Structural bioinformatics

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MTA-ELTE Momentum Bioinformatics Group

4. December 2017

Basic features of protein structures

Structure determination methods

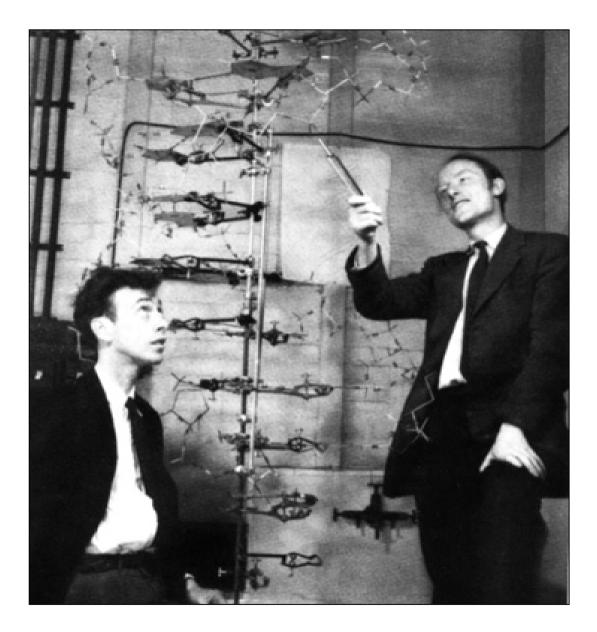
PDB database

Visualization and analysis of structures

Structure comparisons

Structural classification

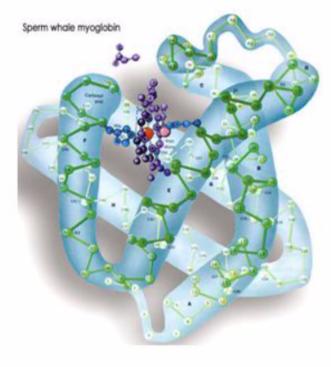
Structure predictions



 "It has not escaped our notice that the specific pairing we have postulated immediately suggests a possible copying mechanism for the genetic material."

1958

- John Kendrew et al., published the first structure of a globular protein, myoglobin.
- Perhaps the most remarkable features of the molecule are its <u>complexity</u> and its <u>lack of</u> <u>symmetry</u>"

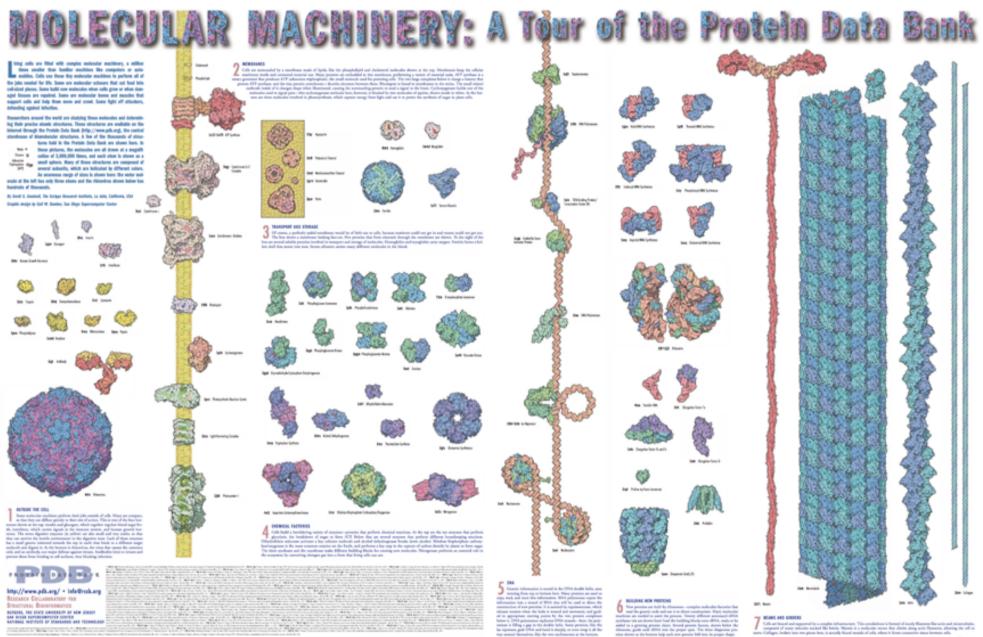


1962

Nobel prize in Chemistry was awarded to Max Perutz and John Kendrew.

Now

~80,000 structures in protein database (PDB)



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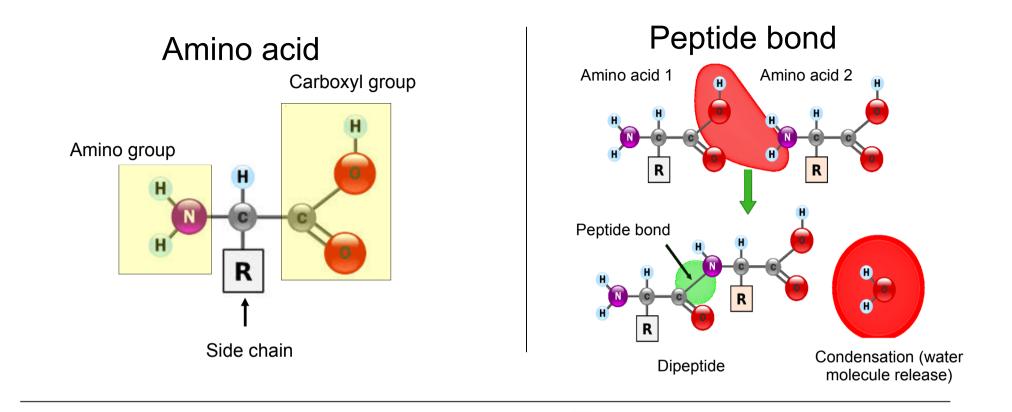
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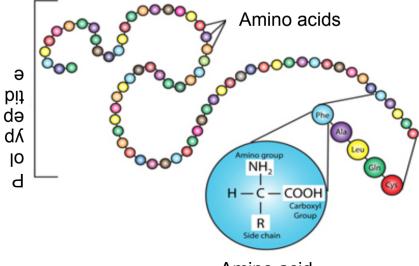
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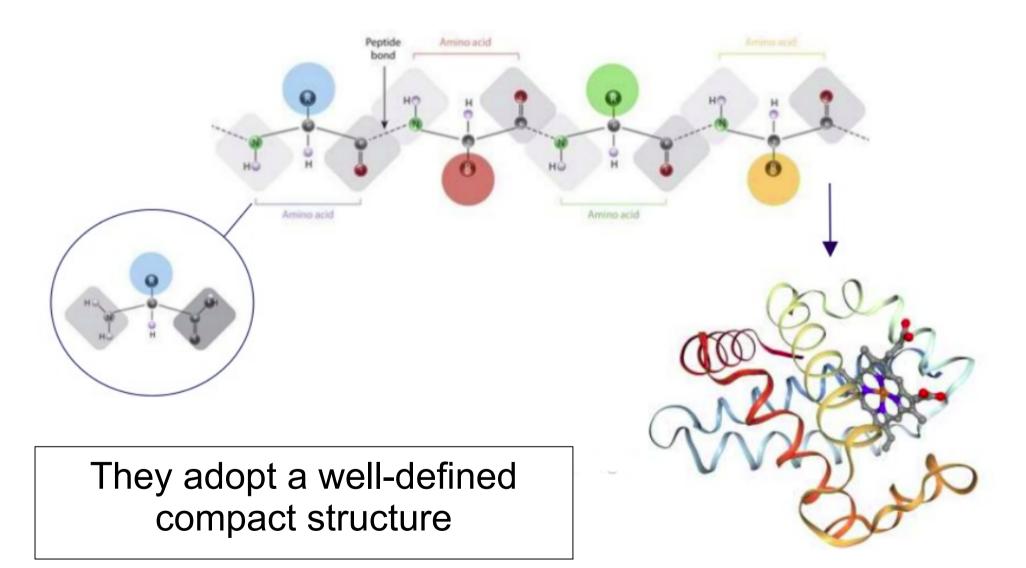




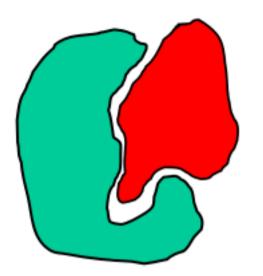


Amino acid

Globular proteins



Why are protein structures interesting?



Function is heavily dependent on the shape of the protein

- Atomic-level understanding of biological processes (DNA, RNA, enzymes, hormones, receptors)
- Understanding the molecular basis of diseases
- Drug design, protein-drug interactions
- No information on e.g. binding strength

Levels of structure for globular proteins

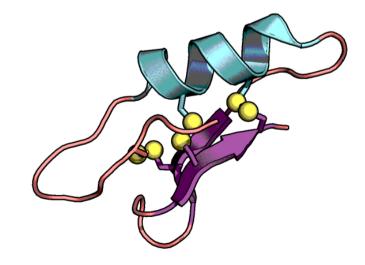
Primary structure = Amino acid sequence MSSVLLGHIKKLEMGHS...

Secondary structure = alpha helix, beta sheets/strands, turns (based on main chain H-bonds)





Tertiary structures = Relative positions of secondary structure elements within the chain



Quaternary structure

Monomer

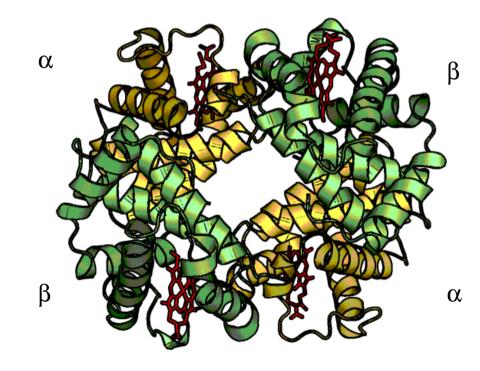
Multi subunit protein complexes

> Homo and hetero oligomers

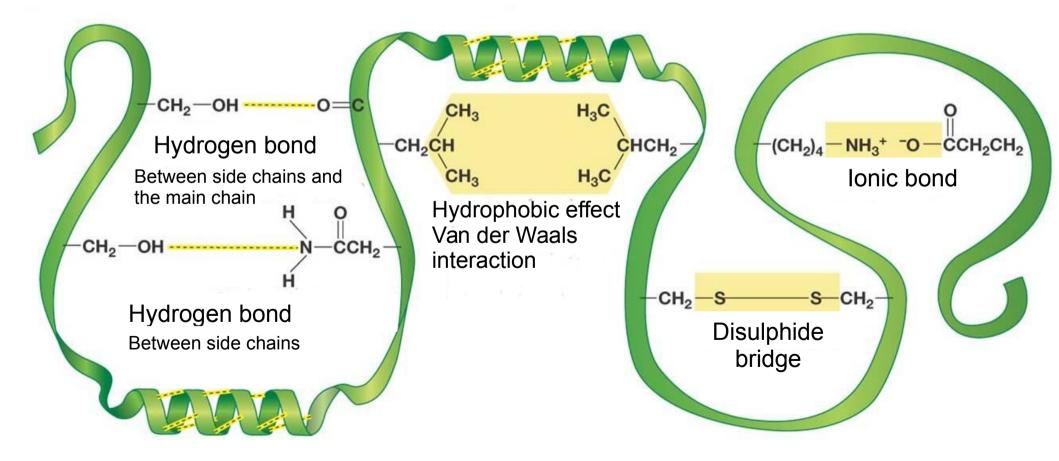
Hemoglobin

Myoglobin

α

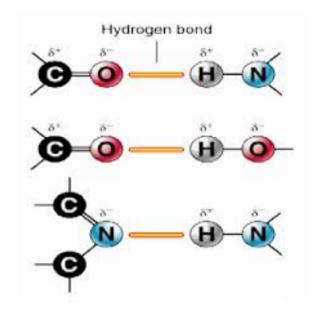


Interactions stabilizing proteins

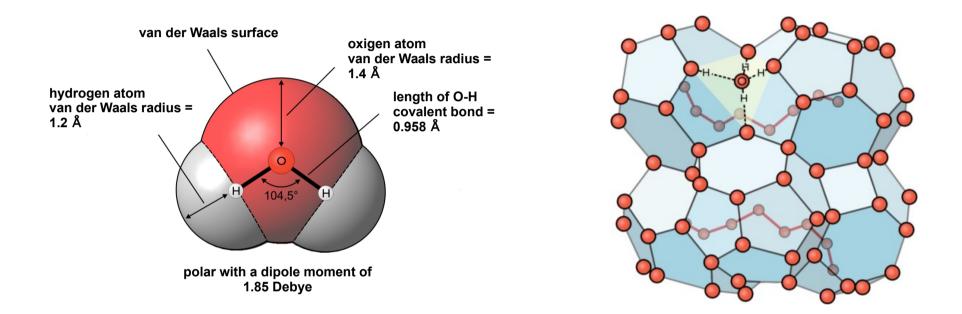


Hydrogen bond

Hydrogen bonds are formed by a H-atom bound in the structure with a **high electronegativity atom** (F, N, O) from a different functional group, i.e. a hydrogen atom establishes a bond between two other atoms.

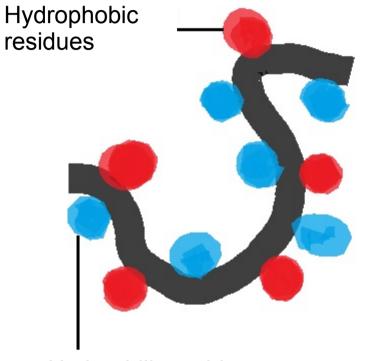


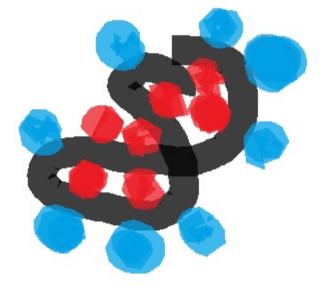
Water molecule



Hydrophobic effect: dominated by entropic terms

Hydrophobicity



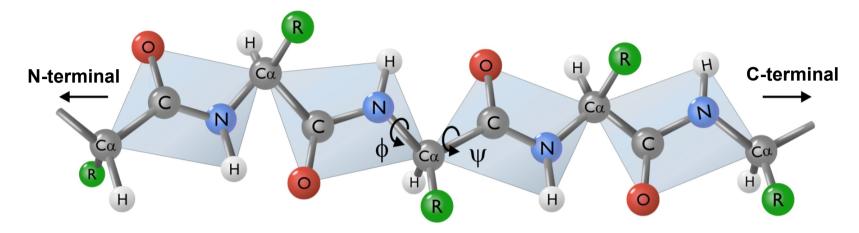


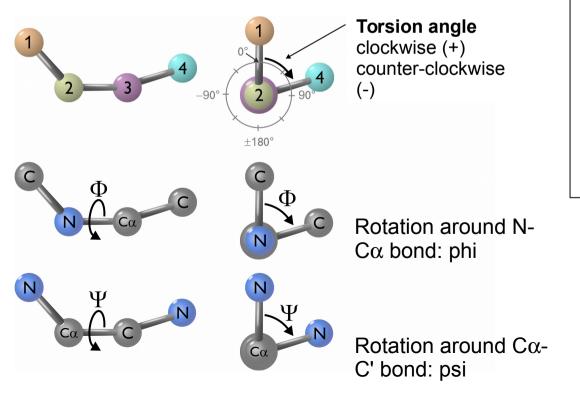
Hydrophilic residues

Protein in isolation

Protein in aquaneous environment

Main chain conformation

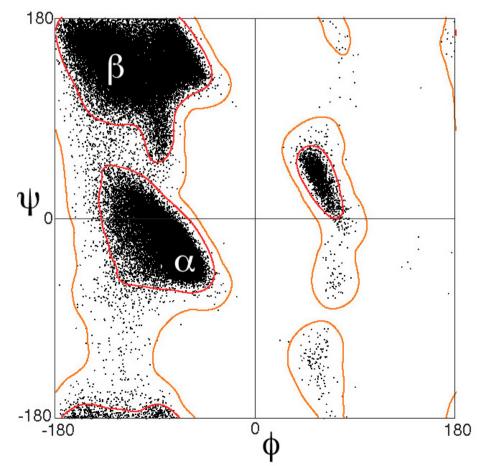




The main chain φ and ψ torsion angles of a protein cannot take arbitrary values, there are preferred conformations.

Ramachandran plot

We can plot the angle-pairs of all residues in a coordinate system using the two torsion angles as X and Y-coordinates.



Glycines and prolines are typically left out from the plot as they have unique conformational preferences. As expected, most residues fall into regions corresponding to α -helices and β strands, but most residues from bends and turns are also within the allowed regions.

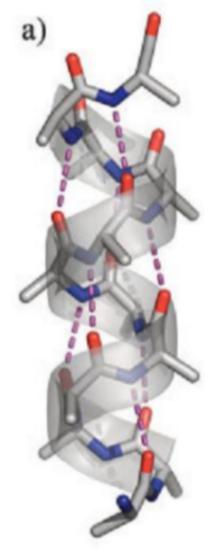
The (right-handed) α helix

Approx. 30% of globular proteins

5-40 residues in length (10 on average)

Individual H-bonds are relatively weak, they have a significant contribution to helix stability

The helix-forming propensity of a peptide segment depends on its sequence



a helix

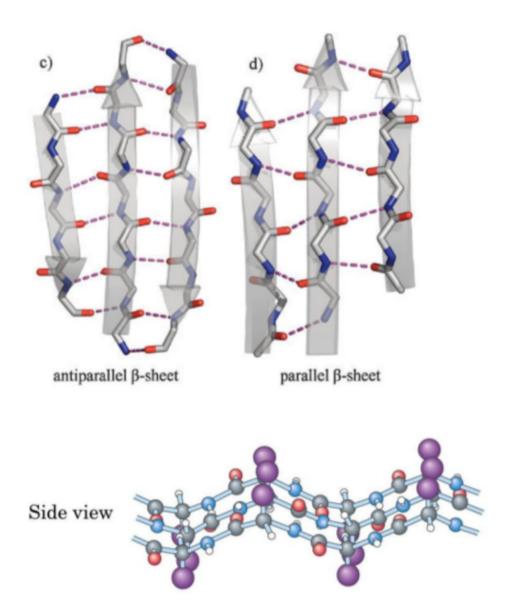
β sheet conformation

Approx. 30% of globular proteins

Strands of 5-10 residues run in parallel

Strands are held together by H-bonds

Different β-sheet forming propensity for various residues

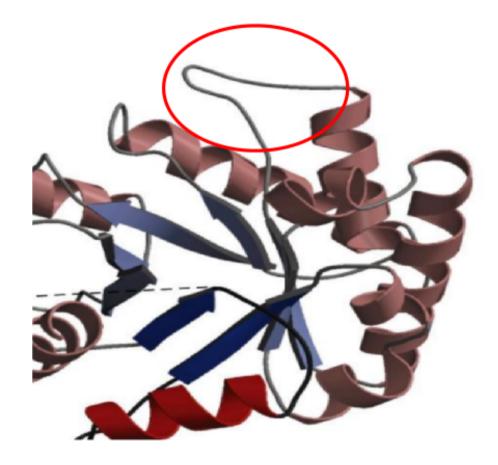


Loops and turns

Typically have hydrophilic characters. Occur on the outer regions of the protein, form H-bonds with water and other molecules

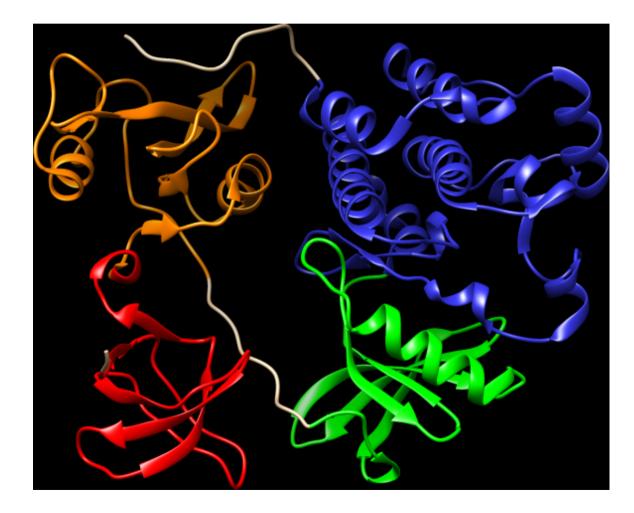
Often form binding regions and active sites in enzymes and receptors

Different loop-forming propensity for various residues



Domains

Many proteins feature distinct compact structural units



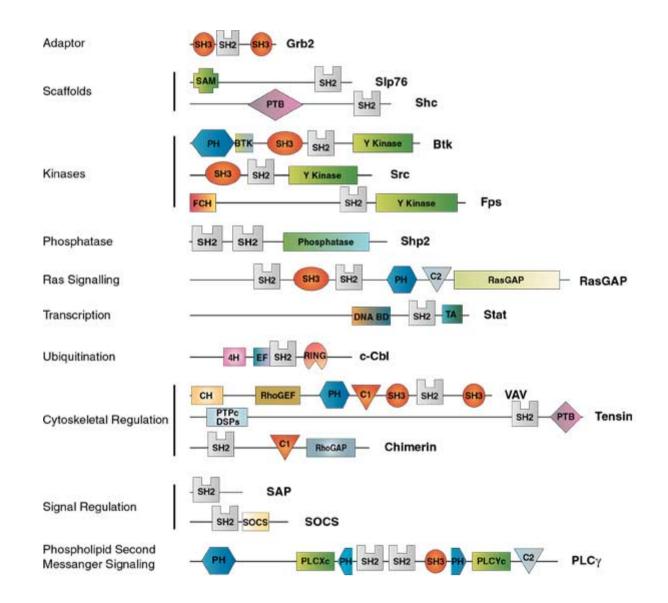
Domains

Compact units with globular-like structures

Domains are basic building blocks of proteins Typically fulfill a well-specified function Can appear in various biological contexts

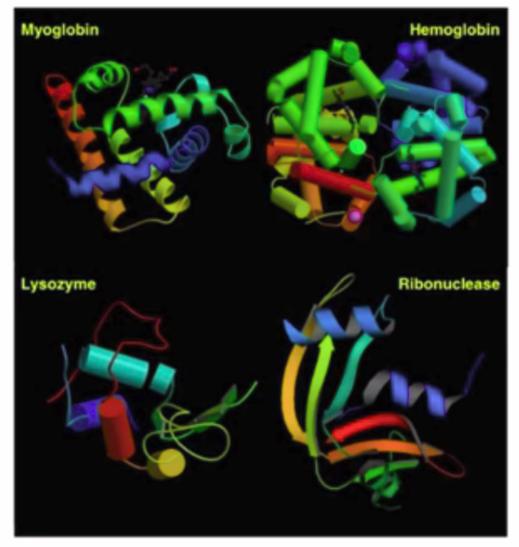


SH2 domain



Protein Data Bank

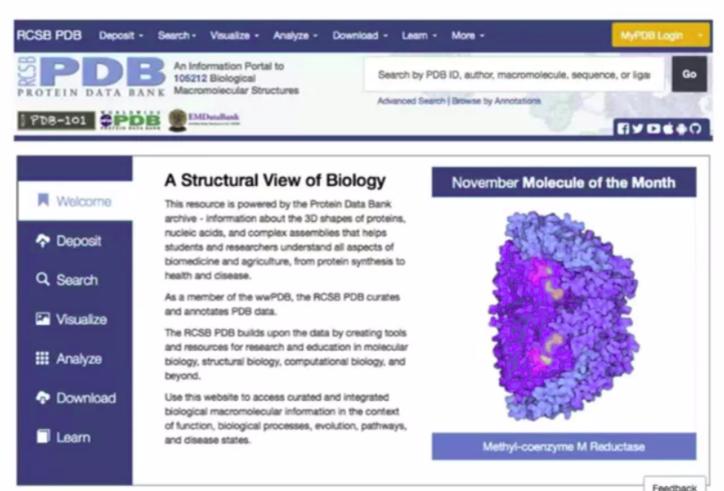
- First open access digital resource in biology (est. 1971 with 7 entries)
- Single global archive of 3-D macromolecular structures (contains >100,000 entries)
- US PDB = RCSB PDB
 - Headquartered at Rutgers/UCSD (NSF, NIH, DOE)
 - Part of Worldwide PDB (with EU and Japan)
- Makes PDB data freely available to all via <u>www.rcsb.org</u>



Some of the first few structures in the PDB

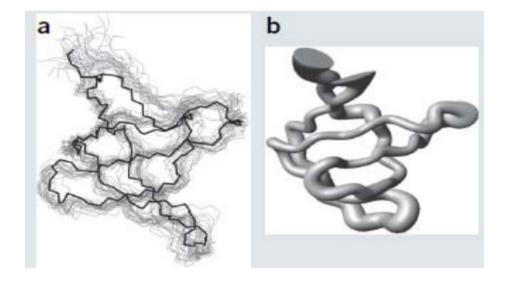
RCSB PDB Portal rcsb.org

- Searching
- Visualizing
- Comparing
- Accessing external data
- Reporting
- PDB-101 resources for education

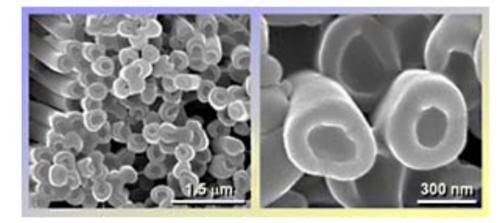


X-ray crystallography





Electron microscopy



X-ray crystallography

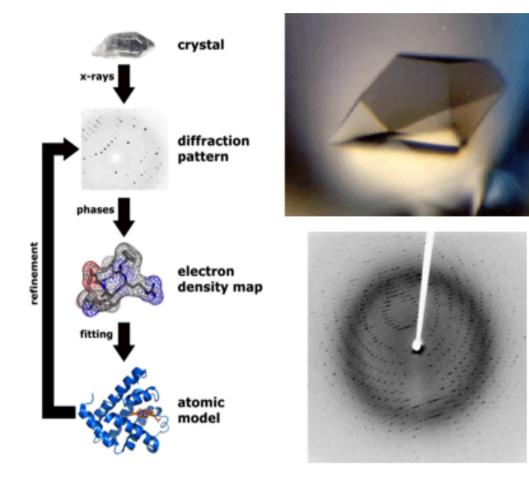


Figure 4: Left: Structure determination by X-ray crystallography. Work by Bragg and others connected spots on diffraction pattern with arrangement of atoms in the crystal to solve simple structures like salts. Work by Perutz, Rossmann, and Blow allowed automated processing of crystal data to solve complex structures. Right (top): typical protein crystal, less than one millimeter in size; Right (bottom): protein crystal diffraction pattern.

X-ray:

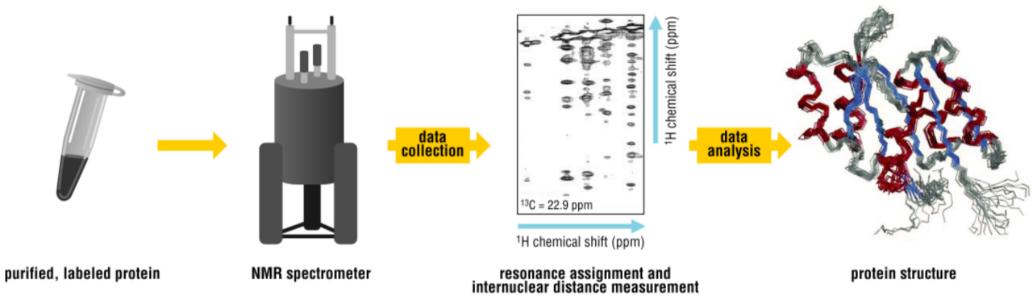
- X-rays have short wave lengths (approx. 1.5 Å) – needed to measure the typical atom-atom distances

- gives information about electron density, the model has to be fit into that

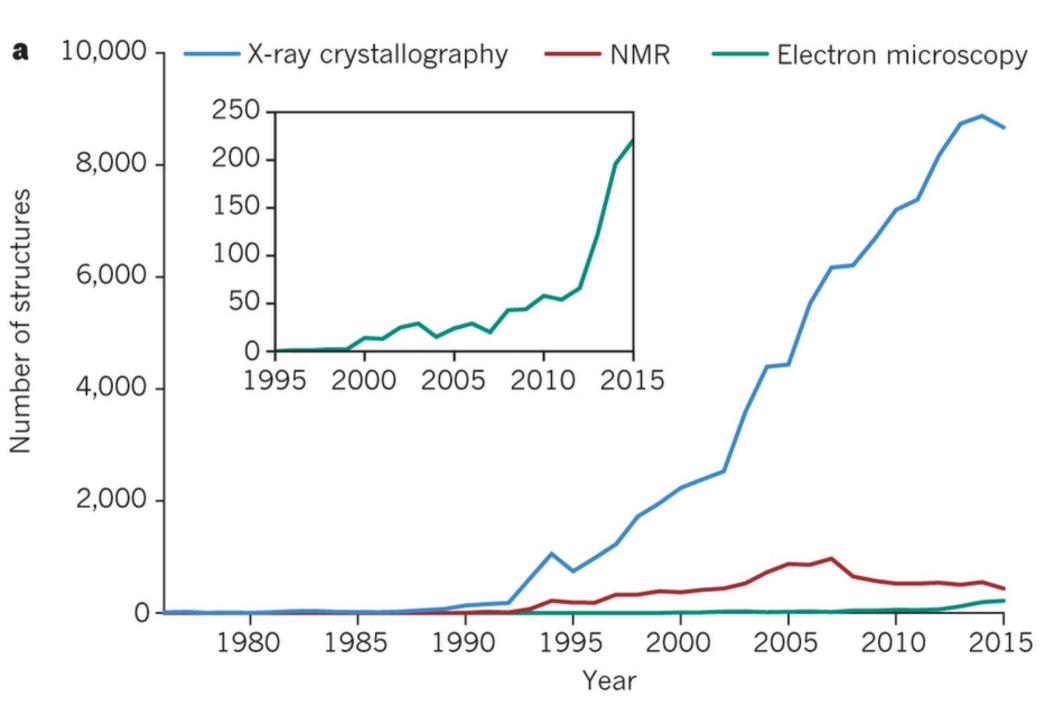
- crystallization artefacts
- non-physiological environment
- no information on hydrogens

NMR

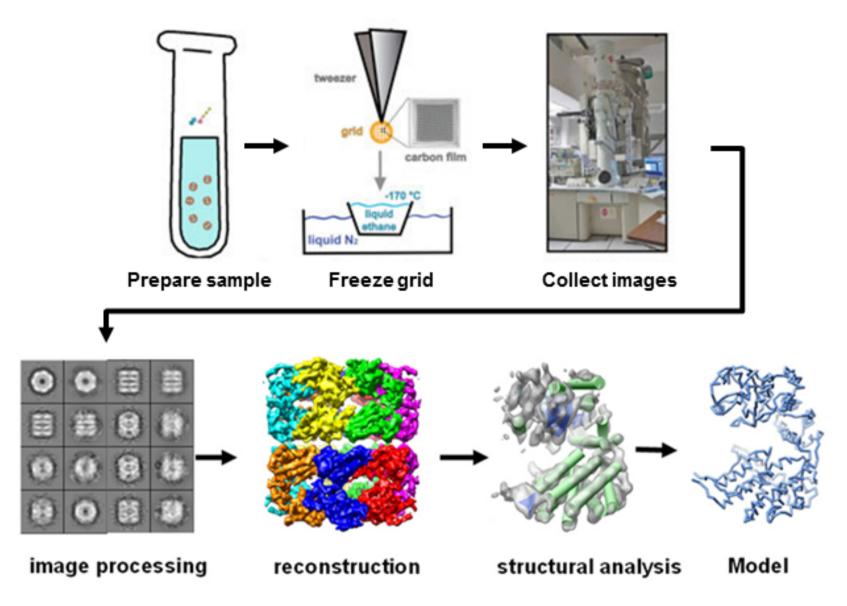
¹H-NMR (Proton <u>Nuclear Magnetic Resonance</u>)



- in solution
- usually yields a structural ensemble that fulfills the distance constraints
- only small proteins
- less precise model
- usable for flexible proteins as well

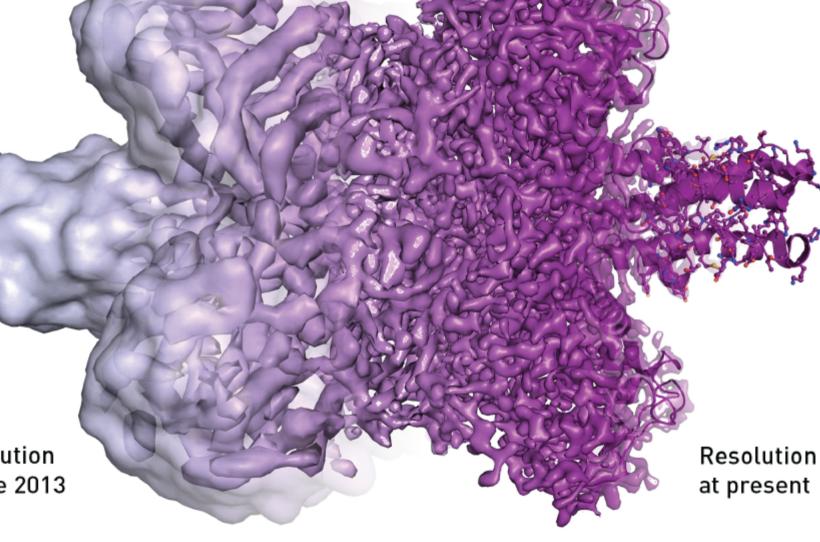


Cryo-EM (atomic resolution)

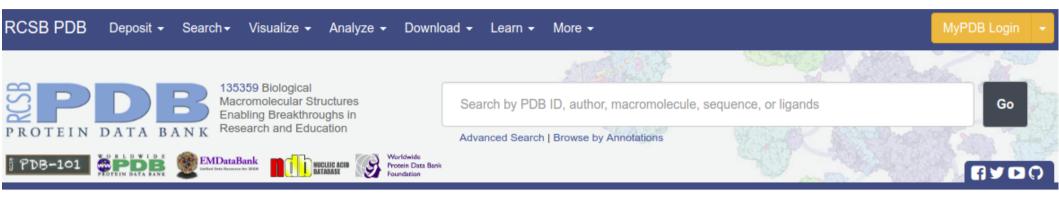


Nobel Prize in Chemistry 2017

Resolution before 2013



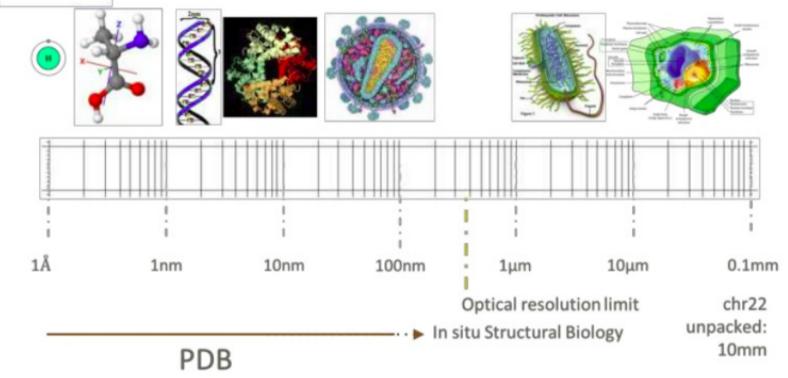
PDB statistics



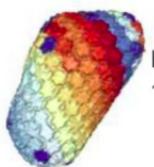
PDB Current Holdings Breakdown

Exp.Method	Proteins	Nucleic Acids	Protein/NA Complexes	Other	Total
X-RAY	113476	1899	5797	4	121176
NMR	10553	1225	246	8	12032
ELECTRON MICROSCOPY	1319	30	468	0	1817
HYBRID	105	3	2	1	111
other	200	4	6	13	223
Total	125653	3161	6519	26	135359

Growing Structure Size and Complexity

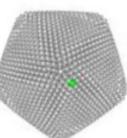


Largest symmetric structure in PDB



HIV-1 capsid: PDB ID 3J3Q ~2.4M unique atoms

Largest asymmetric structure in PDB



Faustovirus major capsid: PDB ID 5J7V ~40M overall atoms

The .pdb file format

The PDB (Protein Data Bank) file format is a text format describing the structure of macromolecules incorporated in the database.

 Description and annotation of the structures of proteins and nucleic acids, including atomic coordinates, side-chain rotamers, secondary structure elements, and atomic connectivity.

 Structures often contain other molecules as well, such as water, ions, ligands, etc. These are also described in the pdb format.

Format description:

http://www.wwpdb.org/documentation/format33/v3.3.html

PDB ID: unique identifier

Each atomic coordinate file in the Protein Data Bank has a unique identifier composed of exactly 4 characters. The first one is always a number, the rest can be either a number or a letter.

There are over 400,000 possible 4-digit PDB IDs (419,904 or 466,560 if "0" can also be the first character). Currently there are approx. 120,000 entries.

Examples:

- 1mbn 1973, the first protein structure model, **myoglobin**
- 1tna 1975, the first RNA structure, yeast phenylalanine transfer RNA
- 1bna 1980, the first **B-DNA** double helix structure (determined using X-ray years after the 1953 theoretically determined structural model of Watson & Crick)
- 2hhd human **hemoglobin**, (deoxy form)
- 9ins insulin

The .pdb file format

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<pre> EXPDTA X-RAY DIFFRACTION AUTHOR R.Z.KRAMER,L.VITAGLIANO,J.BELLA,R.BERISIO,L.MAZZARELLA, AUTHOR 2 B.BRODSKY,A.ZAGARI,H.M.BERMAN</pre>								
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ATOM	2	CA	PRO A		7.608 20.72		1.00 17.44	
ATOM	3	С	PRO A	1	8.487 20.7		1.00 17.44	
ATOM	4	0	PRO A	1	9.466 21.4	57 19.005	1.00 17.44	
ATOM	5	СВ	PRO A	1	6.460 21.72	23 20.211	1.00 22.26	
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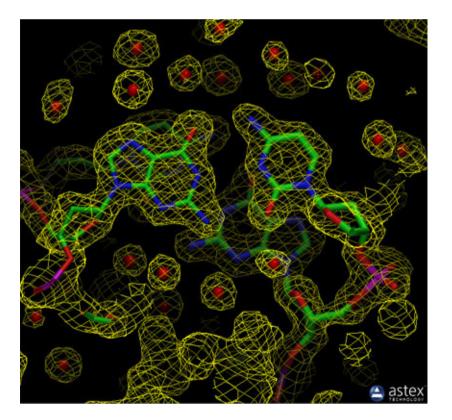
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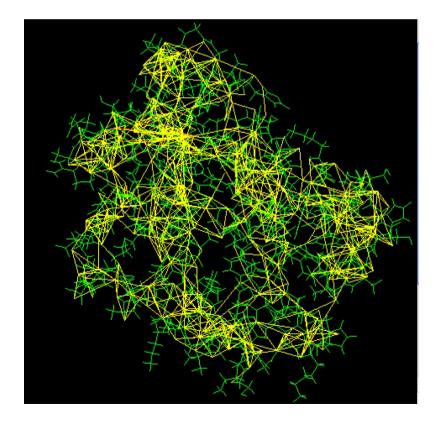
The .pdb file format

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атом atom numbe			ne	A 775	-26.321 3D ID, number	9.177		1.00	67.65 Cy	o t atom type 3-factor

Model

All protein structures are models! Structures are not directly measured, but are generated as models that best fit the collected experimental data.





Resolution (X-ray)

• Describes the reliability of determined atomic coordinates

Very low:>4Å

Individual coordinates cannot be interpreted Low: 3.0-4.0Å

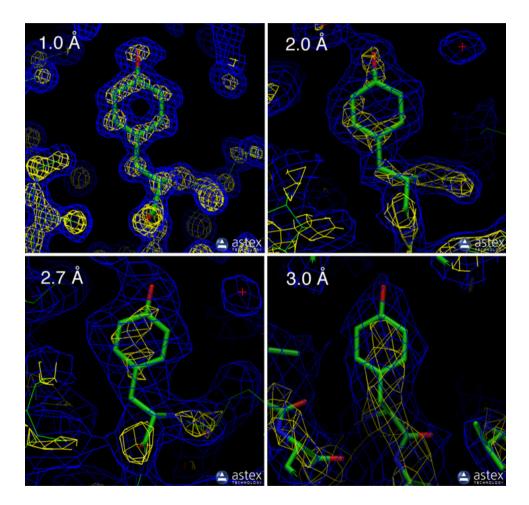
The fold is recognizable

Average: 1.8-3.0Å

The majority of the structure is correct, with incorrect rotamers and unreliable surface loop conformations Good: 1.0 – 1.8Å

Atomic level: <1.0A

Resolution can change for each position!



Describing structure quality

Expected distribution:

Based on small molecules

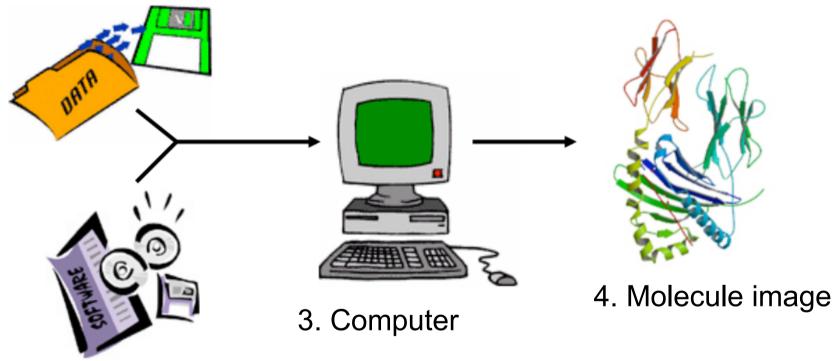
Based on known, high quality structures

Possible parameters

- Correct bond lengths and bond angles
- No atom-atom clashes
- Most buried amid groups form H-bonds
- Based on main chain conformational properties

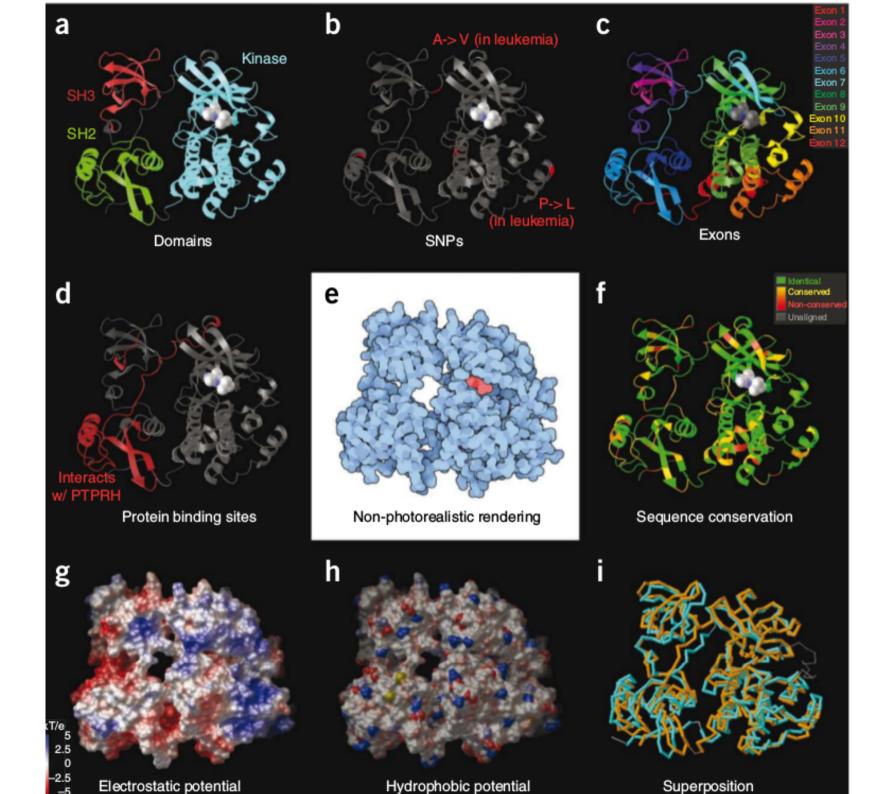
Visualizing protein structures

1. PDB coordinate file



2. Visualization program

Eg.: Rasmol, Pymol, Chimera, VMD, Jmol, Swiss PDB viewer



Secondary structures are stabilized by H-bonds



Secondary structure determination

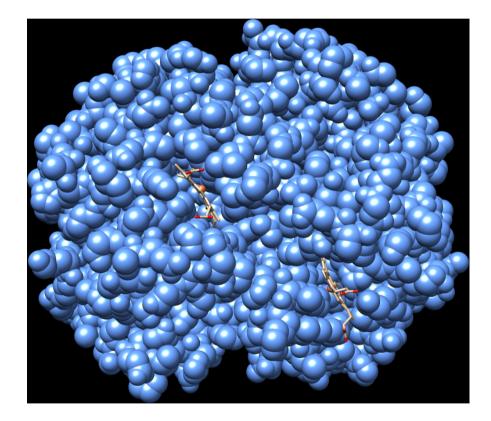
Can be based on: H-bond patterns Dihedral angles

Automatic determination using algorithms DSSP STRIDE

3 (alpha, beta, coil) or more categories (e.g. turn, other helix types)

Do not agree 100%

The inside of the protein is tightly packed

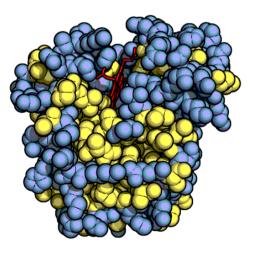


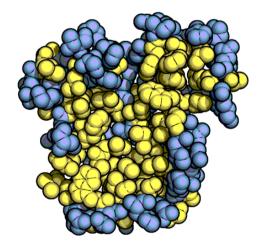
Hydrophobic core

Hydrophobic side chains go into the core of the molecule – but the main chain is highly polar.

The polar groups (C=O and NH) are neutralized through formation of H-bonds.

Myoglobin

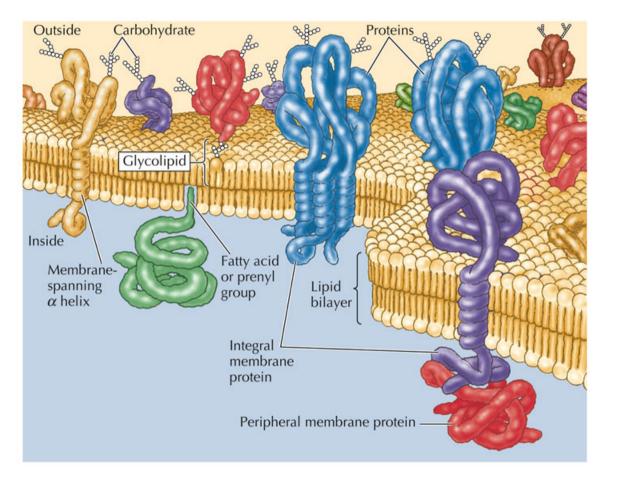




surface

buried

Membrane proteins



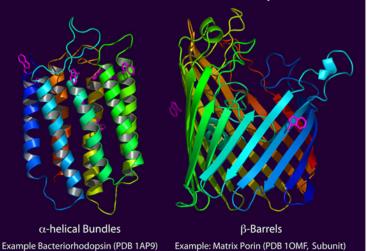
THE CELL, Fourth Edition, Figure 2.25 © 2006 ASM Press and Sinauer Associates, Inc.

Important for:

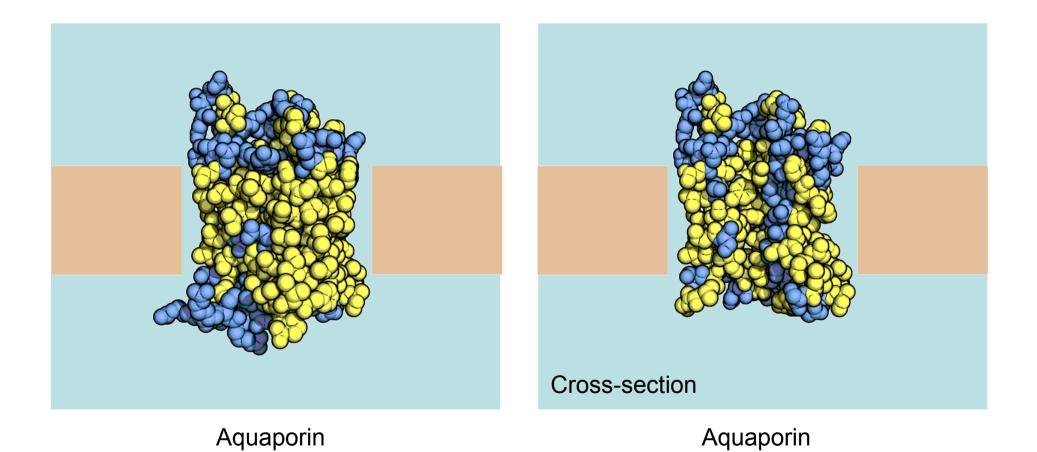
Energy production Transport Cell-cell junction Signaling

Drug targets

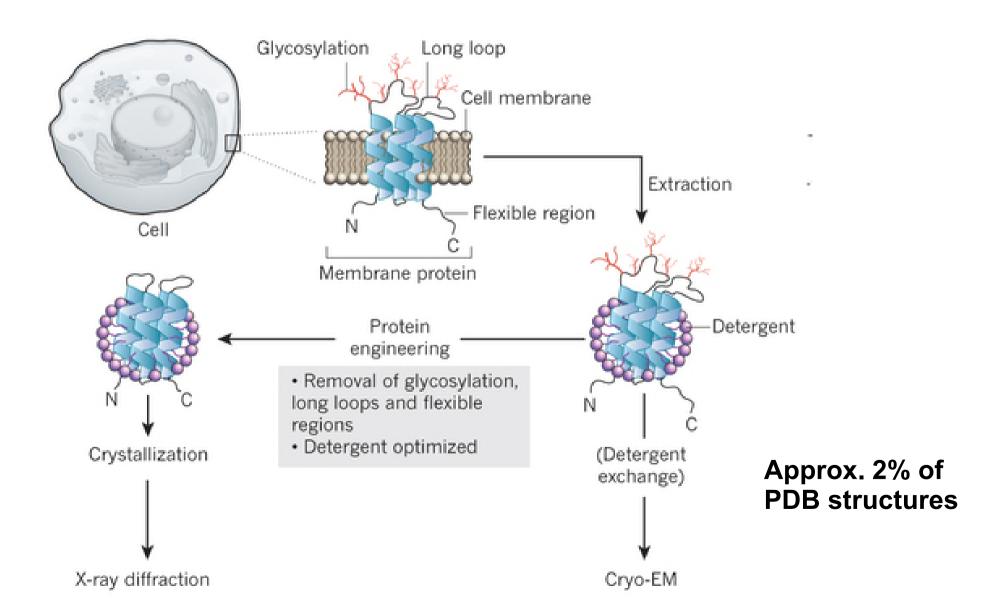
The known structures of transmembrane proteins belong to two classes, based on their transmembrane secondary structure.



Hydrophobicity of membrane proteins



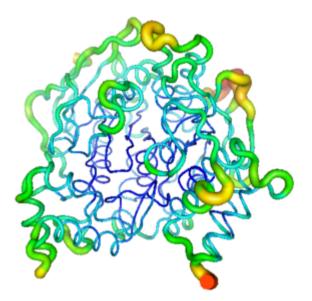
Structure determination of transmembrane proteins

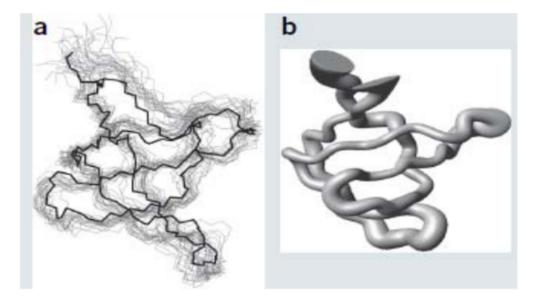


Proteins are dynamic molecules

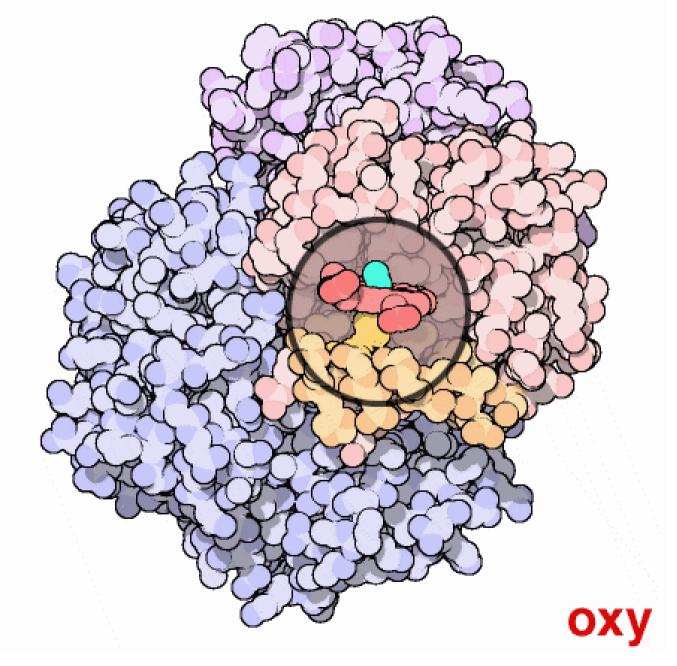
X-ray B-factor



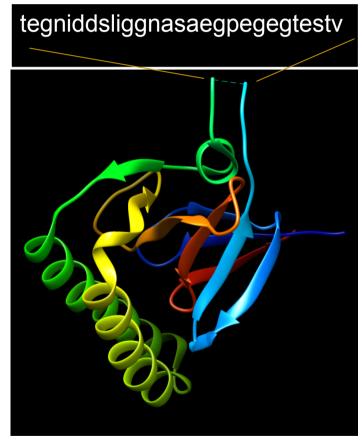




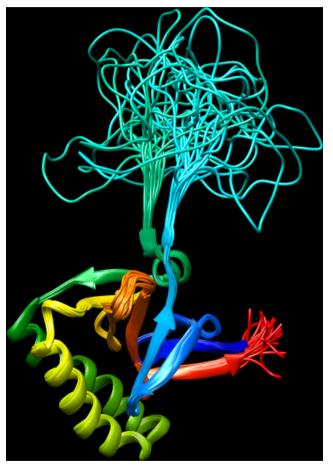
Conformational changes



Missing structure parts



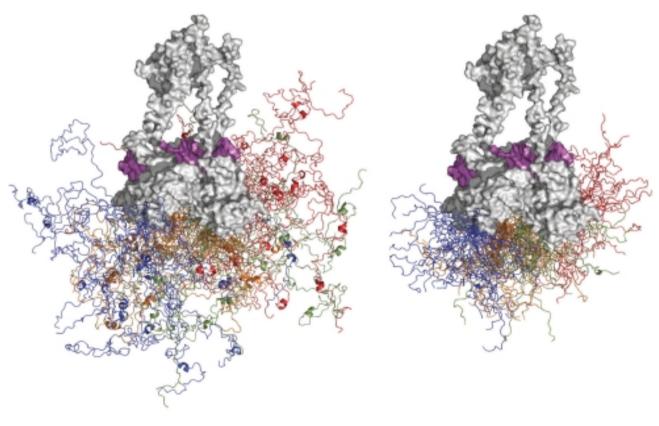
Missing regions in the protein structure



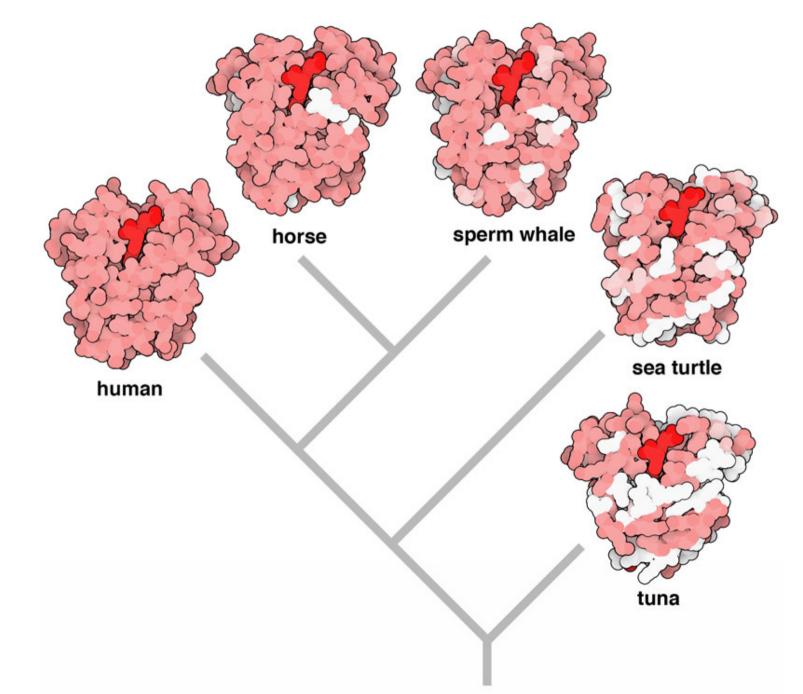
NMR structures with high structural variability

Intrinsically disordered proteins

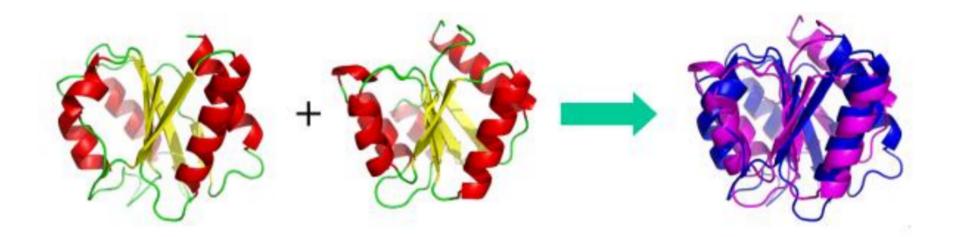
Do not form a well-defined structure on their own under native(-like) conditions



Globin evolution

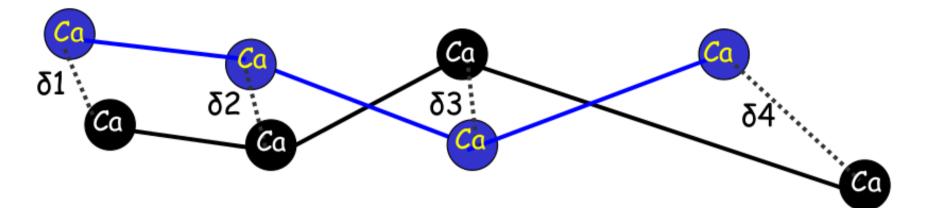


Similarity between two structures



Superposition: minimizing distances between positions

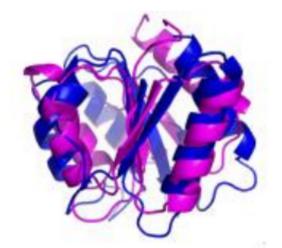
RMSD



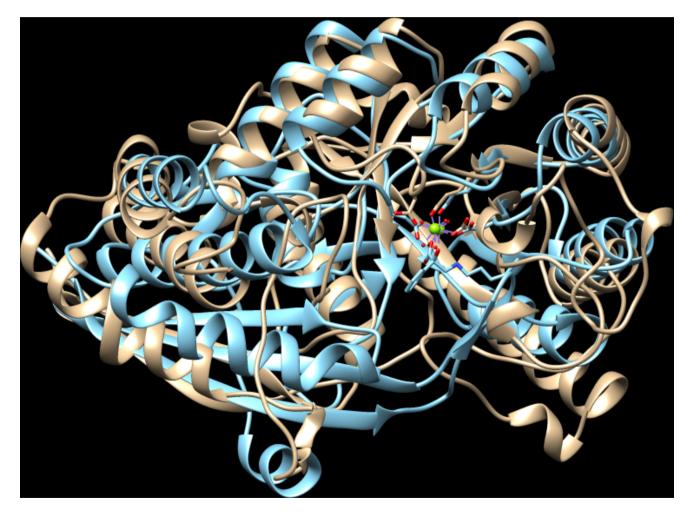
Root Mean Square Deviation (RMSD):

The most commonly used function for measuring structural similarity

RMSD is the average distance between equivalent atoms of superimposed structures $RMSD = \sqrt{\frac{1}{N}\sum_{i=1}^{i=N}\delta_i^2}$



Structures of evolutionarily related proteins are usually similar



1ebhA: enolase 1mns : mandelate racemase

Sequence identity: 25% Active center is very similar Simlar chemical reactions Different substrate

Sequence-structure relationship

The structure is usually more conserved than the sequence

Structures typically tolerate more mutations

Due to physical effects some structures are more common Analogue

The number of folds is limited Currently around 1,200 folds

Structural classification

We can group similar and evolutionarily related protein structures using classification

Example

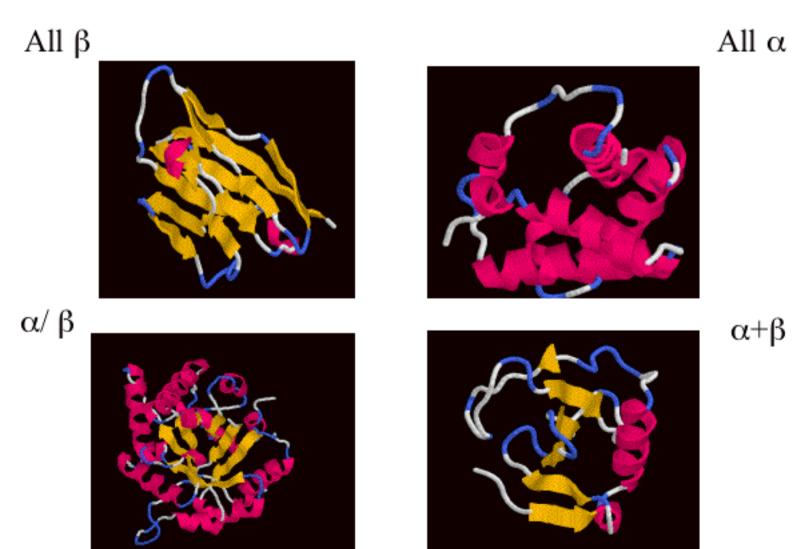
CATH

http://www.cathdb.info/

SCOP

http://scop2.mrc-lmb.cam.ac.uk/

Structural classes

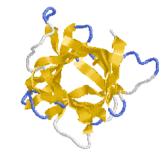


Fold - topology

Proteins belonging to the same fold contain roughly the same secondary structure elements in the same order and similar spatial configuration.



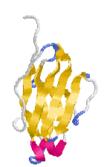
globin





trefoil

up-down





 α/β sandwich

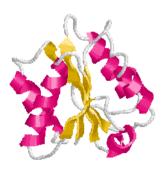


jelly roll



TIM barrell

immunoglobulin







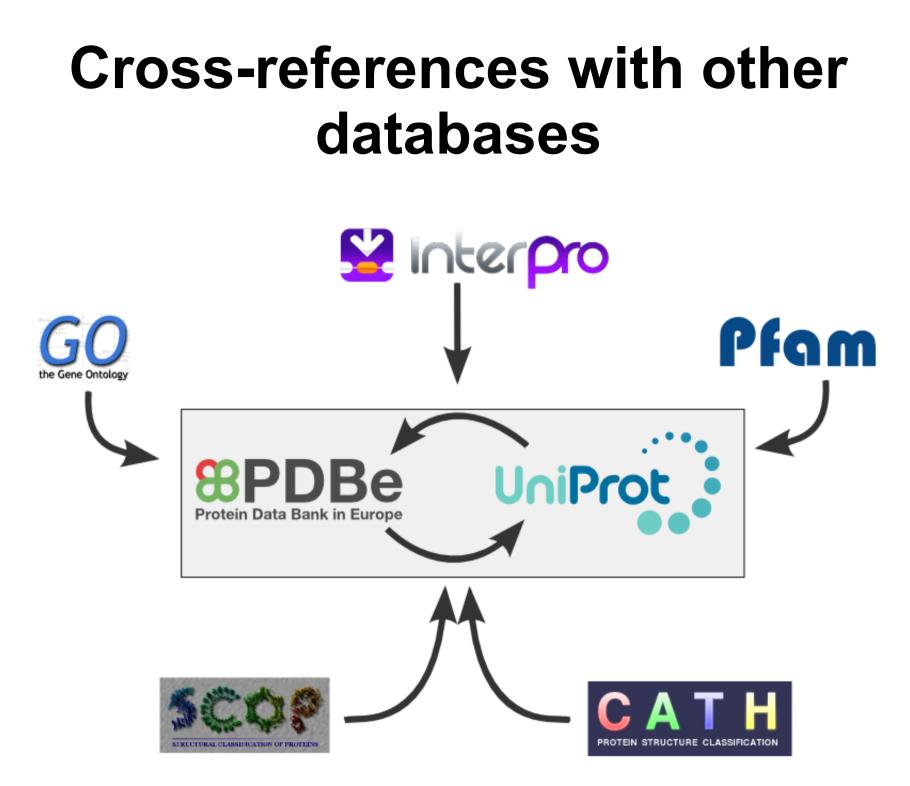
UB α/β roll

Homolous and analogous structures

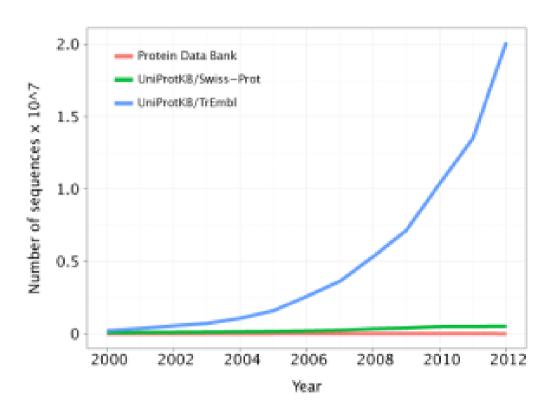
Homolous proteins evolved from a common ancestor via divergence, and share the same fold

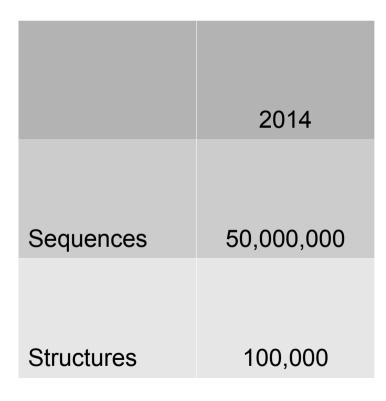
Analogous proteins share the same fold but do not have and evolutionary relationship (or it is undetectable)

Some folds are more common (due to physical effects) Number of folds is limited (1-2,000 folds)



Sequence-structure gap

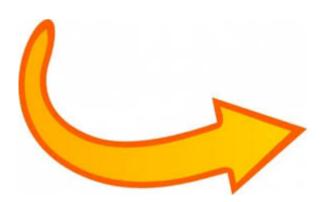


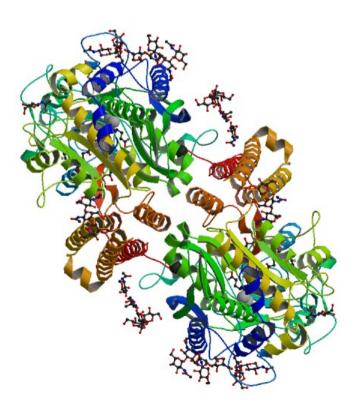


Tertiary structure predictions

>Protein

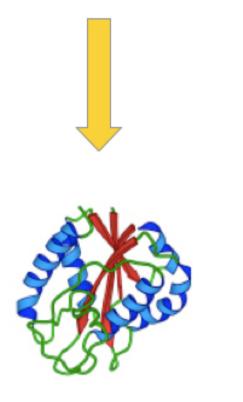
RSKSSNEATNITPKHNMKAFLDELKAENIKKFLYNFTQIPHLAGTEQNFQLAKQIQSQWKEFGLDSVELAHYDVLLSYPN KTHPNYISIINEDGNEIFNTSLFEPPPPGYENVSDIVPPFSAFSPQGMPEGDLVYVNYARTEDFFKLERDMKINCSGKIV IARYGKVFRGNKVKNAQLAGAKGVILYSDPADYFAPGVKSYPDGWNLPGGGVQRGNILNLNGAGDPLTPGYPANEYAYRR GIAEAVGLPSIPVHPIGYYDAQKLLEKMGGSAPPDSSWRGSLKVPYNVGPGFTGNFSTQKVKMHIHSTNEVTRIYNVIGT LRGAVEPDRYVILGGHRDSWVFGGIDPQSGAAVVHEIVRSFGTLKKEGWRPRRTILFASWDAEEFGLLGSTEWAEENSRL LQERGVAYINADSSIEGNYTLRVDCTPLMYSLVHNLTKELKSPDEGFEGKSLYESWTKKSPSPEFSGMPRISKLGSGNDF EVFFQRLGIASGRARYTKNWETNKFSGYPLYHSVYETYELVEKFYDPMFKYHLTVAQVRGGMVFELANSIVLPFDCRDYA VVLRKYADKIYSISMKHPQEMKTYSVSFDSLFSAVKNFTEIASKFSERLQDFDKSNPIVLRMMNDQLMFLERAFIDPLGL PDRPFYRHVIYAPSSHNKYAGESFPGIYDALFDIESKVDPSKAWGEVKRQIYVAAFTVQAAAETLSEVA



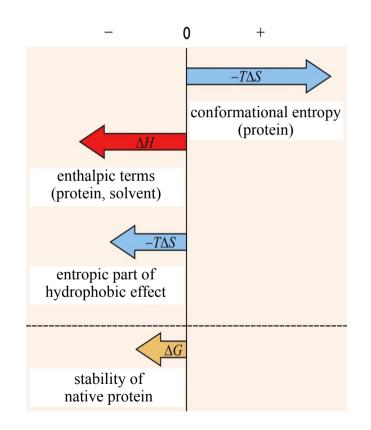


Protein folding

GFCHIKAYTRLIMVG...



 $\Delta G = \Delta H - T \Delta S$



Folding

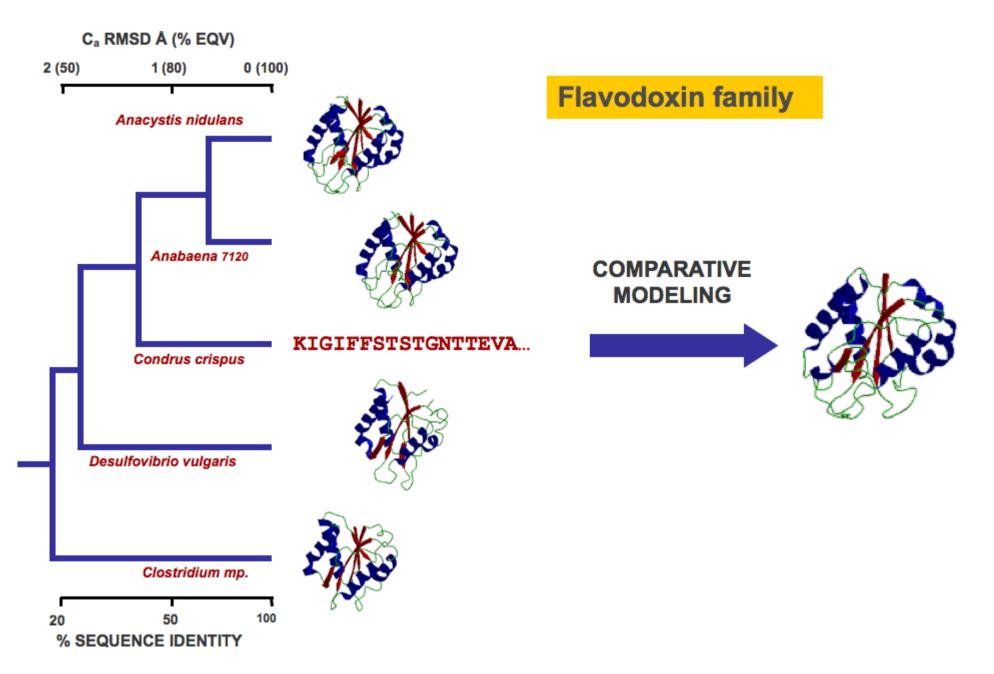
(physics)

Determining tertiary structure based on physical principles

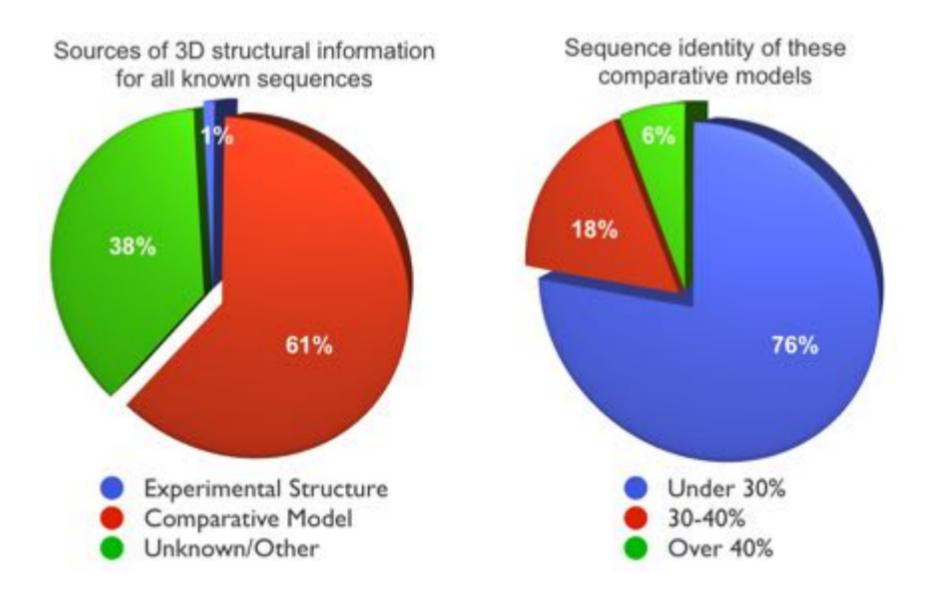
- large number of comformations, huge conformational space

- the physical energy function is not known exactly

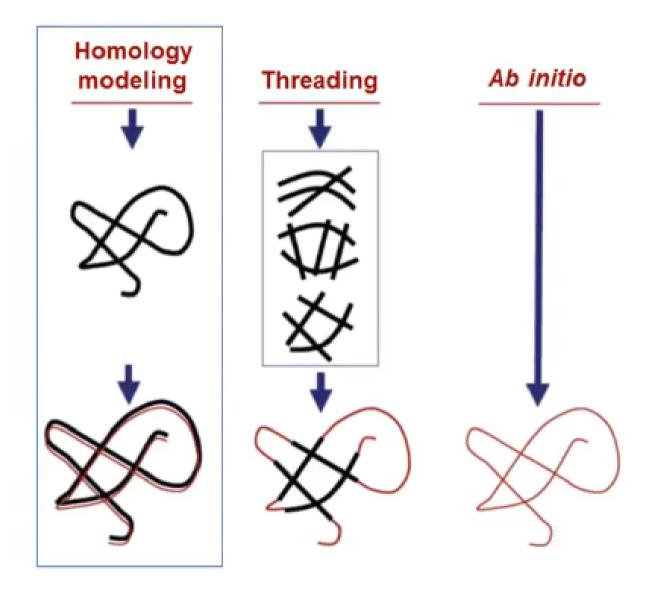
Comparative structure modeling



Structural coverage



Tertiary structure prediction approaches





The Nobel Prize in Chemistry 2013 Martin Karplus, Michael Levitt, Arieh Warshel

The Nobel Prize in Chemistry 2013



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Photo: © S. Fisch Michael Levitt

Photo: Wikimedia

Arieh Warshel

M.L.:

It's sort of nice in more general terms to see that computational science, computational biology is being recognized." He added, "It's become a very large field and it's always in some ways been the poor sister, or the ugly sister, to experimental biology."

The Nobel Prize in Chemistry 2013 was awarded jointly to Martin Karplus, Michael Levitt and Arieh Warshel "for the development of multiscale models for complex chemical systems".