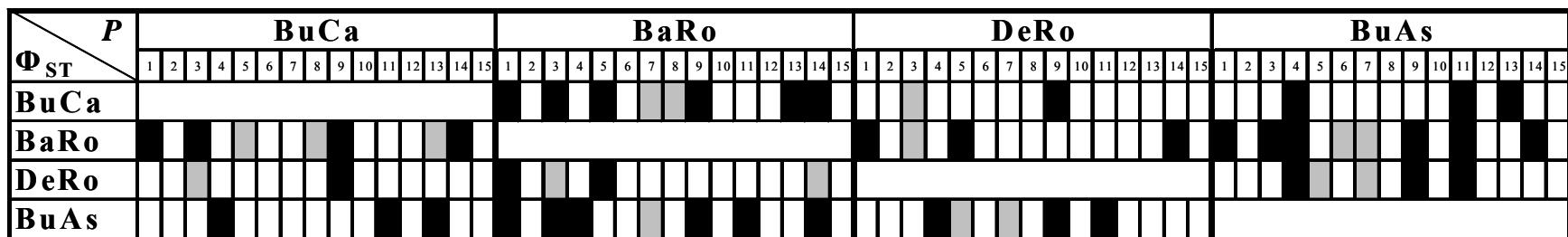


Genetic Structure (AMOVA)

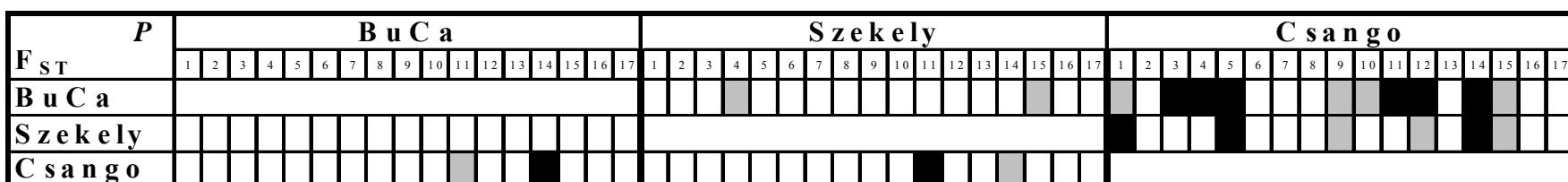
a,



b,



Jelölések: ■ ($F_{ST}, \Phi_{ST} > 0,02; P < 0,1$) ■■ ($0,02 > (F_{ST}, \Phi_{ST}) > 0,01; 0,01 < P < 0,05$) □ ($(F_{ST}, \Phi_{ST}) < 0,01; P > 0,05$)



b ,



Jelölések: ■ ($F_{ST}, \Phi_{ST} > 0,02; P < 0,1$) ■■ ($0,02 > (F_{ST}, \Phi_{ST}) > 0,01; 0,01 < P < 0,05$) □ ($(F_{ST}, \Phi_{ST}) < 0,01; P > 0,05$)

Fajtaazonosítás?....

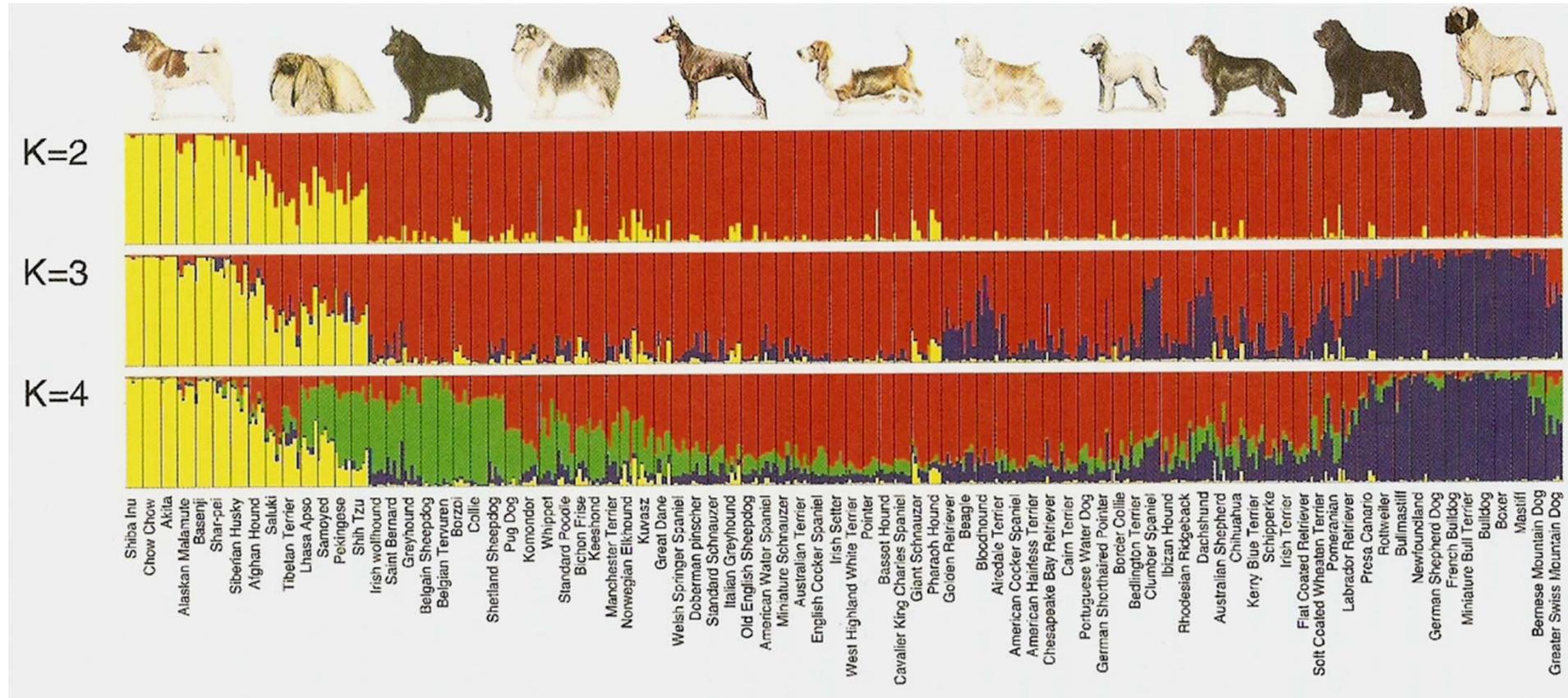
Populációs vizsgálatok!!!

1. Ősi

2. Terelő

3. Vadász

4. Védő

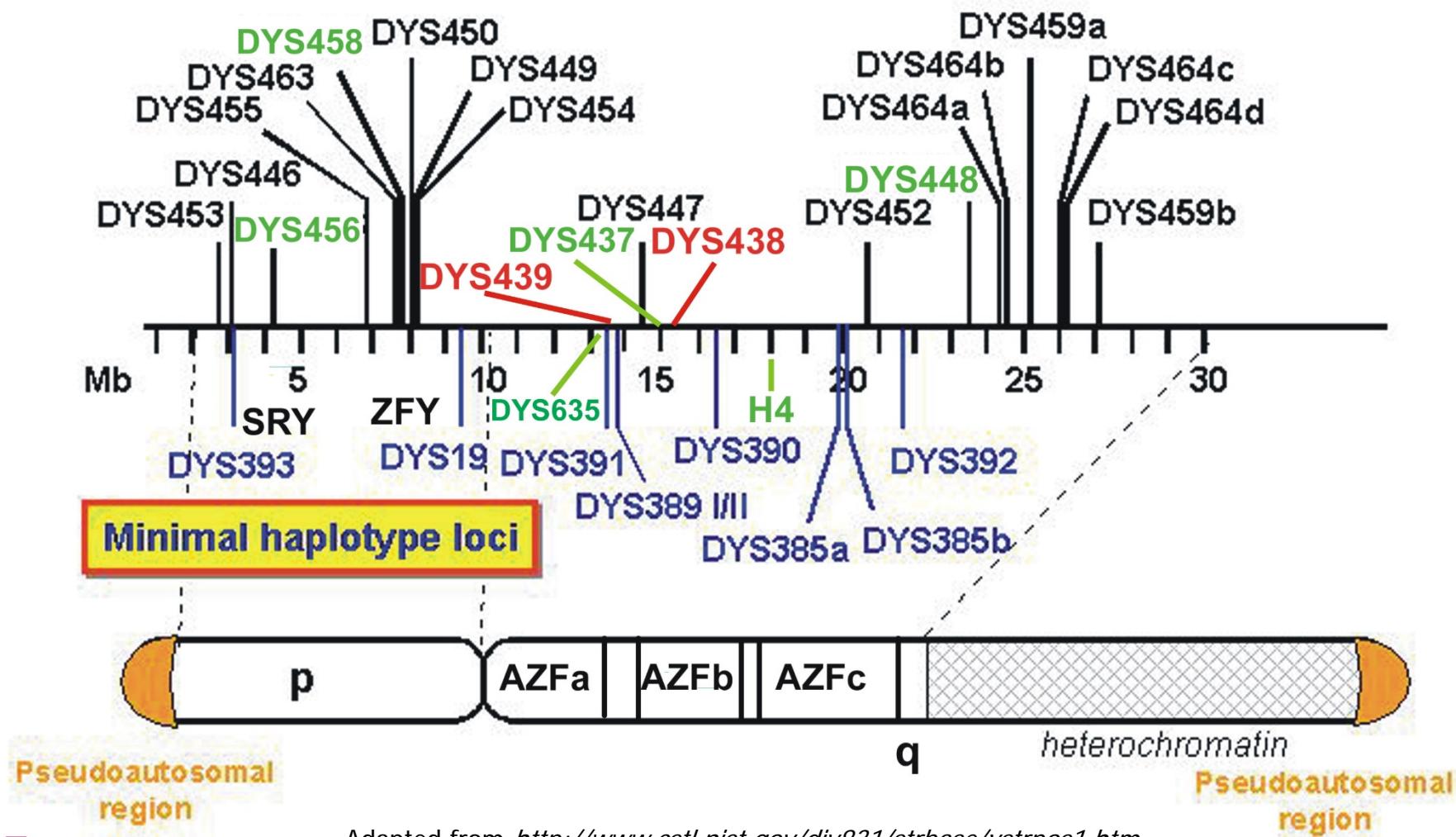


- 85 fajta (n=414)
- 95 mikroszatellita marker (dinukleotid repeat)
- 99%-ban helyes besorolás

Y STR Positions along Y Chromosome

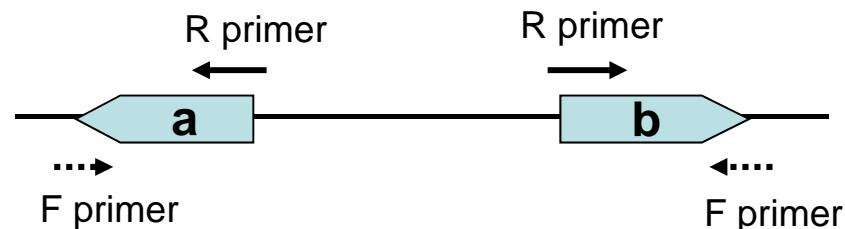
Extended haplotype loci

ABI AmpF/STR Yfiler loci

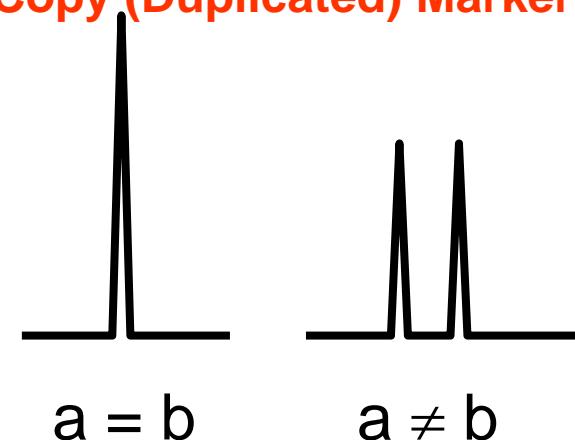


Adapted from <http://www.cstl.nist.gov/div831/strbase/ystrpos1.htm>

(A) DYS385 a/b

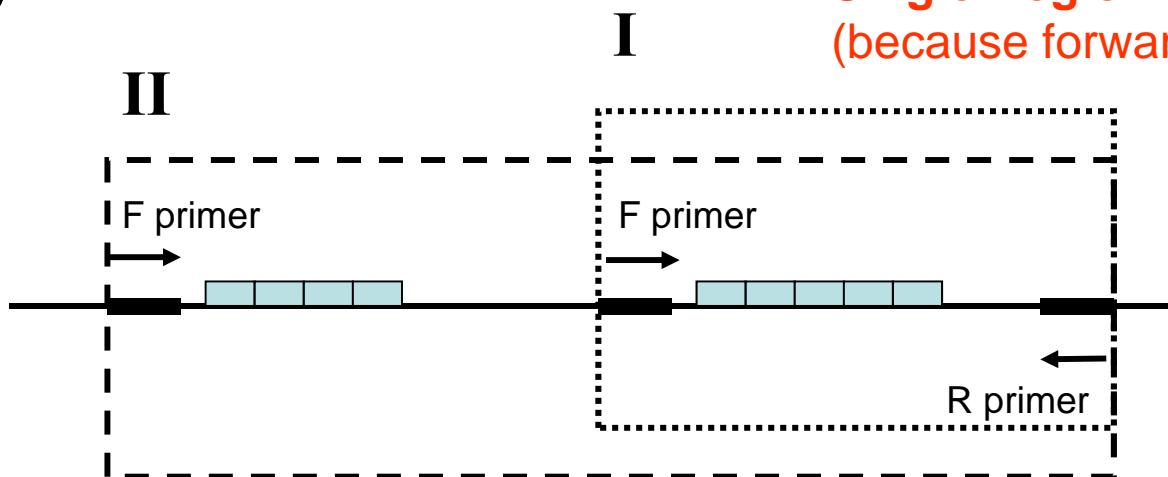


Multi-Copy (Duplicated) Marker

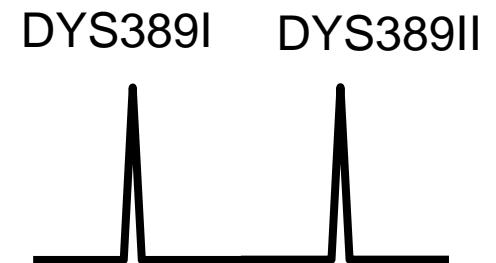


*Duplicated regions are
40,775 bp apart and facing
away from each other*

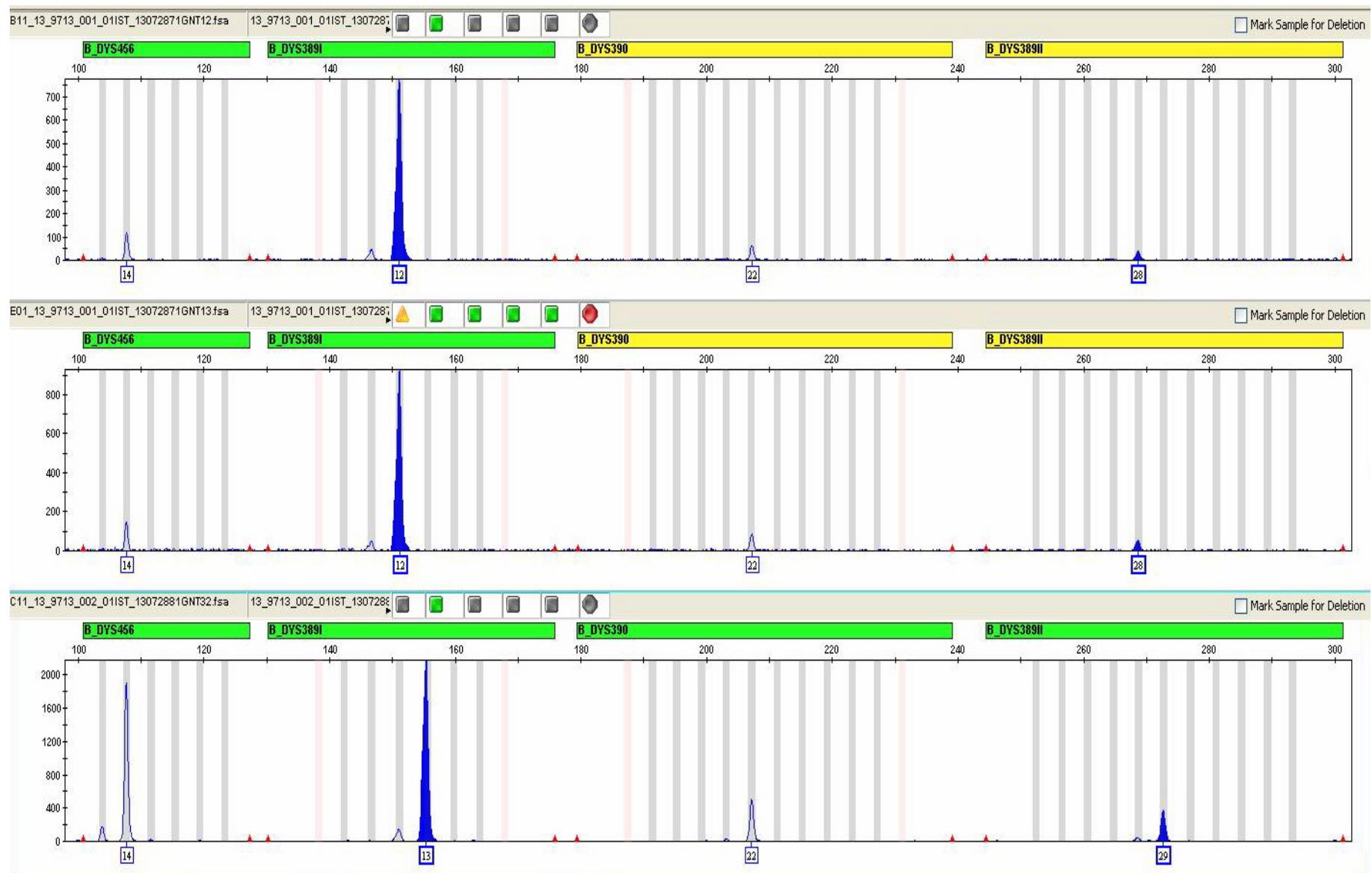
(B) DYS389 I/II



Single Region but Two PCR Products
(because forward primers bind twice)



DYS389 I-II mutation



Mobilis Genetikai Elemek

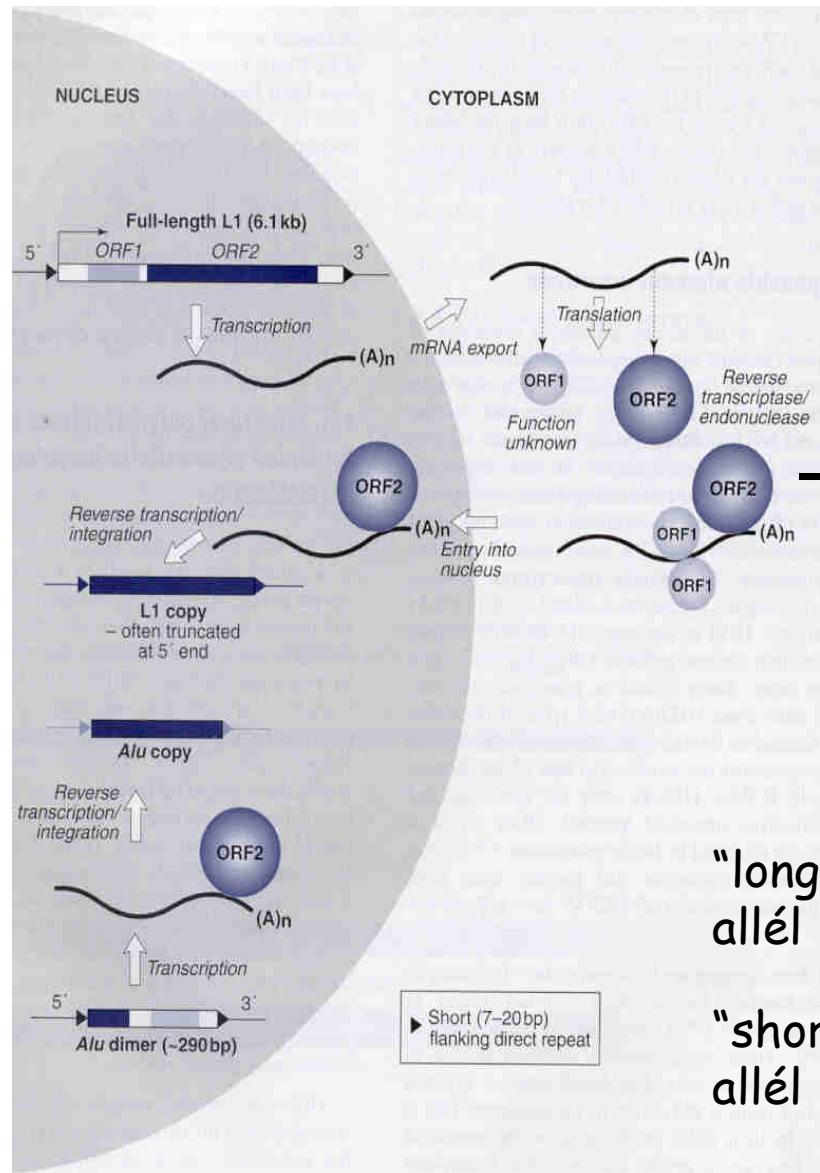
- Extra mennyisége a humán genomban: 100/gén (~ 45 %)
- Humán genom ~ 75 %-a lehet mobilis elem maradványa
- Mutagén hatások: pl. inzerció, nem homológ rekombináció,
→ negatív konzekvencia a gazdára nézve
- Retrotranszpozon: „copy-and-paste”, LINEs, SINEs, LTRs
- Transzpozonok: „cut-and-paste”

TABLE 2.2: CLASSES OF DISPERSED REPEATS IN THE HUMAN GENOME.

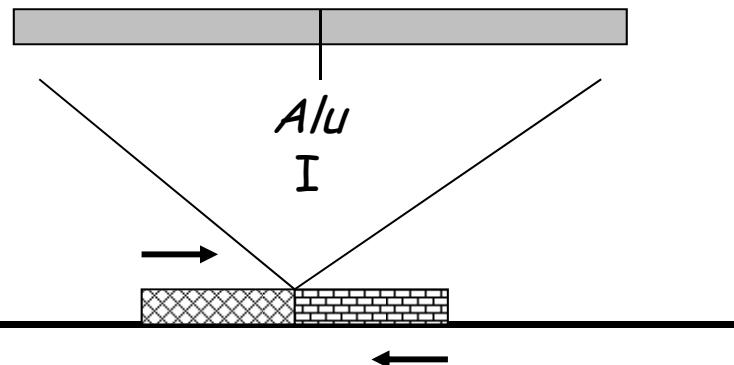
Class	Copy no. per haploid genome	Fraction of genome	Autonomous transposition or retrotransposition?	Length
LINEs	850 000	21%	Yes	Up to 6–8 kb
SINEs	1 500 000	13%	No	Up to 100–300 bp
Retrovirus-like elements	450 000	8%	Complete copies, yes	6–11 kb (1.5–3 kb)
DNA transposon copies	300 000	3%	Complete copies, yes	2–3 kb (80–3000 bp)

Values given in parentheses are lengths of incomplete elements, incapable of autonomous transposition (see Section 3.4). Adapted from Lander *et al.* (2001).

Mobilis elemek: biallélikus hossz-polimorfizmus



Human *A/u* Repeat
(~300 bp)



Kétféle alléltípus

"long" (+)
allél



400
bp

"short" (-)
allél



100
bp

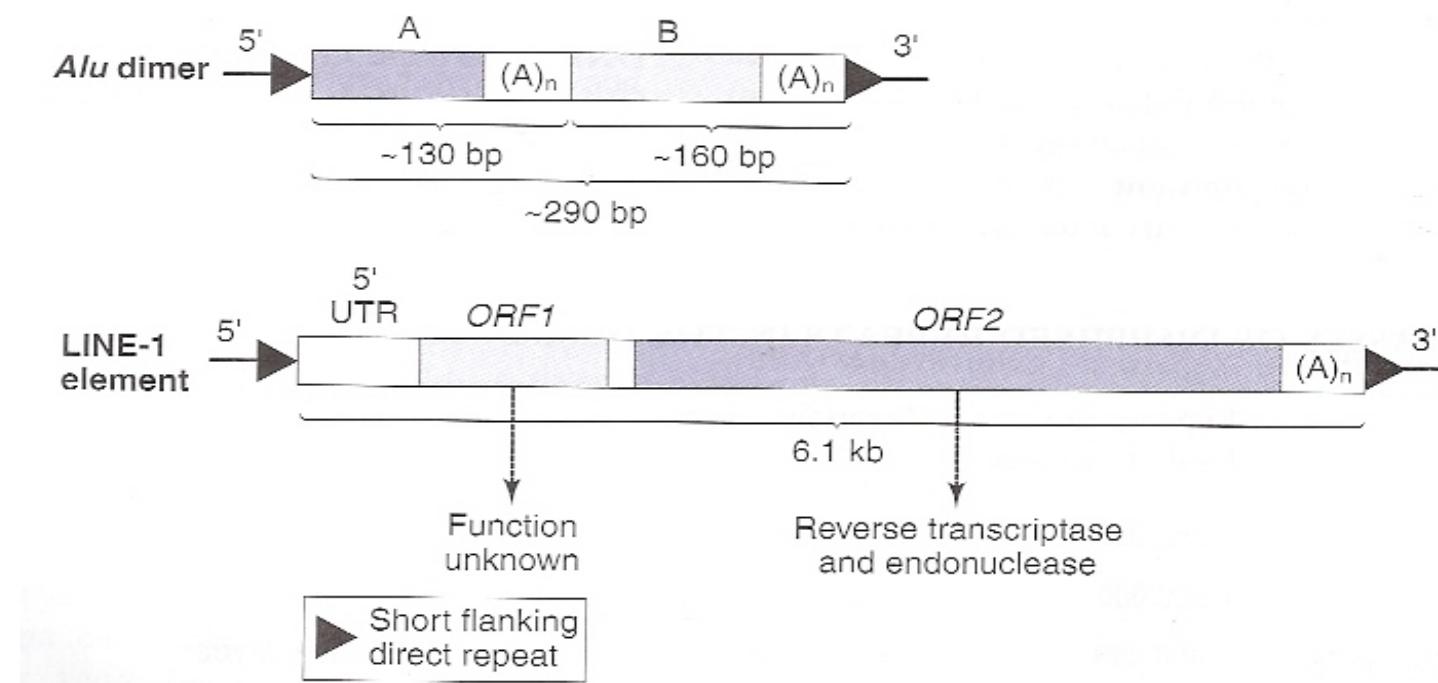
► Short (7–20 bp)
flanking direct repeat

Long Interspersed Nuclear Elements - LINEs

- LINEs v. *Kpn*: emberi genom 20 %-a, 870.000 kópia
- kb. 100 LINEs működőképes retrotranszpozonként
 - ~ 6 kb, belső 5' promóter, 2 ORF (RNS-kötő fehérje, endonukleáz + reverz transzkriptáz), poly(A)-farok,
- Target-primed reverz transzkripció: TT | AAAA - target
- Hanyag másolás
 - (transzkripció „read-through”, „dead-on-arrival”, nagyobb szekvenciarészek átrendeződése, egyéb nem autonóm szekvenciákhoz való kötődés)
- Önmaguktól nem tudnak a genomból kivágódni (deléció)
- ősi és relatíve új szekvenciák (pl. LINE-1: 5 MYS)

Short Interspersed Nuclear Elements - SINEs

- SINEs v. *Alu*: 1.500.000 kópia, 70 % *AluI*, 300 bp,
- Főemlős specifikus, *Alu I*: AGCT, polimorfizmusok,
- Nem kódoló szekvencia, önállóan nem mobilizálódik
- *Alu* - LINE-1 retrotranszpozíció, 0.05 /genom / generáció



Egynukleotid polimorfizmusok (SNPs)

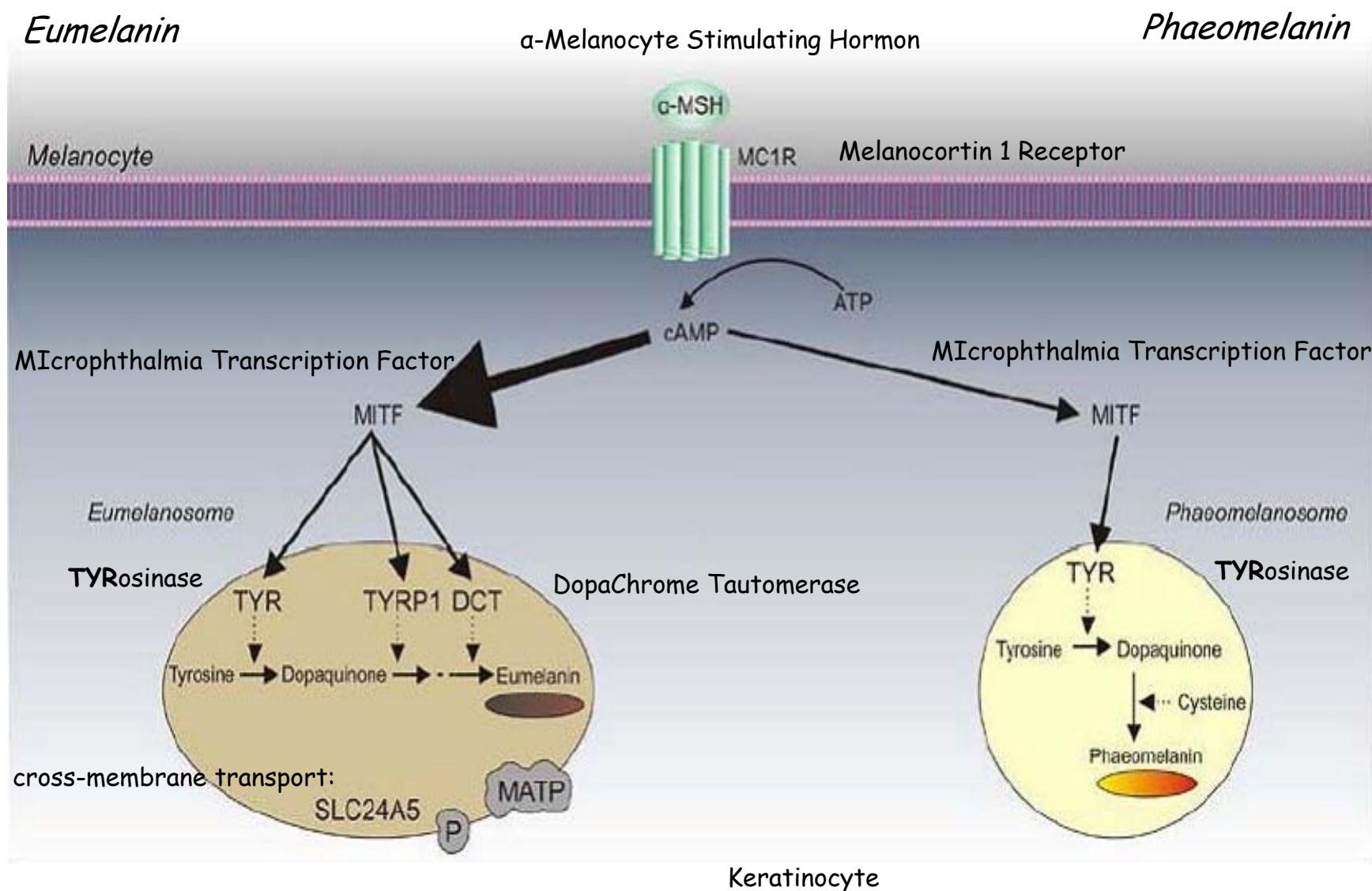
- Biallélikus markerek (6 lehetőség)
 - (A / G, C / T, A / T, C / G, T / G, A / C)
- Több millió SNP a genomban
 - kb. 500-1000 bázispáronként
 - pontmutációk génekben és/vagy regulátor régiókban
- Fenotípus kapcsolatok
 - pigmentáció, testalkat, ...
- Leszármazási vonalak (Y-SNP's)
- Diagnosztika
 - multifaktoriális poligénes betegségek

SNP markerek a humán genomban

TABLE 12.2 Categories of SNP Markers (See Budowle & van Daal 2008, Butler et al. 2008).

Category	Characteristics	Examples
Identity SNPs Individual Identification SNPs (IISNPs)	SNPs that collectively give very low probabilities of two individuals having the same multi-locus genotype	FSS 21plex (Dixon et al. 2005) SNPforID 52plex (Sanchez et al. 2006) Kidd group SNPs (Pakstis et al. 2010)
Lineage SNPs Lineage Informative SNPs (LISNPs)	Sets of tightly linked SNPs that function as multi-allelic markers that can serve to identify relatives with higher probabilities than simple bi-allelic SNPs	mtDNA coding region SNPs (Coble et al. 2004) Japanese Y-SNPs (Mizuno et al. 2010) Haplotype blocks (Ge et al. 2010)
Ancestry SNPs Ancestry Informative SNPs (AISNPs)	SNPs that collectively give a high probability of an individual's ancestry being from one part of the world or being derived from two or more areas of the world	SNPforID 34plex (Phillips et al. 2007b) 24 SNPs (Lao et al. 2010) FSS YSNPs (Wetton et al. 2005)
Phenotype SNPs Phenotype Informative SNPs (PISNPs)	SNPs that provide a high probability that the individual has particular phenotypes, such as a particular skin color, hair color, eye color, etc.	Red hair (Grimes et al. 2001) "Golden" gene pigmentation (Lamason et al. 2005) IrisPlex eye color (Walsh et al. 2010)

A humán melanogenezis



A bőrpigmentáció kialakulásáért felelős gének

Principal skin pigmentation candidate genes

Locus	Chromosome	Protein	Mut phenotype	Function
Melanosome proteins				
TYR	11q14-11q21	Tyrosinase	OCA1	Oxidation of tyrosine
TYRP1	9p23	Gp75, TRYPI	OCA3	DHICA-oxidase, TYR stabilisation
DCT	13q32	DCT, TRYPII		Dopachrome tautomerase
OCA2	15q11.2-15q12	P-protein	OCA2 (eye)	pH of melanosome
SLC45A2	5p14.3-5q12.3	MATP, AIM-1	OCA4 (skin)	Melanosome maturation
SLC24A5	15q21.1	Cation exchanger		Melanosome precursor
Signal proteins				
ASIP	20q11.2-20q12	Agouti signal protein		MC1R antagonist
MC1R	16q24.3	MSH receptor	Red hair (skin)	G-protein coupled receptor
POMC	16q24.3	MSH receptor	Red hair	MC1R antagonist
OA1	Xp22.3	OA1 protein	OA1	G-protein coupled receptor
MITF	3p12.3-3p14.1	MITF	Waardenburg	Transcription factor
Proteins involved in melanosome transport or uptake by keratinocytes				
MYO5A	15q21	MyosinVa	Griselli	Motor protein
RAB27A	15q15-15q21.1	Rab27a	Griselli	RAS family protein
HPS1	10q23.1-10q23.3	HPS1	Hermansky-Pudlak	Organelle biogenesis and size
HPS6	10q24.32	HPS6	Hermansky-Pudlak	Organelle biogenesis

ACTH: adrenocorticotrophin hormone; DCT: dopachrome tautomerase; DHICA: 5,6-dihydroxyindole-2-carboxylic acid; MATP: membrane-associated transporter protein; MC1R: melanocortin-1 receptor; MITF: microphthalmia-associated transcription factor; MSH: melanocyte stimulating hormone; OCA: oculocutaneous albinism; POMC: pro-opiomelanocortin; TYRP1: tyrosinase-related protein 1.

MC1R gén mutációk

Mutations in the MC1R gene, their penetrance and functional significance (where known)

Mutation	Type	Designation	Penetrance (odds ratio)	Functional significance	References (for functional significance and penetrance)
R151C	Mis-sense	R	63.3	Altered cellular location	[16,26]
R160W	Mis-sense	R	63.3	Altered cellular location	[16,26]
D294H	Mis-sense	R	63.3	Impaired G coupling ability	[26,27]
D84E	Mis-sense	R	63.3	Altered cellular location	[16,26]
I155T	Mis-sense	Lack of statistical data—strong familial association		Altered cellular location	[16,26]
V92M	Mis-sense	r	5.1	Reduced α-MSH binding	[26,28,29]
V60L	Mis-sense	r	5.1		[26]
R163Q	Mis-sense	r	5.1	Slightly reduced α-MSH binding	[26,29]
R142H	Mis-sense	Lack of statistical data—strong familial association			[26]

- MC1R allél változatok különböző aktivitással rendelkeznek.
 - 317 AS, 7 transzmembrán domén,
 - SNPs, RHC fenotípus,
 - neandervölgyi pigmentáció
 - genetikai tesztek, fenotípus predikció

SNPs - pigmentációs gének

- ASIP (aguti): 3'UTR 8818A - MSH antagonista - phaeomelanin termelés
- MATP: melanoszóma pH reguláció, 374Leu allél - sötét szín, albinizmus
- SLC24A5: „arany” gén, zebrafish, Ala111Thr allél, világos árnyalat, europid rasszban fixált, szelekciós nyomás?
- OCA2: albinizmus gén, 305 Arg/Trp, Afrika / Európa

Gene	Location	Protein	Reference SNP ID (rs#) ^a	Alleles	Variation type
<i>MC1R</i>	16q24.3	MC1R: melanocortin 1 receptor	rs1805007	C/T	ns coding, c.451C>T, p.R151C
			rs1805008	C/T	ns coding, c.478C>T, p.R160W
<i>HERC2</i>	15q13	Unknown	rs12913832	A/G	Non-coding, intron 86
<i>OCA2</i>	15q11.2-15q12	P-protein: NA+/H+ antiporter or glutamate transporter	rs7495174 rs6497268 or rs4778241 rs11855019 or rs4778138 rs1545397	T/C G/T T/C G/A	Non-coding, intron 1 Non-coding intronic
<i>SLC45A2</i>	5p13.3	MATP: membrane-associated transporter protein	rs16891982	C/G	ns coding, c.1122C>G, p.F374L
<i>SLC24A5</i>	15q21.1	SLC24A5 (or NCKX5): solute carrier family 24, member 5; potassium-dependent sodium–calcium ion exchanger	rs1426654	G/A	ns coding, p.A111T
<i>DCT</i>	13q32	DCT or TYRP2/TRP-2: dopachrome tautomerase or tyrosinase-related protein-2	rs2031526	G/A	Non-coding, intronic

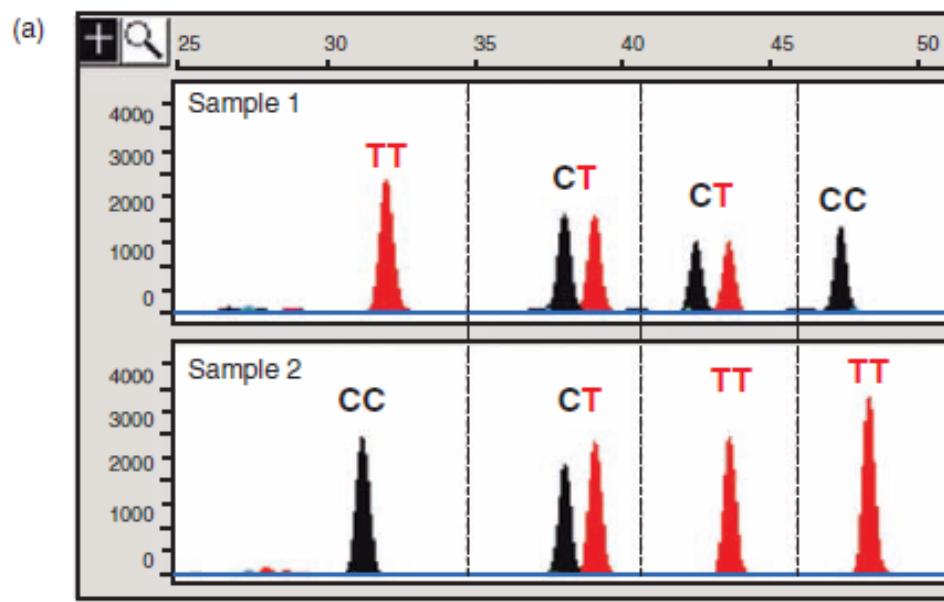
ns non-synonymous

^b Reference SNP ID refer to the reference sequence identifier given to the SNP in the dbSNP database

SNaPshot: A Primer Extension Assay Capable of Multiplex Analysis

Minisequencing
(SNaPshot assay)

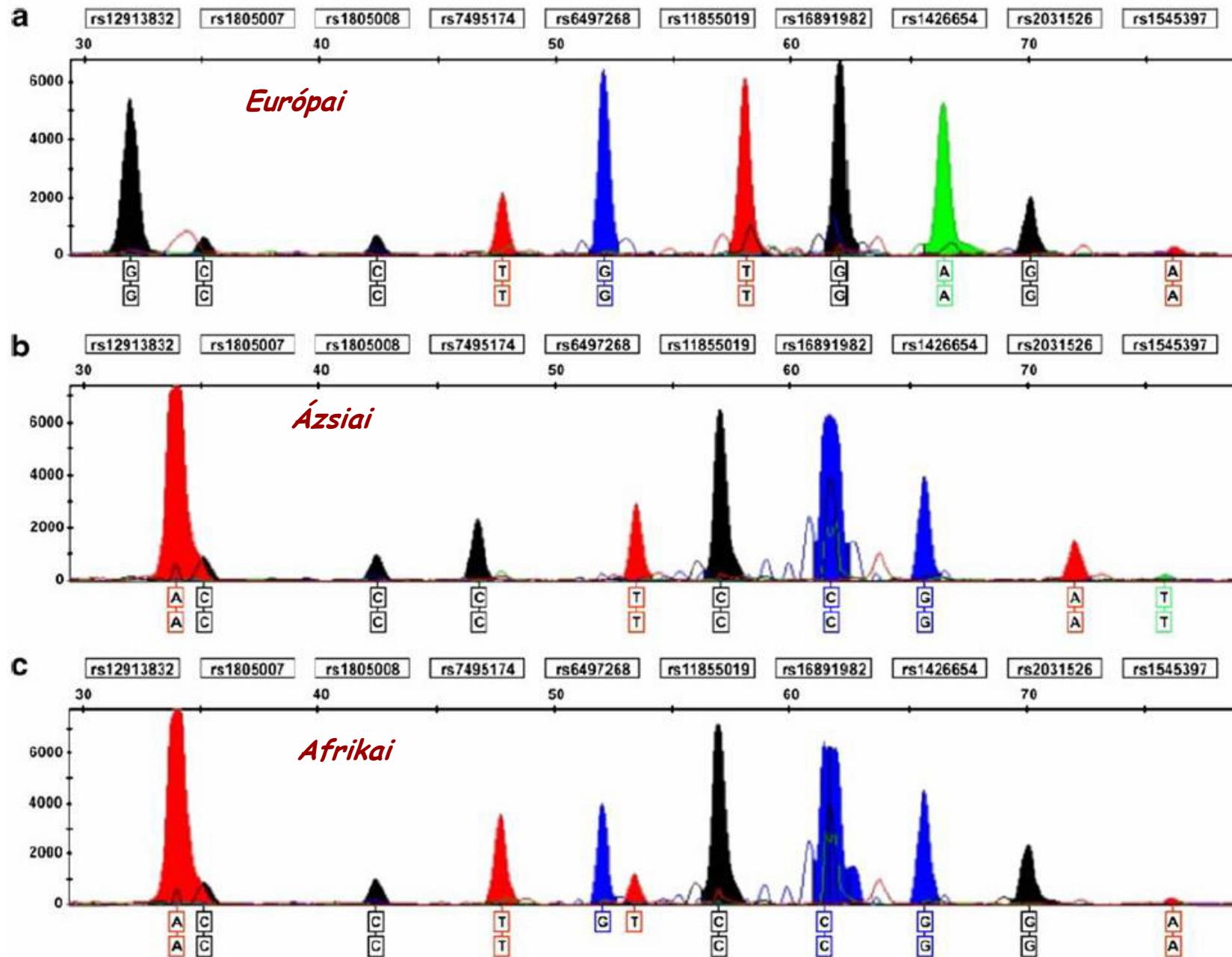
Allele-specific primer extension across the SNP site with fluorescently labeled ddNTPs; mobility modifying tails can be added to the 5'-end of each primer in order to spatially separate them during electrophoresis.



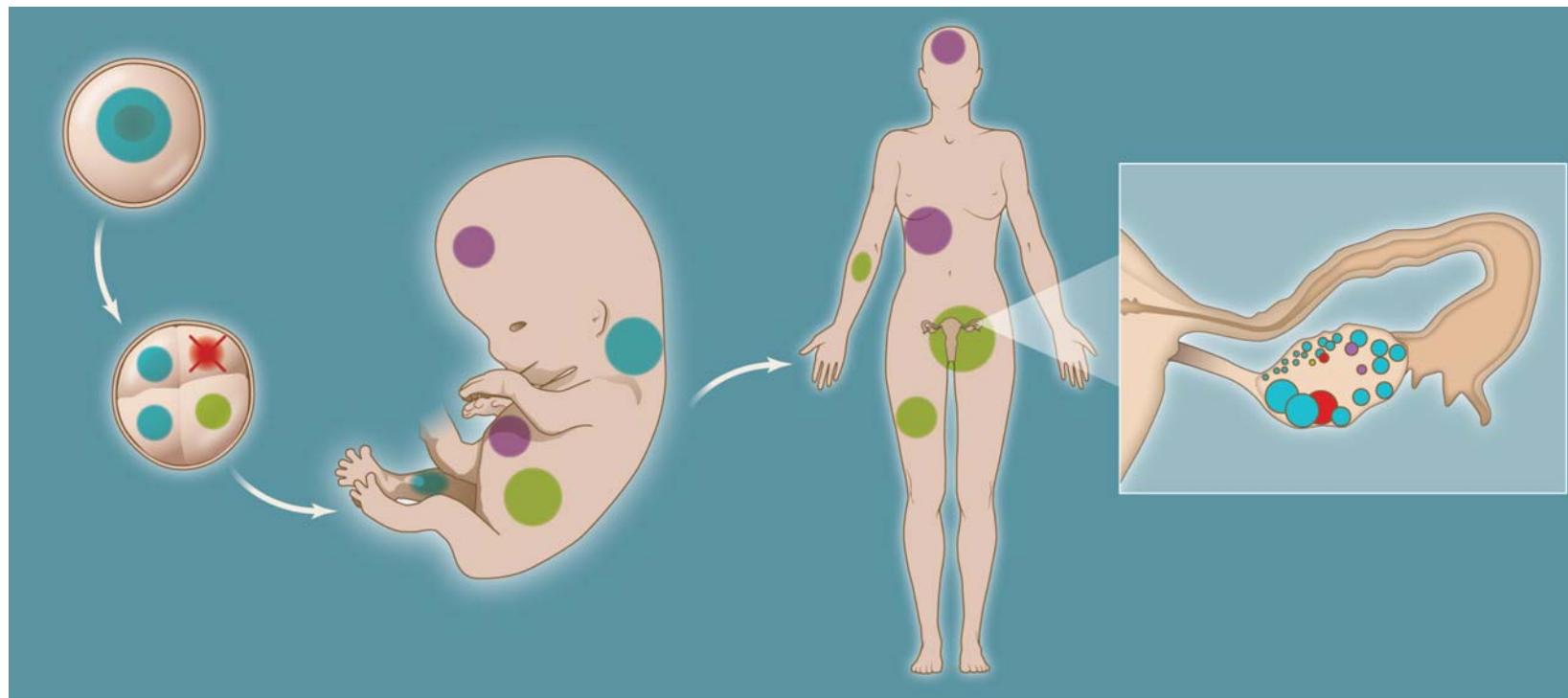
- (b)
- (TTTTT)-primer1 (chromosome 20)-ddT/ddT
 - (TTTTT)-(TTTTT)-primer2 (chromosome 6)-ddC/ddT
 - (TTTTT)-(TTTTT)-(TTTTT)-primer3 (chromosome 14)-ddC/ddT
 - (TTTTT)-(TTTTT)-(TTTTT)-(TTTTT)-primer4 (chromosome 1)-ddC/ddC

FIGURE 12.2 Allele-specific primer extension results using four autosomal SNP markers on two different samples (a). SNP loci are from separate chromosomes (1, 6, 14, and 20) and therefore unlinked. Electrophoretic resolution of the SNP primer extension products occurs due to poly(T) tails that are 5 nucleotides different from one another (b).

10 pigmentációs gén SNP genotipizálás (SNaPshot)



Acquiring mosaicism. Human development from a single fertilized cell to a multicellular organism requires many cell divisions and the genetic material to be replicated many times.



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